DISSERTATIONES MEDICINAE UNIVERSITATIS TARTUENSIS 163

DISSERTATIONES MEDICINAE UNIVERSITATIS TARTUENSIS 163

TRIIN ELLER

Immune markers in major depression and in antidepressive treatment



Department of Psychiatry, University of Tartu, Tartu, Estonia

Dissertation is accepted for the commencement of the degree of *doctor medicinae* on October 13, 2009 by the Council of the Commencement of Doctoral degree in Medicine, University of Tartu, Estonia.

Supervisors:	Veiko Vasar, MD, PhD, Professor, Department of Psychiatry, University of Tartu
	Jakov Šlik, MD, PhD, FRCPC, Associate Professor, Department of Psychiatry, University of Ottawa, Ottawa, Ontario, Canada
	Eduard Maron, MD, PhD, Senior Researcher Fellow, Department of Psychiatry, University of Tartu
Reviewers:	Vallo Volke, MD, PhD, Senior Researcher, Institute of Physiology, University of Tartu
	Külli Kingo, MD, PhD, Associate Professor, Senior Researcher, Chair of Dermatology and Venerology, University of Tartu
Opponent [.]	Raimo K R Salokangas MD PhD Professor Department of

Opponent: Raimo K.R. Salokangas MD, PhD, Professor, Department of Psychiatry, University of Turku, Turku, Finland

Commencement: January 15, 2010

Publication of this dissertation is granted by the University of Tartu

ISSN 1024–395x ISBN 978–9949–19–260–1 (trükis) ISBN 978–9949–19–261–8 (PDF)

Autoriõigus Triin Eller, 2009

Tartu Ülikooli Kirjastus www.tyk.ee Tellimus nr 450

CONT	ENTS
------	------

CONTENTS	5
LIST OF ORIGINAL PUBLICATIONS	7
ABBREVIATIONS	8
INTRODUCTION	10
REVIEW OF LITERATURE	12
1. Major depressive disorder (MDD)	12
1.1. Epidemiology of MDD	13
1.2. Aetiology and pathogenesis of MDD	14
1.2.1. Monoamine neurotransmitters	14
1.2.2. Hypothalamic-Pituitary-Adrenal axis (HPA)	14
1.2.3. Brain structure in depression	14
1.3. Genetics of MDD.	16
	10
2. Immune system cells and cytokines	
2.1. Classification of cytokines	18
2.2. Cytokines and depression	18
3. Thyroid function in major depression	28
3.1. Anti thyroid peroxidise auto-antibodies (anti-TPO)	28
AIMS OF THE STUDY	29
MATERIALS AND METHODS	30
1. Ethical considerations	30
2. Characteristics of study participants and study design	30
3. Laboratory analyses	31
4. Statistical analysis	32
RESULTS	33
1. Differences in cytokine levels between MDD patients and	
healthy controls	33
2. Escitalopram treatment effects on IL-8, TNF- α , and sIL-2R levels	
in MDD patients	34
3. Bupropion augmentation effects on IL-8, TNF- α , and sIL-2R	
levels in escitalopram-resistant MDD patients	36
4. Associations between IL-8, TNF- α , and sIL-2R baseline serum	
concentrations and treatment response in MDD patients	37
5. Thyroid function and treatment response	38
f	

DISCUSSION	39
1. TNF-α	39
2. TNF- α in the escital optimate the phase	40
3. TNF- α in the augmentation phase with bupropion	41
4. IL-8	41
5. IL-8 in the escitalopram – treatment phase	42
6. IL-8 in the augmentation phase with bupropion	42
7. sIL-2R	43
8. sIL-2R in the escitalopram- treatment phase	45
9. sIL-2R in the augmentation phase with bupropion	46
10. Anti-TPO and thyroid hormones	46
11. General discussion and future perspectives	47
CONCLUSIONS	50
REFERENCES	51
SUMMARY IN ESTONIAN	64
ACKNOWLEDGEMENTS	69
PUBLICATONS	71
CURRICULUM VITAE	116

LIST OF ORIGINAL PUBLICATIONS

- I. Eller T, Aluoja A, Maron E, Vasar V. Soluble interleukin 2 receptor and tumour necrosis factor in depressed patients in Estonia. Medicina, 2009 (accepted).
- II. Eller T, Vasar V, Shlik J, Maron E. 2008. Pro-inflammatory cytokines and treatment response to escitalopram in major depressive disorder. Prog Neuropsychopharmacol Biol Psychiatry. 32: 445–450.
- III. Eller T, Vasar V, Shlik J, Maron E. 2008. Effects of bupropion augmentation on pro – inflammatory cytokines in escitalopram-resistant patients with major depressive disorder. J Psychopharmacol 2009 Sep 23(7): 854–858.
- IV. Eller T, Metsküla K, Talja I, Maron E, Uibo R, Vasar V. Thyroid autoimmunity and treatment response to escitalopram in major depression (submitted to Nord J Psychiatry, 2008).

Contribution of the author:

- 1. The author took care of the patient selection and treatment, performed the clinical scales, carried out statistical analysis, and wrote the first draft of the manuscript.
- 2. The author took care for the patient treatment, performed the clinical scales, carried out statistical analysis, and wrote the first draft of the manuscript.
- 3. The author designed the study, performed the clinical scales, carried out statistical analysis, and wrote the first draft of the manuscript.
- 4. The author participated in the study design, performed the clinical scales, carried out statistical analysis, and wrote the first draft of the manuscript.

ABBREVIATIONS

АСТН	_	Adrenocorticotropic hormone
anti-TPO	_	Anti thyroid peroxidise auto-antibodies
AVP	_	Arginine vasopressin
BDI	_	Beck Depression Inventory
BMI	_	Body mass index
CSF	_	Cerebrospinal fluid
CRH	_	Corticotropin releasing hormone
DA	_	Dopamine
DSM-IV	_	Diagnostic and Statistical Manual of Mental Disorders,
		4th edition
FE	_	MDD, first episode
FR	_	MDD, full remission
HAMD	_	Hamilton Depression Rating Scale
НС	_	Healthy controls
HPA axis	_	Hypothalamic-Pituitary-Adrenal axis
ICPE	_	International Consortium of Psychiatric Epidemiology
IDO	_	Indoleamine 2,3-dioxygenase
IFN-α	_	Interferon-a
IFN-γ	_	Interferon-y
IL .	_	Interleukin
IL-1	_	Interleukin-1
IL-1RA	_	Interleukin-1 receptor antagonist
IL-6	_	Interleukin-6
IL-8	_	Interleukin-8
IL-12	_	Interleukin-12
IRS	_	Inflammatory response system
LPS	_	Bacterial cell wall lipopolysaccharide
MADRS	_	Montgomery-Asberg's Depression Rating Scale
MDD	_	Major depressive disorder
M.I.N.I. 5.0.0	-	Mini International Neuropsychiatric Interview Version
		5.0.0
NA	-	Noradrenalin
NIMH	-	National Institute of Mental Health
NK	-	Natural killer
NR	-	Non-responder
NS	-	Non significant
PGE2	-	Prostaglandin E2
R	-	Responder
RE	-	MDD, recurrent depressive episode
S	-	Stimulated
SD	-	Standard deviation
sIL-2R	-	Soluble interleukin-2 receptor

sIL-6R	_	Soluble interleukin-6 receptor
SSRI	_	Selective Serotonin re-uptake inhibitor
TCA	_	Tricyclic antidepressant
TDO	_	Tryptophan 2,3-dioxygenase
Tc cells	_	Cytotoxic T cells
Th cells	_	T-helper cells
TNF-α	_	Tumour necrosis factor α
US	_	Unstimulated
5-HT	_	Serotonin (5-hydroxytryptamine)

INTRODUCTION

Unipolar depression is a common, often chronic and episodic psychiatric disorder (Andrade et al., 2003; Weissman et al., 1996). Depression is a major health problem worldwide for two reasons: it is highly prevalent in the general population, and causes a significant loss of quality of life and social functioning in the affected individual. The prevalence rates vary widely across studies and different countries; for example, the one-month prevalence of depression is found from 2.2–20.7 % (Angst and Merikangas, 1997; Angst et al., 2002; Ialongo et al., 2004; Kessler et al., 1993; Regier, 1993; Regier et al., 1988). Prevalence rates have been found to be higher in women than in men. Further, depression contributes to a poorer outcome of co-morbid mental and somatic conditions.

The fact that mood disorders, including unipolar depression, have a great impact on distress of the affected individual and his or her family, lifetime disability, and suicide highlights the importance of etiologic research for their treatment and prevention (Merikangas et al., 2002). The available data consistently demonstrate an association of all mental disorders with considerable disability burden in terms of the number of work days lost and generally low treatment rates. Only 36.5% of cases with mood disorders had had any consultation with professional health care services; the finding suggests a considerable degree of unmet need (Wittchen and Jacobi 2005). In addition, there is evidence suggesting that biological mechanisms underlie a bidirectional link between mood disorders and many medical illnesses. Moreover, mood disorders may affect the course of medical illnesses (Evans et al., 2005).

Major depression is a complex disorder caused by genetic and environmental factors and interactions between them. There are many factors associated with depression: age, gender, living with a partner, ethnic background, education, immigrant status, urban-rural differences, life stress, childhood traumas, comorbidity with other mental and physical disorders (Aluoja et al., 2004; Paykel et al., 2005; Smit et al., 2004). Despite some promising leads, no confirmed linkage in mood disorders has been established as yet. Impediments to gene findings include the lack of phenotypic validity, variation in ascertainment sources and methodology across studies, and genetic complexity (Merikangas et al., 2002).

A large body of evidence in recent years suggests that major depression is associated with activation of the inflammatory response system (IRS) (Connor and Leonard, 1998; Maes et al., 1995; Schiepers et al., 2005). Cytokines are small glycoproteines that function as signalling molecules between immune cells. When examining systemic activity of the immune system in depressed patients, differentiation into two major groups is justified, namely, proinflammatory and anti-inflammatory cytokines. An increased production of proinflammatory cytokines may play a crucial role in the immune and acute phase responses in depression (Van West 2005). There is evidence implying that antidepressive treatment with various antidepressive agents has an immunomodulative effect. However, the existing data are conflicting.

The purposes of this study were (i) to find associations between depression, depressive symptoms and soluble interleukine- 2 receptor (sIL-2R) and tumour necrosis factor- α . (TNF α) and anti thyroid peroxidise auto-antibodies (anti-TPO); (ii) to investigate the acute and chronic effects of selective serotonin re-uptake inhibitor, escitalopram, on serum levels of interleukin-8 (IL-8), sIL-2R and TNF α in patients with major depression; (iii) to clarify whether the addition of bupropion in escitalopram-resistant patients with major depression causes additional changes in the immune system; (iiii) to explore whether serum cytokine concentrations and/or anti-TPO positivity can predict treatment response to antidepressants.

REVIEW OF LITERATURE

I. Major depressive disorder (MDD)

Mood is defined by DSM-IV as a "pervasive and sustained emotion that colours the perception of the world. In contrast with affect, which refers to more fluctuating changes in emotional 'weather', mood refers to a more pervasive and sustained emotional *climate*" (DSM-IV). Mood is obviously modified by the events that occur in the real world. Negative events lower our mood and positive events tend to make us happier. On the one hand, mood affects all of our cognitions, judgements, and expectations. Through these continuous variations mood is influenced by the ever-changing flow of life events; mood determines our attitude towards life, and the basic characteristic of normal mood is that it is subject to change. Pathological mood is no longer influenced by changes in reality and remains steady and still, regardless of any occurrence. On the other hand, the state of mood continues to determine the cognition and the interpretation of reality (Faravelli et al., 2005). The lack of response to external stimuli is therefore the basis of the pathology of mood, rather than the intensity of the mood.

Four major clusters of symptoms and signs are recognized in the diagnosis of depression. They include mood (anhedonia, dysphoria, guilt, anxiety), cognition (attention deficit, difficulty to make decisions, lack of interest, thought content deals with sadness, hopelessness, lack of future, low selfesteem, death from boredom with life to actual self-harm), psychomotor (retardation, agitation), and neuro-vegetative (disturbed sleep, appetite, and sexuality) areas. The European psychopathological tradition indicates melancholy as the psychopathological nucleus of depression. It includes somatic symptoms and signs (worsening in the morning, early morning awakening, significant anorexia or weight loss), marked psychomotor retardation or agitation, guilty feelings and specific features of the mood (loss of interest or pleasure in all or almost all activities, lack of reactivity to usually pleasurable stimuli, peculiar characteristic of the depressed mood, which is qualitatively different from normal sadness). Atypical symptoms are opposites of melancholic ones. They include mood reactivity (i.e. mood brightens in response to actual or potential positive events) and inverse neuro-vegetative symptoms (increased appetite or significant weight gain, hypersomnia). Psychotic features considered by DSM-IV are delusions and hallucinations that might be mood-congruent or mood-incongruent.

I.I. Epidemiology of MDD

The one-month prevalence of depression is found between 2.2 and 20.7 % (Angst and Merikangas 1997; Angst et al., 2002; Ialongo et al., 2004; Kessler et al., 1993; Regier 1993; Regier et al., 1988). Life-time prevalence varies between 3% in Japan and 16.9% in the US according to the surveys of the International Consortium of Psychiatric Epidemiology (ICPE) (Andrade et al., 2003) and amounts to 16.2% in the National Co-morbidity Survey Replication (NCS-R) (Kessler et al., 2003). The high variation is due to different assessment instruments (e.g. semi-structured interviews or standardized interviews) and classification systems. Very low rates of major depression have been reported in studies conducted in Eastern Asian nations. Sociodemographic differences (e.g. discrepancies in the distribution of marital status) or cross-cultural variations (e.g. different social acceptability of the expression of emotions) could explain the discrepancies between the results. Prevalence rates have been consistently found to be 1.5–2.5 times higher in women than in men (Jacobi et al., 2005).

Depression is highly co-morbid with other mental disorders, especially anxiety. In most surveys, between one-third and half of respondents with a lifetime history of major depressive episode also had a history of at least one anxiety disorder (Andrade et al., 2003; Rush et al., 2005). Major depression and dysthymia frequently coexist; the disorder is sometimes referred to as 'double depression'. The lifetime prevalence of double depression has been reported to range between 1.5% and 2.5% (Bland 1997). The median prevalence of current or lifetime alcohol problems in depression is 16% and 30%, respectively (Sullivan et al., 2005).

The median age of the onset of MDD is in the range 20 to 25 in most countries by the ICPE Survey. Consistent socio-demographic correlates included being female and unmarried (Andrade et al., 2003).

Family studies have shown that the risk of depression onset and severity is associated with family history of depression (Rohde et al., 2005). Sullivan et al. reported the relative risk for MDD subjects versus first-degree relatives of MDD patients 2.84 (Sullivan et al., 2000).

Suicide is one of the most serious aspects of major depression although there is evidence that suicide risk loci are independent of susceptibility loci of mood disorders (Zubenko et al., 2004). Suicide has been reported to occur in 10-15 % of patients previously hospitalized for major depressive disorder (Angst et al., 1999). In a study conducted by Murphy (1998), women were more likely to experience episodes of major depression but were 25% less likely than men to commit suicide.

I.2. Aetiology and pathogenesis of MDD

There is general agreement that the clinical syndrome of major depression must be associated with characteristic neurobiological changes in the brain. However, it is unclear to what extent specific syndromes correlate with particular neurobiological changes. In addition, there is some evidence that depression is associated with a distinct cellular and structural pathology.

1.2.1. Monoamine neurotransmitters

The monoamine hypothesis suggests that depressive disorder is due to an abnormality in the monoamine neurotransmitter system at one or more sites in the brain (Cowen 2005). Alterations were found both in receptors and in the concentrations or the turnover of the amines. Three monoamine transmitters have been implicated: serotonin (5-hydroxytryptamine; 5-HT), noradrenalin (NA), and dopamine (DA).

The synthesis of 5-HT in the brain depends on the availability of L-tryptophan. Plasma tryptophan levels are decreased in depressed patients, particularly in those with melancholic depression (Anderson et al., 1990). Recent development in brain imaging with selectively labelled ligands has allowed assessment of certain brain 5-HT receptor subtypes in vivo. There is evidence that unmedicated depressed patients differ from healthy volunteers in the density of 5-HT2a, 5-HT1a receptors in certain brain regions, and there are probably reductions in brain stem 5-HT transporter sites in depressed subjects, consistent with a decrease in the density of 5-HT cell bodies (Cowen 2005; Willeit et al., 2000). The efficacy of the serotonin re-uptake inhibitors (SSRI) suggests clinically the importance of serotonergic neurotransmission for the pathogenesis of depression.

The function of NA and DA has been studied less than that of 5-HT. However, considerable experimental evidence, neuroimaging techniques, and clinical evidence support the role of NA and DA (Elhwuegi 2004; Nutt et al., 2007). DA neurons play a key role in decreased incentive motivation, anhedonia, loss of interest and reward, the processes that are disrupted in depression, particularly in the case of melancholic states. NA is associated with loss of energy, fatigue, and low mood (Cowen 2005; Nutt et al., 2006). In addition, the reciprocal interactions between 5-HT, NA, and DA systems may account for the full picture of depression (Nutt et al., 2006; Trivedi et al., 2008).

I.2.2. Hypothalamic-Pituitary-Adrenal axis (HPA)

The HPA axis consists of a feedback loop including the hypothalamus, pituitary, and adrenal glands. The axis receives important regulation from the hippocampus, amygdala, bed nucleus of the stria terminalis, and paraventricular nuclei. It is frequently stated that about half of the patients whose depressive disorder is at least moderately severe, and those with melancholic features,

hypersecrete cortisol. In addition, MDD is associated with early escape from dexamethasone-induced cortisol suppression and a blunted adrenocorticotropic hormone (ACTH) response to corticotropin releasing hormone (CRH) or dexamethasone/CRH challenge (Nemeroff and Vale 2005). The cause is not clearly established. Although researchers had initially thought that cortisol changes might simply be a marker of distress or depression, the view that it plays a provocative role in this regard has received increasing attention lately. Nevertheless, it has been suggested that the balance between glycocorticoid and mineralocorticoid receptors may be a pivotal factor in determining stress reactions and depressive outcomes. In addition, there is growing evidence that CRH hyperfunctioning in hypothalamic and extra-hypothalamic sites (locus coeruleus, amygdala, hippocampus, and nucleus accumbens), CRH1 receptor, and arginine vasopressin (AVP) are associated with depression (Anisman et al., 2008). The data indicate that in some patients stressful life experiences may interact with a predisposition to abnormal HPA axis regulation to produce sustained HPA overactivity (Cowen 2005).

There is some evidence that corticosteroids regulate the genomic expression and function of monoamine receptors in the brain, which could lead, for example, to a decrease in 5-HT neurotransmission (Cowen 2005).

1.2.3. Brain structure in depression

Computerized tomography (CT) and magnetic resonance imaging (MRI) have identified a number of abnormalities in MDD patients, particularly in those with more severe and chronic disorders. The most consistent findings include enlarged lateral ventricles, volume loss in frontal and temporal lobes, a decreased hippocampal volume, and a decreased volume of basal ganglia structures. Researchers have suggested that changes in the brain volume may represent long-term consequences of depression, perhaps associated with cortisol hypersecretion. However, there is also some evidence that changes are manifested early in the course of the illness and represent vulnerability factors (Frodl et al, 2002). In MDD, increased deep white matter hyperintensities are associated with the late onset of depression, greater illness severity and poorer treatment response, apathy, psychomotor slowness, and retardation (Chen et al., 2006; Steffens and Potter 2008; Taylor et al., 2003). A general report of imaging studies concludes that patients with structural abnormalities are less likely to respond to treatment (Cowen 2005).

I.3. Genetics of MDD

MDD is a complex disorder that does not result from either genetic or environmental influences alone but rather from both. Family, adoption, and twin studies help to delineate genetic and environmental effects in humans. Sullivan et al. have meta-analysed the available studies and found that in five family studies the odds ratio for proband versus first-degree relative status was 2.84. Statistical summation of five twin studies suggested that familial aggregation was due to additive genetic effects, with a minimal contribution of environmental effects common to siblings and substantial individual-specific environmental effects. The recurrence best predicts the familial aggregation of MDD (Sullivan et al., 2000)

The first genome-wide linkage survey identified nineteen statistically significant chromosomal regions for MDD [1p, 1q, 2q (2), 4q, 5q (2), 8p, 10p, 10q (3), 11pter, 11q, 15q, 18q, 19p, 19pericentric, Xq]; ten of those were highly-significant linkages (Zubenko et al., 2003). A year later the same team reported about chromosomal loci of genes that influence the risk of suicidal behaviour [2p12, 6q12, 8p22-p21 and Xq25-26.1] (Zubenco et al., 2004). Additionally, Zubenko et al. (2002) found 2q33-34 to be related to recurrent, early-onset MDD in women and Abkevich et al. (2003) showed that 12q22-12q23.2 was related to MDD in men. Some studies have focused on chromosome 15q (Holmans et al., 2004; Verma et al., 2008) and chromosome 10 (Neff et al., 2008).

In association studies in MDD, candidate genes regularly include members of the main neurotransmitter systems, such as monoamines, glutamate, and also pathways that influence several neuroendocrine systems, for example, HPA-axis (Lekman et al., 2008). Such candidate alleles are chosen on the basis of the current understanding of the biology of the disease. The simplest and common form of association studies is the case-control approach; unrelated affected individuals and unrelated healthy controls are genotyped to investigate whether the hypothesized susceptibility gene variant is overrepresented in the affected group. Several meta-analyses have been done in that area with conflicting results. Controversial findings have been published, for example, about the serotonin transporter linked promoter region, catehol-O-methyltransferase gene, promoter region of monoamine oxidase A gene, dopamine receptor 2, interleukin-1beta, dopamine β -hydroxylase, tryptophan hydroxylase, tyrosine hydroxylase genes, and brain-derived neurotrophic factor locus (Furlong et al., 1998a,b, 1999; Ho et al., 2000; Johansson et al., 2001; Kõks et al., 2006; Schumacher et al., 2005; Wood et al., 2002; Yu et al., 2003). That is why the NIMH support efforts to identify the most heritable, more homogenous subtypes and endophenotypes of mood disorders for genetic studies (Merikangas et al., 2002). Previous evidence from genetic association studies on cytokine genes in depression has been limited and inconsistent. No significant associations between polymorphisms from IL10 (Jun et al., 2002), IL6 (Hong et

al., 2005) and IL1-beta (Yu et al., 2003) genes and MDD have been found. On the other hand, the same IL1-beta polymorphisms have been found to be a risk factor for the appearance of depressive symptoms in patients with schizophrenia spectrum disorders (Rosa et al., 2004) and with Alzheimer's disease (McCulley et al., 2004). Furthermore, polymorphisms from monocyte chemoattractant protein-1 (MCP1) (Pae et al., 2004) and the tumour necrosis factor-alpha (TNF) (Jun et al., 2003) genes may have a potential role for susceptibility to MDD. Our recent study established an increased risk of MDD related to the IL20 and IL24 haplotype although none of the SNPs were individually associated with MDD (Traks et al., 2009). In another unpublished study we scanned a large number of single-nucleotide polymorphisms (SNPs) located on the chromosomal region 1q32, which contains four genes from IL10 family: IL10, IL19, IL20, and IL24, in groups of patients with major depressive disorder (MDD, n=312) and panic disorder (PD, n=210), and matched the findings with healthy controls (n=356). We found no significant associations between the SNPs of IL-10 family genes and MDD or PD.

One of the latest approaches in genetics is pharmacogenetics. Pharmacogenetic strategy studies how genetic variation could affect the response of patients to psychotropic drugs and their susceptibility to adverse drug reactions. These studies cold be useful to predict response rate to different antidepressants (rev by Rausch 2005; Serretti 2005; Tsao et al., 2006).

2. Immune system cells and cytokines

The primary function of the immune system is to protect the individual from bacterial and viral insults. The most important immune cells are monocytes, Tand B-lymphocytes, neutrophils, and natural killer cells. Monocytes (and also macrophages and dendritic cells) recognize microorganisms, take them up via phagocytosis, and degrade the microorganisms in small peptides that bind with endogenous major histocompatibility class II proteins. This complex is expressed at the cell membrane in such a way that T- and B-lymphocytes can recognize a foreign protein. Neutrofils are phagocytic cells at the site of the infection; natural killer cells destroy infected or malignant cells. Lymphocytes are cells of the acquired immune system; their actions are antigen-specific. B-lymphocytes recognize a membrane-bound antibody and then proliferate and differentiate into antibody-producing plasma cells. These antibodies opsonize the respective microbes, which facilitates phagocytosis by phagocytic cells. T-lymphocytes can be divided into two classes: T-helper cells (Th cells) and cytotoxic T cells (Tc cells). Activated Th-cells secrete certain molecules (cytokines) that regulate the activity of other immunecompetent cells; Tc cells mainly destroy cells infected with intracellular microorganisms (Aniaman et al., 2008; Van West et al., 2005).

5

Cytokines are small (15 to 44 kD) glycoproteines which function as signalling molecules between different immune cells. In addition to immune cells, they are produced by endothelial, epithelial, and neuronal cells.

2.1. Classification of cytokines

In the context of major depression, two major groups of cytokines are important: pro-inflammatory and anti-inflammatory cytokines. Pro-inflammatory cytokines are mainly produced by activated immune cells and stimulate others, so they enhance inflammatory reactions. Anti-inflammatory cells tend to inhibit activated cells. Monocytes and macrophages initially produce pro-inflammatory cytokines interleukin-1 (IL-1), tumour necrosis factor- α (TNF α), interleukin-6 (IL-6) and interleukin-12 (IL-12); after the initial activation anti-inflammatory cytokines or proteins IL-10 and IL-1 receptor antagonist (IL-1RA) are produced by these cells.

Th-1 cells produce interferon- γ (IFN γ), IL-2 and TNF α , tumour necrosis factor- β , IL-12, and IL-18. Th-2 cells produce IL-3, IL-4, IL-5, IL-10, IL-13, and IL-6. Th-1 cytokines stimulate cell-mediated immunity (mainly phagocytic cells); Th-2 cells promote humoral immunity (antibodies, allergic reactions) (Schwarz et al., 2001; Van West et al., 2005).

2.2. Cytokines and depression

Growing evidence suggests that, in addition to providing communication between immune cells, specific cytokines play a role in signalling the brain to produce neurochemical, neuroendocrine, neuroimmune, and behavioural changes. For example, pro-inflammatory cytokines, such as stressors, increased HPA axis functioning and influenced a range of monoaminergic and peptidergic extra-hypothalamic sites (rev by Anisman et al., 2008; Kronfol and Remick 2006). The pro-inflammatory cytokines TNF- α , IL-1 and IL-6 are primary HPA stimulating cytokines. IL-6 is a potent stimulator of CRH production, which leads to elevated HPA activity characterized by increased ACTH and cortisol levels (O'Brien et al., 2004). In addition, pro-inflammatory cytokines acutely stimulate 5-HT turnover and reduce the production of 5-HT by stimulating the enzyme indoleamine 2,3-dioxygenase (IDO), which converts tryptophan the precursor of 5-HT, into kynureine (Wichers and Maes 2002; Schiepers et al., 2005). IL-1, interferon- α (IFN- α), IFN- γ , and TNF- α have been shown to upregulate the serotonin transporter, which may reduce extracellular 5-HT levels (Hayley et al., 2005; Miller and Raison 2006; Wichers and Maes 2002) and anti-inflammatory cytokine IL-4 was shown to induce a reduction of 5-HT uptake, so that the synaptic level of 5-HT synaptic level increases (Mössner et al., 2001). Some underlying mechanisms have been reported from animal studies; for example, IL-1 and TNF- α act on serotonin transporter by activating P38 mitogen-activated protein (MAP) kinase (Miller and Raison 2006).

There is evidence that patients who receive cytokine immunotherapy frequently show depressive symptoms, which may be attenuated by antidepressant medication. This fact supports the causal role of cytokines in MDD (Capuron et al., 2004; rev Capuron and Miller 2004). However, despite the link with depression and immunotherapy, the question remains open whether the effects observed are a genuine manifestation of neurochemical changes underlying depression, or whether the symptoms are rather a reflection of general malaise or toxicity. Recent studies seem to have indicated that the neurovegetative and mood-related features introduced by IFN- α therapy are independent of one another, as paroxetine primarily affected the mood-related symptoms, with only minor effects on fatigue and anorexia (Raison et al., 2005).

Because cytokines are closely associated with central neurotransmitters and cytokine regulation is affected by stress, a number of studies have investigated the possible role of cytokines in major psychiatric disorder, including major depression. Maes et al. reported an increased plasma concentration of IL-1 (1993), IL-6 (1995 and 1997), sIL-2R, soluble IL-6 receptors (sIL-6R), and acute phase proteins (1995) in depressed patients. They concluded that there is an increase in pro-inflammatory cytokines in MDD patients. Unfortunately, these observations have not been consistently replicated. Table 1 provides a brief summary of controversial studies on cytokines in MDD patients versus healthy controls.

The association between cytokine levels and scores of depression rating scales is also unclear. Leo et al. (2006) reported IL-1 β , IL-6, and TNF- α level to be related to HAMD scores. By contrast, Kagaya et al. (2001) and Tuglu et al. (2003) did not found any correlation between TNF- α concentration and HAMD scores; the latter authors used additionally the BDI scale, but no correlation was still found. There is evidence that cytokine activity may be related to chronicity of illness, neurovegetative features, or some other aspects (such as typical or atypical, melancholic or non-melancholic, sleep pattern) or symptoms of MDD (Anisman and Merali 2002).

There is evidence implying that antidepressive treatment with various antidepressive agents has an immunomodulative effect. In vitro studies suggest that when human monocytes are incubated with different classes of antidepressants together with bacterial cell wall lipopolysaccharide (LPS), which stimulate the release of pro-inflammatory cytokines, the synthesis and release of IL-1, IL-6, and TNF- α is markedly inhibited (Xia et al., 1996). Additionally, Kubera et al. (2000a) showed that an increased concentration of serotonin after administration of SSRI antidepressants is associated with an increased release of anti-inflammatory cytokine IL-10 and a decreased synthesis of INF- γ .

The effects of antidepressants on the activity of the immune system have been studied in animal studies. The enhanced lymphocyte proliferation and an increased production of IL-1 and IL-2 in rats under chronic mild stress is reversed following chronic treatment with imipramine (Kubera 1996) while the production of IL-10, an anti-inflammatory cytokine, increases after desipramin treatment (Kubera 2001b). A decreased production of TNF- α after the administration of different antidepressants has been reported several times in animal studies (Brustolim et al., 2006; Obuchowicz et al., 2005, Reynolds et al., 2005; Roumestan et al., 2006). However, data about treatment effects on immune system activity are controversial. Table 2 summarizes the findings of antidepressant effects in HC, and Table 3 does the same in MDD patients.

Authors	IL-1 α	L-1 α IL-1 β	IL-2	IL-4	IL-6	IL-7	IL-8	IL-10	IL-12	IL-15	IFN-γ	$TNF-\alpha$	TGF-β1	sIL-2R	sIL-6R	II-1Ra
Maes et al., 1993	Ļ															
Weizman et al., 1994		\rightarrow	\rightarrow													
Maes et al., 1995					Ļ									←	Ļ	
Maes et al., 1997					Ļ										N	←
Frommberger et al.,					←											
1997 Song et al., 1998					~		~									~
Kubera et al., 2000b					ŧ			ŧ								ŧ
Lanquillon et al., 2000					u							~				
Kagaya et al., 2001		N			u							и		u		
Mikova et al., 2001					N									22		
Rief et al., 2001					N										N	Ļ
Rothermundt et al.,		N		·												
2001												_				
Nunes et al., 2002		22			\rightarrow							22		$\overrightarrow{\imath}$		
Tuglu et al., 2003												Ļ				
Bauer et al., 2003												N		22		
Brambilla et al., 2004		22										N				
Kubera et al., 2004					Ļ							N				
Schlatter et al., 2004			Ļ	22												
Basterzi et al., 2005					N											
Myint et al., 2005				N							22		\overrightarrow{v}			

Table 1. Cytokine levels in different studies in MDD patients vs healthy controls

Table 1. Continued

Authors	IL- 1α	IL-1 β	IL-2	IL-4		IL-7	IL-8	IL-10	IL-12	IL-15	IFN-γ	TNF- α	IL-6 IL-7 IL-8 IL-10 IL-12 IL-15 IFN-y TNF-a TGF-B1 sIL-2R sIL-6R II-1Ra	sIL-2R	sIL-6R	II-1Ra
Lee and Kim, 2006									~				и			
Leo et al., 2006		←			~						<u> </u>	←				
Pike and Irwin, 2006					~						<u> </u>			N		
Kim et al., 2007			\rightarrow	\rightarrow	←						\rightarrow	←	~			
Marques-Deak et al., 2007		u			u						u					
O'Brien et al., 2007					←		11	22				¢			u	
Sutcigil et al., 2007			¢	22					←			¢	22			
Simon et al., 2008	←	←	~	~	~	~	←	←	~	~	~					

Higher levels in MDD patients than in healthy controls Lower levels in MDD patients than in healthy controls $\leftarrow \rightarrow u \quad \overleftarrow{u} \quad \overrightarrow{v}$

No statistical difference between MDD patients and healthy controls

A slight increase, statistically not significant A slight decrease, statistically not significant 1 1

Authors	Study group	Antidepressant	IL-1β	IL-2	IL-4	IL-6	IL-8	1L- 10	IL-12	INF- γ	$\begin{array}{ c c c c c c c c c c c c c c c c c c c$	TGF-β	IL-1Ra
Xia et al., 1996	Healthy volunteers	TCA	\rightarrow			\rightarrow				\rightarrow	\rightarrow		
Maes et al., 1999	Healthy volunteers	Clomipramine Sertraline Trazodone						<i>←</i>		\rightarrow			
Lin et al., 2000	Healthy voluteers	Moclobemide				11	\rightarrow	←		u	\rightarrow		22
Szuster-Ciesielska et al., 2003	Healthy volunteers	Imipramine, Mianserin Lithium		$\approx (US) \approx (US)$	≈(US) ↓(S)			← ←	$\approx (OS) \uparrow (S) \uparrow (S) \uparrow (S) \approx (S) \uparrow (S)$	t (S)↓		≈(US) ↑	
Maes et al., 2005	Healthy volunteers	fluoxetine						22		\rightarrow	\rightarrow		
Diamond et al., 2006	Healthy volunteers	Desipramine Clomipramine Fluoxetine, Reboxetine	$\leftarrow \rightarrow \wr$					\overleftarrow{u} \overleftarrow{u} \overleftarrow{u}		$\rightarrow \rightarrow \rightarrow$	<i>u u u</i>		

Table 2. Effect of antidepressants on different markers of the immune system in healthy volunteers

increased after treatment

I

decreased after treatment no difference between the baseline and at the end of treatment tend to increase after treatment tend to decrease after treatment Stimulated production Unstimulated production I

I

Ι

I $\underset{\mathbf{CS}}{\operatorname{cs}} \underbrace{\mathbf{s}} \underbrace{\mathbf{s}} \underbrace{\mathbf{s}} \underbrace{\mathbf{s}} \underbrace{\mathbf{s}} \leftarrow \rightarrow \\$

I

Table 3. Effect of antidepressants on different markers of the immune system in patients with MDD

						I (
Authors	Study group	Antidepressant $ IL-1\beta IL-2 IL-4$	IL-1 β	IL-2	IL-6	П-8	IL-10	IL-12	INF- γ	TNF-α	TGF-β	IL-8 IL-10 IL-12 INF- γ TNF- α TGF- β sIL-2R sIL-6R IL-1Ra	sIL-6R	IL-1Ra
Weizman et	MDD	Clommipramine	←	22										
al., 1994	patients	4 weeks												
Maes et al.,	MDD	Fluoxetine			22							22	N	
1995	patients	TCA												
Frommberger MDD	MDD	i			\rightarrow									
et al., 1997	patients													
Maes et al.,	MDD	Trazodone			u								\uparrow	u
1997	patients	Trazodone +												
		pindolol												
		Trazodone +												
		fluoxetine												
		5 weeks												
Kubera et al., MDD	MDD	i			N		N							22
2000b	patients	6 weeks												
Lanquillon et MDD	MDD	Amitiiptyline			↑(R)					↓(R)				
al., 2000	patients	6 weeks			↓(NR)					\approx (NR)				
Kagaya et al., MDD	MDD	Mainly	N		N					←		22		
2001	patients	clomipramine,												
		combinations												
		with others												
		1 month												

L-6R IL-1R																				
sIL-2R sII					22												←			
TGF-β																				
TNF-a					N		\rightarrow					22								
INF-7	≈ —	•																		
IL-12																				
IL-10	← ←	-																		-
IL-8					11														_	
IL-6					U							~								
IL4																				
IL-2																				
IL-1β																	\rightarrow			
Antidepressant IL-1 β IL-2 IL-4 IL-6 IL-8 IL-10 IL-12 INF- γ TNF- α TGF- β sIL-2R sIL-6R IL-1Ra	Imipramine, Venlafaxine	Fluoxetine			Different	8 weeks	Sertraline,	Citalopram,	Fluoxetin,	Fluvoxamine	Paroxetine	Imipramine	Venlafaxine	Fluoxetine			Double-blind	(citalopram,	sertraline,	
Study group	ent-	resistant	patients	in vitro	MDD	patients	MDD	patients					treatment-	resistant	patients	in vitro	Chronic	posttraum	atic stress	
Authors	Kubera et al., MDD, 2001a treatme				Mikova et al., MDD	2001	Tuglu et al.,	2003				Kubera et al., MDD,	2004				Tucker et al., Chronic	2004		

ned
ntin
ů.
le 1
Tab

Authors	Study group	Antidepressant	IL-1β IL-2 IL-4	IL-2		IL-6	IL-8	IL-10	IL-12	INF-γ	TNF-α	TGF-β	IL-6 IL-8 IL-10 IL-12 INF- γ TNF- α TGF- β sIL-2R sIL-6R IL-1Ra	sIL-6R	IL-1Ra
Basterzi et	MDD	SSRI				\rightarrow									
al., 2005	patients	6 weeks													
t al.,	MDD	different			\rightarrow					u		←			
2005	patients														
Himmerich	MDD	Different									U				
et al., 2006	patients	groups, also													
		combinations,													
		Li allowed;													
		8 weeks													
Lee and Kim, MDD	MDD	Different							$\overrightarrow{\mathcal{U}}$			↓			
2006	patients	6 weeks													
t al.,	Drug-	Sertralin	\rightarrow			\rightarrow					\rightarrow				
2006	naïve first	Citalopram													
	episode	6 weeks													
	MDD														
	patients														
a et al.,	DDD	Fluvoxamine									\rightarrow				
2006	patients	Paroxetine													
		Milnacipran													
		Longer than													
		6 months													

led
ntinu
Cor
Ι.
le
ab
Ë

Table 1. Continued

Authors	Study	$\Delta ntidemescant = \frac{1}{11 - 18} = \frac{1}{11 - 2} = \frac{1}{11 - 4} = \frac{1}{11 - 6} = \frac{1}{11 - 8} = \frac{1}{10} = \frac{1}{11 - 12} = \frac{1}{101 - 12} = 1$	П_1В	ι τ	V- II	9- Ш	8- П	IT _10	11 _17	INF_~	$TMF_{-\alpha}$	TGF_R	ara	еП _6Р	П_1Ра
CIOINITY	group	MINUTE PROPERTY AND	d1-71	7-11			0-11	N1-11	71-71		m- INTI	d- ID I	117-7110	VID-TIC	11-11/0
Kim et al.,	DDD	Different		\rightarrow	U	\rightarrow				N	22	22			
2007	patients	classes													
		6 weeks													
		in vitro													
Sutcigil et al., MDD	MDD	Sertraline			←				\rightarrow			~			
2007	patients	8 weeks													

1 1

increased after treatment
decreased after treatment
no difference between baseline and at the end of treatment
tend to increase after treatment
tend to decrease after treatment
Stimulated production
Unstimulated production I

3. Thyroid function in major depression

There is evidence that abnormalities in thyroid function are more common in patients with mood disorders than in healthy subjects (Bauer et al., 2008; Joffe and Marriott 2000). Subclinical hypothyroidism is found to be related to an increased risk of elderly depression (Chueire et al., 2007). Thyroid hormones, particularly T3, are known to accelerate the clinical response to antidepressant therapy in MDD (Aronson et al., 1996; Abraham et al., 2006; Agid and Lerer 2003). Additionally, there is evidence that depressive patients with subclinical hypothyroidism respond worse to antidepressant intervention (Duval et al., 1996; Joffe and Levitt 1992). Gitlin et al. (2004) found that low values of the thyroid-stimulating hormone (TSH) correlated with greater improvement of depressive symptoms during treatment with SSRIs.

3.1. Anti thyroid peroxidise auto-antibodies (anti-TPO)

The association between thyroid autoimmunity and mood and anxiety disorders was found in several studies (Carta et al., 2004; Pop et al., 1998), but there have also been some negative findings in that field (Chueire et al., 2007; Engum et al., 2005; Horning et al., 1999). Only a few studies have focused on the impact of anti-TPO positivity to the treatment effect of MDD, and this data is also controversial. Haggerty et al. (1997) reported that the presence of antithyroid antibodies predicts a poor response to antidepressant treatment. However, Fountoulakis et al. (2004) failed to demonstrate such an association.

AIMS OF THE STUDY

The general aim of the study was to explore a possible involvement of IL-8, TNF- α , and sIL-2R in the pathogenesis of MDD. Based on this, the specific aims of the study were as follows:

- 1. To compare TNF- α and sIL-2R serum levels between patients with major depression and healthy controls.
- 2. To find correlations between the levels of cytokines and the severity of depression measured by HAM-D in patients with major depression.
- 3. To find correlations between the levels of cytokines and single symptoms of depression according to HAMD items.
- 4. To examine effects of escitalopram treatment on the levels of IL-8, TNF- α , and sIL-2R in patients with major depression.
- 5. To find out whether bupropion augmentation changes the production of IL-8, TNF- α , and sIL-2R in escitalopram-resistant patients with major depression.
- 6. To detect possible associations between IL-8, TNF- α , and sIL-2R serum concentrations and treatment response in patients with major depression.
- 7. To find out whether anti-TPO positivity or thyroid hormones have an impact on efficacy of escitalopram treatment in patients with major depression.

8

MATERIALS AND METHODS

I. Ethical considerations

The Ethics Review Committee on Human Research of the University of Tartu approved the study protocols and the informed consent forms of the subjects. All participants signed the written informed consent.

2. Characteristics of study participants and study design

All the subjects who participated in this study – patients and healthy controls – were Caucasians living in Estonia. MDD patients were recruited at the Psychiatric Clinic of the Tartu University Hospital. The age of all subjects was between 15 and 65 years. The diagnosis according to DSM-IV criteria was verified using M.I.N.I. 5.0.0 and substantiated by psychiatric history and medical records. To assess the severity of depressiveness, HAMD and BDI scales were used; MADRS was used additionally in the treatment phase. All healthy subjects were interviewed using the M.I.N.I. 5.0.0, and only those without a personal or family (defined as first-degree relatives) history of psychiatric disorders and not taking medications were included in the study. There were no significant age and gender differences between the patients and the healthy volunteers.

129 MDD patients were selected for the treatment phase with escitalopram. These subjects were required to have a MADRS score at least 23 or higher and the wash-out period from previous antidepressive drugs had to be at least two weeks, if the subject had received treatment for current depressive episode. No other regular medication, including anti-inflammatory drugs, was allowed during the study, except for hormonal contraceptives and zolpidem or zopiclon for insomnia. All patients started treatment with 10 mg escitalopram per day for the first 4 weeks. The patients showing at least a 50% decline in the MADRS total score at week 4 continued taking 10 mg of escitalopram until the end of the study. The dose of escitalopram was increased and kept at 20 mg in patients who demonstrated less than a 50% decrease in the MADRS score at week 4 or who showed exacerbation of depressive symptoms during any of the following visits. At the end of week 12 the patients were defined as responders (R) if the decrease in the MADRS total score was at least 50% and as remitters if the score was less than 12. The patients who did not meet these criteria were defined as non-responders (NR). As almost all responders fulfilled the criteria of remission on the MADRS, the analyses were made only between the groups of R and NR. Bupropion 150-300 mg per day was added to escitalopram in those NRs who agreed to continue the study (n=28) for additional 6 weeks. At the end of the augmentation period the patients were again defined as R if the decrease in the MADRS total score was at least 50% during this period or as NR if the MADRS total score decreased less than 50%. All the patients were visited every two weeks; blood samples for cytokines were taken at week 0, week 4, week 12, and week 18; for antibodies blood was collected only once, during the baseline visit. Table 4 shows the characteristics of the study participants.

	Total number of participants	Sex (male/female)	Age (year) Mean ± SD	Age range (years)
MDD patients	247	73/174	33.5 ± 12.8	17–63
MDD patients in	100	35/65	32.1 ± 11.9	19–63
the first treatment				
phase				
MDD patients in	28	11/17	31.2 ± 9.5	19–48
the augmentation				
phase				
Healthy controls	94	36/58	32.5 ± 13.3	15-66

Table 4. Characteristics of study participants

3. Laboratory analyses

Initial cytokine selection was made on the basis of the literature: IL-1 β , IL-6, IL-8, IL-10, TNF- α , and sIL-2R. Unfortunately, it was not possible to measure IL-1 β , IL-6, and IL-10 in our patients. The blood was collected between 9.00 and 11.30 a.m. for all the study groups. After complete clot formation the samples were centrifuged, and the serum was divided. The probes were collected and analysed in one batch by means of the IMMULITE system using solid-phase, enzyme labelled, and chemiluminescent sequential immunometric assay. The intra-assay coefficient of variation for sIL-2R was 3.7%, 3.8% for IL-8, and 3.6% for TNF- α ; the inter-assay coefficients were 8.1%, 7.4%, and 6.5%, respectively.

Anti-TPO testing was performed for 129 patients using the ImmunoCAP 100 system (Phadia, Uppsala, Sweden). TSH, total T3, freeT3, and freeT4 were assessed by means of the chemiluminescence method, using the IMMULITE200 analyser. The reference values are 0.4–4.0 mU/L for TSH; 1.3–2.8 nmol/L for total T3, 2.7–6.5 pmol/L for free T3, and 10.3–25.0 pmol/L for free T4. The coefficients of variance for these hormones were less than 10% for freeT3, 9% for free T4, 12.5% for TSH, and 15% for T3. The anti-TPO test values over 100 IU/ml were taken as positives.

4. Statistical analysis

The analyses were performed using the software package Statistica 7.0 (Tulsa, OK, USA). As cytokine levels did not follow the Gaussian distribution, logarithms were used to normalize the data. The significance level of the tests for declaring a probability value as significant was set at 0.05. Different statistical tests were used in different studies and are described in the publications.

RESULTS

I. Differences in cytokine levels between MDD patients and healthy controls

The levels of sIL-2R and TNF- α were compared between 75 currently depressed subjects, 17 patients in full remission and 55 healthy controls in Study I (demographic data in Table 5). First, sIL-2R and TNF- α were compared in the 4 study groups: MDD with the recurrent depressive episode (RE), MDD with the first episode (FE), MDD in full remission (FR) and healthy controls (HC). The results showed a significant difference in the level of sIL-2R between the groups (Table 5). The levels of sIL-2R were significantly lower in FR than in RE and HC. There was a trend towards a lower level of sIL-2R in FR compared to FE. Previous use of antidepressants did not influence these results.

No group differences were found in the levels of TNF- α between 4 groups (Table 5), but a comparison of the currently euthymic subjects (HC and FR) and depressed subjects (FE and RE groups) showed lower levels of TNF- α in the currently depressed subjects. Additionally, the subjects with previous antidepressive treatment had significantly lower levels of TNF- α and differed significantly from drug-naïve patients and HC. There was no difference between HC and drug naïve patients.

When only drug-naïve patients, drug naïve remissions, and HC were included in the analysis, REs were associated with increased levels of sIL-2R by comparison with FE, FR, and HC. There was no difference in the levels of TNF- α between the groups.

HAM-D scores were significantly and positively associated with TNF- α but not with sIL-2R levels in the currently depressed patients. BDI scores were not related to the levels of TNF- α and sIL-2R. Both biomarkers did not correlate with the number of depressive episodes, with the duration of the current episode, smoking habits, or melancholic features. Additionally, it appeared that sIL-2R levels were related to two HAM-D items: decreased activity (the 7th item of HAM-D) and agitation (the 9th item of HAM-D). TNF α levels were associated with decreased activity and suicidality (the 3rd item of HAM-D). IL-2R and TNF α levels were not related to any BDI items.

Table 5. The demographic data, the mean scores of HAM-D and BDI and the concentrations of interleukin-2 receptor (IL-2R) and tumour necrosis factor alpha (TNF α) in the Study I groups with statistical comparisons.

	FE	RE	FR	HC	p-value
Male/female	4/8	14/49	7/10	23/32	*NS
$A = (\pm SD)$	32.50	37.24	35.76	32.75	**NS
Age (±SD)	(14.28)	(12.35)	(14.54)	(14.10)	IND IND
BMI (±SD)	25.38	22.22	22.99	24.15	**NS
$DMI(\pm 5D)$	(4.02)	(3.24)	(2.90)	(4.47)	
HAMD (±SD)	24.27	24.14	3.19	1.08	***<0.001
$HAMD(\pm SD)$	(4.54)	(3.11)	(3.54)	(1.22)	0.001
PDI (+SD)	30.40	30.08	4.85	4.32	***<0.001
BDI (±SD)	(7.88)	(8.86)	(3.98)	(4.81)	0.001
Melancholic	8/4	56/7			*=0.050
symptoms Yes/No	0/4	30/7			-0.030
IL-2R (kU/l)	431.75	506.90	354.94	453.55	***<0.001
Mean (±SD)	(111.24)	(174.06)	(142.78)	(136.10)	0.001
TNFα (ng/l)	5 48 (1 72)	6.46	7.72	7.29	***NS
Mean (±SD)	5.48 (1.72)	(3.16)	(3.74)	(3.56)	113

*Fisher's exact test

**Kruskal-Wallis' test

*** Group - effect of ANOVA

post hoc for HAMD: FE/RE (p=0.889), FR/HC (p=0.008), FE/HC (p=0.000), FE/FR (p=0.000), RE/HC (p=0.000), RE/FR (p=0.000)

post hoc for BDI: FE/RE (p=0.894), FR/HC (p=0.812), FE/HC (p=0.000), FE/FR (p=0.000), RE/HC (p=0.000), RE/FR (p=0.000)

post hoc for IL-2R: FE/RE (p>0.05), FR/HC (p=0.004), FE/HC (p>0.05), FE/FR (p=0.080), RE/HC (p>0.05), RE/FR (p=0.0001)

2. Escitalopram treatment effects on IL-8, TNF-α, and sIL-2R levels in MDD patients

The treatment effects of escitalopram on IL-8, TNF- α , and sIL-2R levels in MDD patients were assessed in Study II. In this study, the study group consisted of 100 patients (35 males and 65 females) and 45 HC (19 males and 26 females). The demographic and clinical data of the study cohort are presented in Table 6.

There were no significant differences in age or sex distribution between the R and NR or between the patient groups and HC. The NR had more previous depressive episodes, earlier age of disease onset, and was more melancholic and less drug-naive than R. At baseline, the severity of depression on MADRS did not significantly differ between the R and NR groups.

There was a statistically significant effect of group x time interaction but no group effect in sIL-2R measurements during the study (weeks 0, 4, and 12). There were different patterns of sIL-2R changes for R and NR – in the NR group sIL-2R decreased significantly between weeks 4 and 12 and in the R group between weeks 0 and 4. No significant effects of escitalopram treatment could be reported for either IL-8 or the TNF- α level. By week 12 there were no differences in cytokine levels between the 3 study groups (Table 7).

Table 6. Demographic an	d clinical data a	nd the baseline r	neasurements of t	he cytokines
of the study II cohort: he	althy volunteers	s, responders and	d non-responders	to treatment
with escitalopram				

Variables	Healthy	Responders	Non-	p-value
	volunteers		responders	
Number of patients	45	74	26	
Gender (male/female)	19/26	29/45	6/20	** NS
Age	32.9. (14.1)	31.5 (±12.2)	34.2 (±11.0)	****NS
Number of episodes		4.60 (±5.08)	7.25 (±6.63)	*<0.01
Age of onset of the first		23.62 (±10.47)	19.67 (±10.32)	*=0.050
episode				
Duration of current		10.8 (±14.3)	14.7 (±16.8)	*NS
episode (months)				
Melancholic symptoms		50/24	23/3	**<0.05
(with/without)				
Drug-naive/previously		44/32	9/15	**<0.05
treated				
MADRS before the		28.5 (±5.9)	29.5 (±4.4)	*NS
treatment				
MADRS at the end of the		4.5 (±5.1)	22.8 (±6.3)	*<0.001
treatment				
Baseline sIL-2R (kU/l)	471.17	524.56	499.18	***NS
	(±136.57)	(±175.33)	(±138.90)	
Baseline IL-8 (ng/l)	7.74 (±1.95)	6.31 (±1.95)	6.64 (±1.99)	***NS
Baseline TNF-α (ng/l)	6.42 (±1.94)	5.70 (±1.55)	6.38 (±2.02)	*** <0.05

*t-test

**Chi-squire test

***group-effect of ANCOVA with age as covariance and gender as second factor

****t-test with Bonferroni correction

	sIL-2R	(kU/l)	IL-8	(ng/l)	TNF-α	(ng/l)
Time	Responders	Non- responders	Responders	Non- responders	Responders	Non- responders
Week 0	524.56	499.18	6.31	6.64	5.70	6.38
	(±175.33)	(±138.90)	(±1.95)	(±1.99)	(±1.55)	(±2.02)
Week 4	493.99	518.18	6.46	7.66	5.91	6.79
	(±167.42)	(±154.08)	(±2.04)	(±2.67)	(±1.90)	(±2.24)
Week 12	515.42	451.00	6.93	7.09	6.27	6.40
	(±208.65)	(±109.66)	(±2.32)	(±2.64)	(±1.94)	(±2.38)
Р*	< 0.05		NS		NS	

Table 7. Measurements of the cytokines in escitalopram treatment-week 0, 4 and 12 in responder and non-responder groups

*time × group effect of RM design ANOVAs

3. Bupropion augmentation effects on IL-8, TNF-α, and sIL-2R levels in escitalopram-resistant MDD patients

The MDD patients who did not respond to 20 mg escitalopram treatment had a possibility to continue the study in the augmentation phase. Twenty-eight patients were selected for Study III. The HC group was the same as in Study II. The demographic and clinical assessment data of patients are presented in Table 8. There were no significant differences in age, sex, or body mass index (BMI) between R, NR, and HC. The NR scored significantly higher on MADRS both before and after the treatment.

There were no group or group x time interaction effects in the augmentation phase. However, there was a significant time effect for IL-8 as the levels of IL-8 increased during 6 weeks of treatment (Table 8). No correlations were noticed between cytokine levels and the severity of depression on MADRS total scores at any time of measurement.

Variables	Responders	Non-responders	p-value
Number of patients	18	10	
Gender (male/female)	5/13	6/4	NS
Age	31.7 (±9.8)	30.4 (±9.4)	NS
BMI	23.9 (±3.7)	24.9 (±3.2)	NS
Number of episodes	7.3 (±8.0)	6.7 (±9.7)	NS
Age of onset of the first episode	18.3 (±6.7)	18.6 (±8.8)	NS
Duration of current episode (months)	13.9 (±16.8)	14.9 (±15.1)	NS
Melancholic symptoms (Yes/No)	15/3	8/2	NS
MADRS before the combined	19.8 (±4.3)	25.7 (±6.9)	=0.010
treatment			
MADRS at the end of the treatment	7.3 (±4.6)	24.6 (±5.5)	<0.001
sIL-2R kU/l (±SD) week 0	525.47	493.11 (143.56)	a) NS
	(265.11)		b) NS
sIL-2R kU/l (±SD) week 6	563.78	534.80 (151.47)	c) NS
	(216.47)		
IL-8 ng/l (±SD) week 0	6.54 (2.13)	7.35 (3.36)	a) NS
			b) < 0.05
IL-8 ng/l (±SD) week 6	6.94 (2.34)	7.64 (2.29)	c) NS
TNF-α ng/l (±SD) week 0	6.55 (2.89)	6.37 (2.47)	a) NS
			b) NS
TNF-α ng/l (±SD) week 6	7.04 (2.85)	7.21 (2.41)	c) NS

Table 8. Demographic and clinical characteristics and concentrations of sIL-2R, IL-8 and TNF- α in responders (R) and non-responders (NR) in bupropion augmentation

a) RM ANOVA Group effect

b) RM ANOVA Time effect

c) RM ANOVA Time x group effect

4. Associations between IL-8, TNF-α, and sIL-2R baseline serum concentrations and treatment response in MDD patients

In the treatment phase I (Study II) the comparison of baseline cytokine levels between R, NR, and HC demonstrated a statistically significant between-group difference for TNF- α but not for other cytokines (Table 6). R showed a lower baseline TNF- α level in comparison with NR or HC, whereas the two latter groups did not differ from each other. However, there was a significant gender effect – NR males had a higher level of TNF- α than R males, NR females, or R females.

In the augmentation phase, the baseline levels of IL-8, TNF- α , and sIL-2R did not significantly differ between the R, NR and HC groups (Table 6).

5. Thyroid function and treatment response

Anti-TPO positivity was found in eight depressive and two healthy females without a statistically significant difference between these groups. As anti-TPO was not seen in either of the male groups, all further statistical analyses were carried out only in females. There were no significant differences in the levels of thyroid hormones (particularly, total T3, free T3, freeT4, and TSH) between female responders and non-responders; however the latter group showed a tendency for a higher prevalence of anti-TPO than the responders. Eleven patients had elevated total T3 and/or free T3 and/or free T4 levels, and one of them had anti-TPO.

Variable	Responders (n=60)	Non- responders (n=30)	P (Mann-Whitney) P* (Chi-square) P** (Fisher exact test)
	25.2 . 12.1	22.5 . 10.5	210
Age (years \pm SD)	35.2 ± 13.1	32.7 ± 10.5	NS
Anti-TPO (pos/neg)	3/57 (5.3 %)	5/25 (20.0 %)	***NS
MADRS baseline $(\pm SD)$	28.1 ± 4.7	29.1 ± 5.3	NS
MADRS endpoint (± SD)	3.8 ± 3.7	23.0 ± 6.4	<0.001
HAMD baseline $(\pm SD)$	20.2 ± 4.0	21.4 ± 5.0	NS
HAMD endpoint $(\pm SD)$	3.8 ± 3.2	16.8 ± 5.6	<0.001
Duration of current episode (months ± SD)	13.1 ± 17.8	12.1 ± 15.2	NS
Age of onset of the first episode (years \pm SD)	26.8 ± 12.3	18.6 ± 8.9	<0.005
Number of previous episodes (± SD)	4.2 ± 5.0	7.7 ± 8.1	<0.05
Comorbid anxiety	23/60	17/30	*NS
Comorbid melancholia	45/60	25/30	*NS

Table 9. Demographic and clinical data of female responders and non-responders.

DISCUSSION

Ι. ΤΝF-α

There were no significant differences in the TNF- α levels between main groups: RE, FE, FR, and HC. Study I showed lower TNF- α serum levels in the currently depressed than euthymic subjects. Further analysis revealed that the lower levels of TNF- α were associated with previous antidepressive treatment and were not found in drug-naïve patients. Narita et al. (2006) reported that the levels of TNF- α were significantly lower in remitted MDD patients receiving maintenance antidepressive treatment for longer than 6 months in comparison with the healthy controls. Unfortunately, it is not clear how long this immunosuppressive effect could last after the discontinuation of depression treatment. The lower than normal levels of TNF- α were also observed in young patients with dysthymia, but not in those with MDD (Brambilla et al., 2004).

TNF- α is a multifunctional cytokine which participates in the pathogenesis of various diseases, including autoimmune, inflammatory, neurodegenerative diseases, diabetes, septic shock, and congestive heart failure (Tayal and Kalra 2008), and it has been associated with psychiatric disorders, including MDD (Table 1).

Although the general opinion is that MDD is associated with higher levels of pro-inflammatory cytokines, especially IL-6, IL-1 β , and TNF- α , not all human studies, including study I for TNF- α , have reported an increase in pro-inflammatory cytokines in depressed patients versus healthy controls (Table 1). Like in HPA axis activity, which could be hyper- or hypoactive depending on subtype of depression (Antonijevic 2006; De Beaurepaire 2002), it may be that some subgroups have opposite reactions in cytokine profiles, e.g. a decrease in pro-inflammatory cytokine production. Our study suggests this hypothesis with the finding of lower levels of TNF- α in currently depressed patients. Lower TNF- α level could be a state marker, as in remission phase there were no differences between healthy and affected subjects. This is in agreement with a study by Kagaya et al. (2001), showing that after pharmacotherapy TNF- α levels of depressed patients increased. This hypothesis needs to be further tested in different subgroups of patients with MDD.

There is also a finding of midlife women reporting higher levels of depressive symptoms associated with a decreased in vitro production of IL-1 β , IL-6, and TNF- α compared with their less-depressed counterparts (Cyranowski et al., 2007). Similarly, TNF- α level was significantly lower in healthy students with high anxiety scores during psychological stress (Chandrashekara et al., 2007). These reports suggest the relationship between decreased synthesis of pro-inflammatory cytokines and symptoms of depression and anxiety. Like in major depression, the data are also controversial in healthy controls. Maes et al. (1997a) found that, in students, examination stress significantly increased the stimulated production of TNF- α . Higher BDI scores were associated with

greater expression of TNF- α (Suarez et al., 2004). However, Marsland et al. (2007) did not find significant associations between TNF- α and any psychosocial parameters in middle-aged community volunteers.

The possible reasons for conflicting findings could be explained by different study cohorts, subtypes of depressive disorders, and different cytokine measurement techniques. Depression is often (approximately 50%) associated with HPA axis hyperactivity (Cowen 2005), and deregulation of the feedback mechanism appears to occur in depressive disorders (Schiepers et al., 2005). Pro-inflammatory cytokines are potent activators of the HPA axis and play a critical role in activating the HPA axis in MDD. These cytokines counteract the negative feedback action of corticosteroids on the HPA axis (Myint and Kim 2003; Schiepers et al., 2005). There is a hypothesis that cytokines could induce corticosteroid receptor resistance in the hypothalamus and the pituitary gland (Pace et al., 2007) – a higher level of pro-inflammatory cytokines implies a stronger resistance of corticosteroid receptors.

2. TNF- α in the escitalopram-treatment phase

The lower levels of TNF- α in Study II were associated with a better treatment response. The R group of patients had lower levels of TNF- α than NR or HC. There was no difference between NR and HC. Bauer et al. (2003) have previously reported that patients with treatment-resistant depression did not differ from HC in their baseline levels of TNF- α and sIL-2R. There is also evidence that elevated HPA axis activity in acute depression suppresses TNF- α activity, while in remission, when HPA axis activity is normalized, the TNF- α system seems to gain influence on the HPA system (Himmerich et al., 2006).

The difference between responders and non-responders has been assessed in several studies. After a six-week treatment period with amitriptyline, TNF- α levels normalized only in responders (Lanquillon et al., 2000). However, the pre-treatment levels of TNF- α in this study were increased in both, responders and non-responders as compared to healthy controls.

TNF- α activates serotonin transporters, providing a mechanism by which cytokines can modulate serotonergic signalling and influence emotional cognitive processing (Miller and Raison 2006; Zhu et al., 2006). Additionally, there is evidence that antidepressants have an effect on the production of proinflammatory cytokines, including TNF- α (Diamond et al., 2006). Table 2 contains results of different studies of the treatment effect on TNF- α . Heiser et al. (2008) found that the incubation of the platelets in vitro MDD patients with cortisol and dexamethazone at baseline resulted in an apparent increase in the secretion of TNF- α in the R group compared with HC while the values of the NR group did not differ from the data of the HC group in this respect. These data underscore the likelihood that this type of glycocorticoid actions may be present under special conditions despite the commonly assumed immunosupressive effects of the steroids. Therefore, it could well be possible that such a mechanism of glycocorticoids is at least partly responsible for the increased levels of this cytokine in MDD patients (Heiser et al., 2008; Lanquillon et al., 2000, Mikova et al., 2001). Heiser et al. (2008) believe that glycocorticoids may be involved in the psychopathology of MDD and the TNF- α system, supported by the correlation between these parameters in the responder subgroup of their study. They also found that the dynamics of the glycocorticoid receptor system is related to psychopathological normalization in the other system. In that study responders to antidepressive therapy showed more dynamic changes with the time of treatment under both basal and stimulated immune conditions. It suggests indeed that the dynamics of neuroendocrine-immune interactive system are related to a positive therapy response.

3. TNF- α in the augmentation phase with bupropion

Previous studies have reported that bupropion may lower the TNF- α level in various physical illnesses including Crohn's disease (Kast 2003). In this study bupropion was added to escitalopram in treatment-resistant patients, and the effect was not detectable. To our knowledge there were no previous studies on the effects of antidepressant augmentation on cytokines production. Hernandez et al. (2008) studied MDD patients during antidepressive treatment with SSRIs for 52 weeks and measured several cytokines at weeks 0, 5, 20, 36, and 52. They found that the changes in certain cytokine levels were not linear. The production of some cytokines changed only at the beginning of treatment (interferon- α), but IL-4, for example, increased from the baseline to week 20 and started to decrease after that. Unfortunately, TNF- α , IL-8 and sIL-2R were not assessed in their study. In this study, at the end of the augmentation phase the treatment had lasted for 18 weeks and the reactions of the immune system might be much faster at the beginning of the treatment. Among the limitations was the relatively small study group in the particular treatment phase.

4. IL-8

There were no differences in levels of IL-8 between the groups in our study. IL-8 is a chemokine produced by monocytes, macrophages, fibroblasts, keratinocytes, and endothelian cells after stimulation by IL-1 and TNF- α (Janeway et al., 2005). It includes several functions of human polymorphonuclear leukocytes, such as chemotaxis, release of granule components and respiratory burst (Mikova et al., 2001). There are only a few studies on IL-8 in major depression and even in animal studies this marker has been rarely measured. Simon and Song found elevated levels of IL-8 in major depression (Simon et al., 2008; Song et al., 1998). As in our study, Mikova et al. (2001) could not repeat that

11

finding. There is evidence that in the case of bipolar depression the levels of IL-8 are higher than in HC (O'Brien et al., 2006). Also, some papers have reported higher IL-8 levels in schizophrenia (Maes et al., 2002) or in negative symptoms of schizophrenia (Zhang et al., 2002). However, Song et al. (2007) reported lower levels of IL-8 in post-traumatic stress disorder compared with HC.

There is a study showing higher IL-8 concentrations in patients with fibromyalgia (Wang et al., 2009). Notably, pain is common in patients with major depression. Severity and duration of chronic pain was reported as directly proportional to severity of depression (Evans et al., 2005; Fishbain 2002). Some theories support the immunological link between pain and depression (Campbell et al., 2003). It is possible that IL-8 levels are related to pain symptoms of depression, but this hypothesis need to be tested.

There is no knowledge of the exact functions of IL-8 in the brain and its relationships with other cytokines and neurotransmitters. Thus, the results are rather difficult to interpret. There are two in vitro studies in healthy men and women showing a greater stimulated production of IL-8 correlating with higher BDI scores (Suarez et al., 2003, 2004), but this finding has not been replicated in depressed patients. Marsland et al. (2007) found among a middle-aged community volunteers a positive relationship between stimulated production of IL-8 and symptoms of depression and an inverse relationship between stimulated IL-8 production and perceived social support. Taken together, there are no affirmative data for a relationship between IL-8 and depression and broader assessments, including other immune markers, are necessary.

5. IL-8 in the escitalopram - treatment phase

The second study observed no changes in IL-8 production in MDD patients treated with escitalopram. There were only two previous studies evaluating associations of IL-8 with treatment outcomes in depression. Mikova et al. (2001) treated 14 patients with different antidepressants and did not find any effect on IL-8 levels. However, due to small sample size these findings remained inconclusive. O'Brien et al. (2007) assessed SSRI-resistant patients in comparison to HC. They measured cytokines once, upon determining treatment-resistant status. The higher production of pro-inflammatory cytokines IL-6 and TNF- α was detected in currently euthymic patients with previous SSRI-resistant depression with no differences in IL-8 levels.

6. IL-8 in the augmentation phase with bupropion

Production of IL-8 during 12 weeks of escitalopram treatment did not change significantly, but started to increase during bupropion augmentation. This may be explained by differences in the effects of various antidepressants on IL-8

synthesis. For example, Kast et al. (2003) found that bupropion decreased and mirtazapine increased the production of TNF- α . Lin et al. (2000) reported that moclobemide suppressed unstimulated production of IL-8 and the TNF- α effect in HC in vitro. However, these data can not be equally transferred to in vivo findings of MDD patients. There is also a possibility that the changes in IL-8 production need more time to become statistically significant. Only Hernandez et al. (2008) have evaluated levels of different cytokines repeatedly during 52 weeks of antidepressive treatment. Unfortunately, IL-8 was not measured in this study. More studies on the effects of augmentation with another antidepressant on immune markers' production are necessary to confirm the trends seen in our study.

7. sIL-2R

Few studies have focused on cytokine receptors in MDD, and the results are even more inconsistent than in cytokines (Table 1; rev by Kronfol and Remick 2000). Cells that express a functional receptor for a cytokine will respond to the presence of that cytokine. Cytokine receptors can also be found in soluble form. Usually, a soluble receptor for a specific cytokine can inhibit the biological activity of the cytokine by inhibiting the binding of the cytokine to its membrane-anchored receptor (TNF- α). However, on rare occasions sIL-R and the specific cytokine complex add to the activity of this cytokine (IL-6) (Kronfol and Remick 2000). There is evidence that sIL-2R appears to regulate the immune function by binding IL-2, and thereby neutralizes its cellular effects (Bien and Balcerska 2008; Levine et al., 2001).

IL-2 is the growth factor of T-cells, which is involved mainly in the proliferation and differentiation of T-cells but also activates B-cells and natural killer (NK) cells (Tayal and Kalra 2008). sIL-2R is produced by T-cells similarly to IL-2; thus, it is a marker of T-cell activity (Janeway et al., 2005). IL-2 is also identified in various brain regions, including hippocampus, neostriatum, and the cortex. IL-2 is produced by cells in the central nervous system including astrocytes and microglial cells (Levine et al., 2001). Elevated levels of sIL-2R have been observed in a variety of autoimmune and inflammatory diseases, including rheumatoid arthritis, multiple sclerosis, systemic lupus erythematosus, systemic sclerosis, myasthenia gravis, sarcoidosis, celiac disease and Crohn's disease, among others (Bien and Balcerska 2007). In major depression, serum IL-2 is often immeasurable because of low concentration (Maes et al., 1995; Klabusay et al., 2006), and usually sIL-2R is measured for assessing T-cell activity. There is an opinion that increased sIL-2R is an adaptation mechanism to increased IL-2 (Levine et al., 2001). Nevertheless, there are currently insufficient data to confirm this hypothesis. On the contrary, there is evidence that IL-2 accumulation in the serum of patients with unipolar and bipolar depression is strongly correlated with a decrease in sIL-2 serum levels (Androsova et al., 2001). An inverse relationship between IL-2 and sIL-2R levels has also been reported in studies in healthy individuals. Increased IL-2 and decreased sIL-2R levels, in addition to decreased IL-2R mRNA, were observed during examination periods in medical students compared with levels measured during low-stress periods (Mittwosh-Jaffe et al., 1995). Moreover, decreased IL-2 levels have been associated with negative emotions (Nakano et al., 1998) and economic stress (Anisman et al, 1999). sIL-2R levels were not measured in these studies.

Findings in patients with depression and healthy individuals have been, and in most studies only IL-2 levels, but not sIL-R, were measured. Some studies demonstrated lower levels of IL-2 in patients with depression compared with healthy controls (Hernandez et al., 2008; Kim et al, 2007; Pavon et al., 2006; Mendlovic et al., 1999). One of these studies included only inpatients with MDD (Pavon et al, 2006), while in the other studies, outpatients were included. In one study, no differences were observed between patients with MDD and healthy controls (Schlatter et al., 2004), although the cohort in this study was small, with 9 patients with depression and 9 healthy individuals. In contrast, higher IL-2 levels were observed in patients with depression compared with in healthy subjects in three separate studies (Simon et al., 2008; Sutcigil et al., 2007; Kim et al., 2008).

In this study decreased sIL-2R serum levels were found in MDD patients in full remission compared with MDD patients with a recurrent episode (trends for the first episode) and healthy controls. Maes et al. (1995) reported that sIL-2R concentrations appear to correlate with IL-2 secretion, and higher levels of sIL-2R may suggest an up-regulated production of IL-2. If so, then MDD patients in the remission phase have lower T-cell activation than other groups in the cohort of this study, The finding may suggest secondary adaptive changes in the immune system activity in the remission phase of MDD. The limitation of this study is a relatively small and heterogeneous FR group. The criteria for the duration of the remission have to be at least 2 months, but no further questions were asked to this effect. Therefore, one can not draw any conclusions about the time period when such changes appear.

On the other hand, Nunes et al. (2002) reported reduced levels of sIL-2R in moderate and severe depression, Kim et al. (2008) reported decreased IL-2 in suicidal MDD patients, Levine et al. (1999) reported lower cerebrospinal fluid (CRF) sIL-2R in MDD compared with healthy controls, and they failed to provide a good explanation to this effect. If lower levels of sIL-2R are related to MDD, it could well be that the finding of this study about lower levels of sIL-2R in the remission phase predicts future MDD episodes in those patients.

Increased serum levels of sIL-2R were observed in patients with MDD compared with healthy individuals in two previous papers (Maes et al., 1995; Seidel et al., 1995).

IL-2 is hypothesized to activate dopaminergic pathways (Levine et al., 2001; Müller and Ackenheil 1998). Study I found that sIL-2R concentrations associated with decreased activity and agitation items of the HAM-D scale, which are symptoms that the dopamine system is involved (Nutt et al., 2007).

8. sIL-2R in the escitalopram- treatment phase

There are many possible mechanisms of the SSRI effect on the activity of the immune system. Chronic antidepressant treatment re-establishes the feedback mechanisms of glycocorticoid receptors, and the activity of the HPA axis normalizes (Holsboer 2000). In addition, there is evidence that both noradrenalin and serotonin act as immuno-modulators. Thus, a functional increase in the activities of the neurotransmitter systems by effective antidepressants could contribute to the normalization of the immune function that occurs in depressed patients following effective treatment (Leonard 2001).

In study II, changes in sIL-2R levels followed different patterns in responders and non-responders to escitalopram. While an increase in sIL-2R is an adaptive immunosuppressive mechanism, the R group reacts immunosuppressively with an increased production of sIL-2R during the first four weeks, and production decreases after week 4. In the NR group the secretion of sIL-2R decreases between the baseline and week 4 and slightly increases after week 4. These findings, in combination with the results of Hernandez et al. (2008), suggest that the timepoint at which sIL-2R is measured is an important factor, and whether the concentration increases or decreases may depend on the duration of treatment.

Some other studies did not detect any changes in sIL-2R levels during antidepressive treatment when R and NR were analysed together. Subchronic use (~3 months') of fluoxetine or tricyclic antidepressants was not associated with changes in sIL-2R levels (measures at baseline and at treatment endpoint) in patients with major depression (Maes et al., 1995). In addition, treatment of patients with depression with clomipramine for 8 weeks did not have an effect on sIL-2R concentrations (Kagaya et al., 2001), and similar results were obtained using antidepressants from different classes, such as clomipramine (n=8), paroxetine (n=4), mianserin (n=1) and amitriptyline (n=1) (Mikova et al., 2001). Nor is there any difference between serum sIL-2R concentrations at baseline and week12 when R and NR were put together in this study. At the time of publication, Mikova et al. (2001) exhibited significantly higher serum IL-2R values in non-responders than in responders after treatment. However, the patient group that was assessed in this study was small (n=14), treatment was not standardized and, moreover, it included antidepressants from different classes.

9. sIL-2R in the augmentation phase with bupropion

In the augmentation phase the secretion of sIL-2R did not change; nor were there any differences between R and NR groups. It seems that differences in immune system reactions occur at the beginning of the treatment. Again, the timepoint of assessment that was discussed in the preceding section may play a role here. As this study is the first to explore the effect of bupropion augmentation on cytokine profiles, further studies are needed to clarify the reactions of the immune system in augmentation or treatment changes.

10. Anti-TPO and thyroid hormones

Thyroid autoimmunity is one of the aspects of immune system activation. The study IV found a somewhat higher frequency of baseline anti-TPO positive cases in non-responders to escitalopram monotherapy as compared with responders, suggesting that anti-TPO positivity may predict the treatment response to antidepressant medication. A larger study group may be needed to have significant statistical power. There was no relationship between thyroid hormones and treatment response.

Notably, anti-TPO positivity has often been associated with bipolar depression (Haggerty et al., 1997, Kupka et al., 2001) and also with unipolar depression (Carta et al., 2004). There is evidence that atypical depression is related to higher thyroid microsomal antibodies (synonymous with anti-TPO). In addition, the presence of anti-TPO during early pregnancy was associated with the development of postpartum depression (Kuijpens et al., 2001). There are several negative findings in unipolar depression (Engum et al., 2005; Fountoulakis et al., 2004; Haggerty et al., 1997, Horning et al., 1999). One hypothesis might be that anti-TPO positivity predicts bipolar disorder for depressed patients without previous manias. Undiagnosed bipolar disorder may be a possible reason for poorer improvement with antidepressants (Berk and Dodd 2005).

The association between presence of anti-TPO and immune system is demonstrated in bipolar patients, while the level of anti-TPO was negatively correlated with the serum level of sIL-2R (Padmos et al., 2004). The previous finding of study II is that the responders showed a lower baseline TNF- α level in comparison with non-responders or healthy subjects, whereas the two latter groups did not differ from each other. So, both findings suggest that immune system activation is involved in the treatment response in major depression.

Alteration of thyroid function has also been reported in depressed patients. Compared with controls, patients with depression showed lower basal serum evening TSH (Duval et al., 1999). However, this has not been a consistent finding (Frye et al., 1999; Fountoulakis et al., 2004; Pop et al., 1998). Gitlin et al. (2004) have demonstrated that low baseline TSH correlated strongly with

greater improvement in depressive symptoms assessed by change in HAM-D scores. Unfortunately, assessment of thyroid hormones was included to our study protocol later and we could assess hormones only once in patients with depression but not in healthy individuals. Previously, Sagud et al. (2002) demonstarted that 4-week treatment with sertraline increased plasma T3 levels in depressed patients. The baseline levels of thyroid hormones (TSH, T3 and T4) of patients with depression did not differ from the values in healthy controls in their study. These results are similar to the findings in study IV.

II. General discussion and future perspectives

We previously stated that about half of the patients whose depressive disorder is at least moderately severe, and those with melancholic features, have HPA-axis hyperactivity. However, not all melancholic patients have this kind of feature. Despite efforts over many years it is still not known how to clinically differentiate these patients from others or what the exact endophenotype of these depressive patients is. In addition, HPA hypofunction was found in a subgroup of depressed patients with hypersonnia, hyperphagia and lethargy or fatigue, commonly referred to as atypical depression (Antonijevic 2006; Levitan et al., 2002). Similarly, there is evidence that some of the depressive patients have changes in immune system activity, but the coexisting endophenotype is still unknown. There is some evidence that different subtypes of depression have different immune patterns, e.g. in melancholic versus non-melancholic depression (Kaestner et al., 2005; Rothermundt et al., 2001), but not all studies confirmed this relationship (Margues-Deak et al., 2007). Unfortunately, these studies investigated different cytokines making direct comparisons not feasible. This formed the basis for our evaluation of the relationships between single depressive symptoms and immune system markers as with study I. The available data are rather inconsistent and controversial, and do not allow clear conclusions to be drawn regarding the involvement of cytokines in depression. Most of the previous studies were limited by small or heterogeneous samples. whereas additional bias might be caused by concomitant use of antidepressive medications during the measurement of cytokines. Moreover, only a few of the previous studies have addressed the association between cytokines and specific symptoms of depression or the different stages of clinical course. Therefore, future studies need very homogenous cohorts; enough patients in each study group, including remission group. In addition, different studies assess different cytokines with different methods. It would be highly useful to assess the concentration of several cytokines and their receptors in one study so as to evaluate the co-effects. Moreover, there is evidence that cytokines are influenced by several factors, such as biological rhythms, circadian rhythms, smoking habits, previous food intake, and physical activity (Gokhale et al., 2007; Haack et al., 2004; Majde and Krueger 2005; Reichenberg et al., 2002).

There is no much knowledge about the influence of factors such as climate, race, gender and seasons. As concentrations of some (TNF-receptor), but not all (IL-2 receptor), immune markers change during the day, repeated measurements would be helpful (Haack et al., 2004). Measurement of certain cytokines, including TNF- α and sIL-2R, may predict response to treatment, but the association is likely to depend on the specific action of the antidepressant used. Clinical trials with standardized treatment with different classes of antidepressant should help to determine whether immune system reactions are associated with distinctive neuropharmacological profiles of antidepressant medication. Furthermore, as demonstrated by Hernandez et al. (2008), longer treatment periods with regular clinical and biomarker assessments are necessary to make further progress in this field. A framework of the effects of immune system in the pathogenesis of depression is presented in Figure 1.

Our studies were limited to measuring only three immune markers: TNF- α , sIL-2R and IL-8. In future studies more complex laboratory analyses would be needed. Using these three markers we could see that being drug-naive or not, could strongly influence results for an unknown time-period. More to the point, previous antidepressive treatment lowers the levels of TNF- α . There was no immune activation in depressed patients in our Estonian study cohort. However, lower levels of baseline TNF- α predicted better treatment response to escitalopram. Different patterns of change in the sIL-2R levels occurred in responders and non-responders which affirms the need for repeated measurements during a longer treatment period.

The weak result that anti-TPO positivity could negatively influence the response to escitalopram needs to be repeated with a larger study cohort because of the low prevalence rate. Thyroid hormones are known to be related to mood symptoms. However, repeated assessment during the treatment with different antidepressants would be better for ascertaining the associations between thyroid function and depressive states.

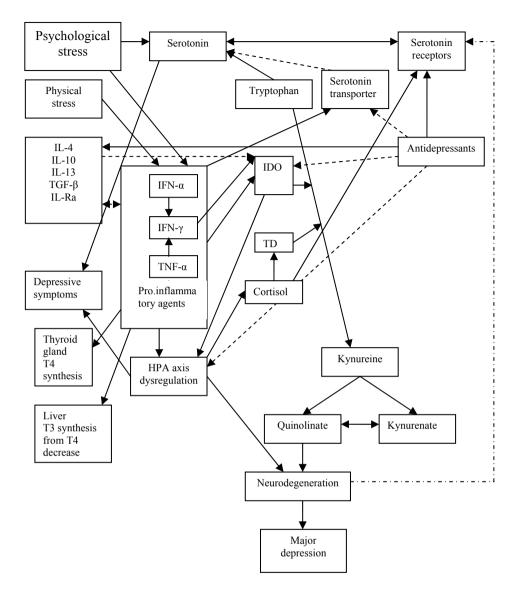


Figure 1. Immune system's effects in the pathogenesis of depression.

CONCLUSIONS

- 1. The levels of TNF- α were lower in currently depressed subjects compared with euthymic subjects in the study cohort. MDD patients with previous antidepressive treatment had significantly lower levels of TNF- α than drug-naïve patients and HC.
- 2. There was a positive correlation between TNF- α (but not sIL-2R) and the HAM-D total score in currently depressed subjects.
- 3. The levels of TNF- α were positively related to HAM-D items of decreased activity and agitation. The levels of sIL-2R were positively associated with HAM-D items of decreased activity and suicidality.
- 4. There were different patterns of changes in the levels of sIL-2R in responders and non-responders to escitalopram treatment: The concentrations of sIL-2R decreased later in non-responders than in responders. Treatment with escitalopram had no significant effect on the levels of IL-8 and TNF- α .
- 5. Augmentation of escitalopram treatment with bupropion caused a significant increase in IL-8 serum concentrations during 6 weeks of augmentation therapy. There was no effect on the levels of sIL-2R and TNF- α .
- 6. The lower baseline TNF- α level was found in the responder group in the escitalopram treatment phase. More specifically, male non-responders had higher levels of TNF- α than male responders or female responders and non-responders.
- 7. There was a trend for higher frequency of baseline anti-TPO positive cases in female non-responders to escitalopram monotherapy as compared with responders. There were no significant differences in the levels of thyroid hormones (particularly, total T3, free T3, free T4, and TSH) between female responders and non-responders.

REFERENCES

- Abraham G, Milev R, Lawson JS. T3 augmentation of SSRI resistant depression. J Affect Disord 2006; 91: 211–215.
- 2. Agid O, Lerer B. Algorithm-based treatment of major depression in an outpatient clinic: clinical correlates of response to a specific serotonin reuptake inhibitor and to triiodothyronine augmentation. Int J Neuropsychopharmacol 2003; 6: 41–49.
- Aluoja A, Leinsalu M, Shlik J, Vasar v, Luuk K. Symptoms of depression in the Estonian population: Prevalence, sociodemographic correlates and social adjustment. J Affect Disord 2004; 78: 27–35.
- Anderson IM, Parry-Billings M, Newsholme EA, Poortmans JR, Cowen PJ Decreased plasma tryptophan concentration in major depression: relationship to melancholia and weight loss. J Affect Disord 1990; 20:185–91.
- Andrade L, Caraveo-Anduaga JJ, Berglund P, Bijl RV, De Graaf R, Vollebergh W, Dragomirecka E, Kohn R, Keller M, Kessler RC, Kawakami N, Kiliç C, Offord D, Ustun TB, Wittchen HU. The epidemiology of major depressive episodes: results from the International Consortium of Psychiatric Epidemiology (ICPE) Surveys. Int J Methods Psychiatr Res 2003; 12: 3–21.
- 6. Androsova LV, Kushner SG, Abramova LI, Oleichik IV, Egorova MIU: Interleukine level in endogeneous depression. Zh Nevrol Psikhiatr Im S S Korsakova 2001; 101: 45–48 (Abstract).
- 7. Angst J, Angst F, Stassen HH. Suicide risk in patients with major depressive disorder. J Clin Psychiatry 1999; 60 Suppl 2:57–62.
- Angst J, Gamma A, Sellaro R, Zhang H, Merikangas K. Toward validation of atypical depression in the community: Results of the Zurich cohort study. J Affect Disord 2002; 72: 125–138.
- 9. Angst J, Merikangas K. The depressive spectrum: Diagnostic classification and course. J Affect Disord 1997; 45: 31–40.
- 10. Anisman H, Merali Z. Cytokines, stress, and depressive illness. Brain Behav Immun 2002; 16: 513–524.
- 11. Anisman H, Merali Z, Hayley S. Neurotransmitter, peptide and cytokine in relation to depressive disorder: Comorbidity between depression and neuro-degenerative disorders. Prog Neurobiology 2008; 85: 1–74.
- 12. Anisman H, Ravindran AV, Griffiths J, Merali Z.: Endocrine and cytokine correlates of major depression and dysthymia with typical or atypical features. Mol Psychiatry 1999; 4: 182–188.
- 13. Antonijevic IA. Depressive disorders is it time to endorse different pathophysiologies? Psychoneuroendocrinology 2006; 31: 1–15.
- 14. APA. Diagnostic and Statistical Manual of Mental Disorders (DSM-IV) 4th edition. 1994; Washington, DC, American Psychiatric Association.
- 15. Aronson R, Offman HJ, Joffe RT, Naylor CD. Triiodothyronine augmentation in the treatment of refractory depression. A meta-analysis. Arch Gen Psychiatry 1996; 53: 842–848.
- 16. Basterzi AD, Aydemir Ç, Kisa C, Aksaray S, Tuzer V, Yazici K, Göka E. IL-6 levels decrease with SSRI treatment in patients with major depression. Hum Psychopharmacol Clin Exp 2005; 20: 473–476.

- 17. Bauer M, Goetz T, Glenn T, Whybrow PC. The thyroid brain interaction in thyroid disorders and mood disorders. Journal of Neuroendocrinology 2008; 20: 1101–1114.
- Bauer ME, Papadopoulos A, Poon L, Perks P, Lightman SL, Checkley S, Shanks N. Altered glucocorticoid immunoregulation in treatment resistant depression. Psychoneuroendocrinology 2003; 28: 49–65.
- 19. Berk M, Dodd S. Are treatment emergent suicidality and decreased response to antidepressants in younger patients due to bipolar disorder being misdiagnosed as unipolar depression? Med Hypotheses 2005; 65:39–43.
- 20. Bien E, Balcerska A. Serum soluble interleukin 2 receptor α in human cancer of adults and children: a review. Biomarkers 2007; 13: 1–26.
- Bland RC. Epidemiology of affective disorders: a review. Can J Psychiatry 1997; 42: 367–377.
- Brambilla F, Monteleone P, Maj M. Interleukin-1β and tumor necrosis factor-α in children with major depressive disorder or dysthymia. J Affect Disord 2004; 78: 273–277.
- 23. Brustolim, D, Ribeiro-dos-Santos R, Kast RE, Altschuler EL, Soares MBP. A new chapter opens in anti-inflammatory treatments: the antidepressant bupropion lowers production of tumor necrosis factor-alpha and interferon-gamma in mice. Int Immunopharmacol 2006; 6: 903–907.
- 24. Campbell LC, Clauw DJ, Keefe FJ. Persistent pain and depression: a biopsychosocial perspective. Biol Psychiatry 2003; 54: 399–409.
- 25. Capuron L, Miller AH. Cytokines and Psychopathology: lessons from interferonα. Biol Psychiatry 2004; 56: 819–824.
- Capuron L, Ravaud A, Miller A, Dantzer R. Baseline mood and psychosocial symptoms during interleukin-2 and/or interferon-alpha cancer therapy. Brain Behav Immun 2004; 18: 205–213.
- 27. Carta MG, Loviselli A, Hardoy MC, Massa S, Cadeddu M, Sardu C, Carpiniello B, Dell'Osso L, Mariotti S. The link between thyroid autoimmunity (antithyroid peroxidase autoantibodies) with anxiety and mood disorders in the community: a field of interest for public health in the future. BMC Psychiatry 2004; 4:25.
- 28. Chanrashekara S, Jayashree K, Veeranna HB, Vadiraj HS, Ramesh MN, Shobha A, Sarvanan Y, Vikram YK. Effects of anxiety on TNF-alpha levels during psychological stress. J Psychosom Res. 2007; 63: 65–69.
- 29. Chen PS, McQuoid DR, Payne ME, Steffens DC. White matter and subcortical gray matter lesion volume changes and late-life depression outcome: 4-year magnetic resonance imaging study. Int Psychogeriatr 2006; 18: 445–456.
- 30. Chueire VB, Romaldini JH, Ward LS. Subclinical hypothyroidism increase the risk for depression in the elderly. Arch Gerontol Geriatr 2007; 44: 21–28.
- 31. Connor TJ, Leonard BE. Depression, stress and immunological activation: the role of cytokines in depressive disorders. Life Sci 1998; 62: 583–606.
- Cowen PJ. Neurobiology of depression. In: Griez EJL, Faravelli C, Nutt DJ, Zohar J. (Eds.) Mood disorders: clinical management and research issues. John Wiley & Sons Ltd, England, 2005; pp. 191–210.
- 33. Cyranowski JM, Marsland AL, Bromberger JT, Whiteside TL, Chang Y, Matthews KA. Depressive symptoms and production of proinflammatory cytokines by peripheral blood mononuclear cells stimulated in vitro. Brain Behav Immun 2007; 21: 229–237.

- 34. De Beaurepaire R. Questions raised by the cytokine hypothesis of depression. Brain Behav Immun 2002; 16: 610–617.
- Diamond M, Kelly JP, Connor TJ. Antidepressants suppress production of the Th1 cytokine interferon-γ, independent of monoamine transporter blockade. Eur Neuropsychopharmacol 2006; 16: 481–490.
- 36. Duval F, Mokrani MC, Bailey P, Correa H, Diep TS, Crocq MA, Macher JP. Thyroid axis activity and serotonin function in major depressive episode. Psychoneuroendocrinology 1999; 24: 695–712.
- Duval F, Mokrani MC, Crocq MA, Jautz M, Bailey P, Diep TS, Macher JP. Effect of antidepressant medication on morning and evening thyroid function tests during depressive episode. Arch Gen Psychiatry 1996; 53: 833–840.
- 38. Elhwuegi AS. Central monoamines and their role in major depression. Prog Neuropsychopharmacol Biol Psychiatry 2004; 28: 435–451.
- 39. Engum A, Bjøro T, Mykletun A, Dahl AA. Thyroid autoimmunity, depression and anxiety; are there any connections? An epidemiological study of a large population. Journal Psychosom Res 2005; 59: 263–268.
- 40. Evans DL, Charney DS, Lewis L, Golden RN, Gorman JM, Krishnan KR, Nemeroff CB, Bremner JD, Carney RM, Coyne JC, Delong MR, Frasure-Smith N, Glassman AH, Gold PW, Grant I, Gwyther L, Ironson G, Johnson RL, Kanner AM, Katon WJ, Kaufmann PG, Keefe FJ, Ketter T, Laughren TP, Leserman J, Lyketsos CG, McDonald WM, McEwen BS, Miller AH, Musselman D, O'Connor C, Petitto JM, Pollock BG, Robinson RG, Roose SP, Rowland J, Sheline Y, Sheps DS, Simon G, Spiegel D, Stunkard A, Sunderland T, Tibbits P Jr, Valvo WJ. Mood disorders in the medically ill: scientific review and recommendations. Biol Psychiatry 2005; 58: 175–189.
- Faravelli C, Ravaldi C, Truglia E. Unipolar depression. In: Griez EJL, Faravelli C, Nutt DJ, Zohar J. (Eds.) Mood disorders: clinical management and research issues. John Wiley & Sons Ltd, England, 2005; pp. 79–101.
- Fishbain DA. The pain-depression relationship. Psychosomatics 2002; 43: 341– 342.
- 43. Fountoulakis KN, Iacovides A, Grammaticos P, St Kaprinis G, Bech P. Thyroid function in clinical subtypes of major depression: an exploratory study. BMC Psychiatry 2004, 4:6.
- 44. Frodl T, Meisenzahl EM, Zetzsche T, Born C, Groll C, Jäger M, Leinsinger G, Bottlender R, Hahn K, Möller HJ. Hippocampal changes in patients with a first episode of major depression. Am J Psychiatry 2002; 159: 1112–1118.
- 45. Frommberger UH, Bauer J, Haselbauer P, Fräulin A, Riemann D, Berger M. Interleukin-6-(IL-6) plasma levels in depression and schizophrenia: comparison between the acute state and after remission. Eur Arch Psychiatry Clin Neurosci. 1997; 247: 228–233.
- 46. Frye MA, Dunn RT, Gary KA, Kimbrell TA, Callahan AM, Luckenbaugh DA, Corá-Locatelli G, Vanderham E, Winokur A, Post RM. Lack of correlation between cerebrospinal fluid thyrotropin-releasing hormone (TRH) and TRHstimulated thyroid-stimulating hormone in patients with depression. Biol Psychiatry. 1999; 15; 45:1049–1052.
- 47. Furlong RA, Coleman TA, Ho L, Rubinsztein JS, Walsh C, Paykel ES, Rubinsztein DC. No association of a functional polymorphism in the dopamine D2 receptor promoter region with bipolar or unipolar affective disorders. Am J Med Genet 1998a; 81: 385–387.

- Furlong RA, Ho L, Rubinsztein JS, Walsh C, Paykel ES, Rubinsztein DC. No association of the tryptophan hydroxylase gene with bipolar affective disorder, unipolar affective disorder, or suicidal behaviour in major affective disorder. Am J Med Genet 1998b; 81: 245–247.
- 49. Furlong RA, Rubinsztein JS, Ho L, Walsh C, Coleman TA, Muir WJ, Paykel ES, Blackwood DHR, Rubinsztein DC. Analysis and metaanalysis of two polymorphisms within the tyrosine hydroxylase gene in bipolar and unipolar affective disorders. Am J Med Genet 1999; 88: 88–94.
- Gitlin M, Altshuler LL, Frye MA, Suri R, Huynh EL, Fairbanks L, Bauer M, Korenman S. Peripherial thyroid hormones and response to selective serotonin reuptake inhibitors. J Psychiatry Neurosci 2004; 29: 383–386.
- 51. Gokhale R, Chanddrashekara S, Vasanthakumar KC. Cytokine response to strenuous exercise in athletes and non-athletes an adaptive response. Cytokine 2007; 40: 123–127.
- 52. Haack M, Pollmächer T, Mullington M. Diurnal and sleep-wake dependent variations of soluble TNF- and IL-2 receptors in healthy volunteers. Brain Behav Immun 2004; 18:361–367.
- 53. Haggerty JJ Jr, Silva SG, Marquardt M, Mason GA, Chang HY, Evans DL, Golden RN, Pederssen C. Prevalence of antithyroid antibodies in mood disorders. Depress Anxiety 1997; 5: 91–96.
- 54. Hayley S, Poulter MO, Merali Z, Anisman H. The pathogenesis of clinical depression: stressor- and cytokine-induced alterations of neuroplasticity. Neuroscience 2005; 135: 659–678.
- 55. Heiser P, Lanquillon S, Krieg J-C, Vedder H. Differential modulation of cytokine production in major depressive disorder by cortisol and dexamethasone. Eur Neuropsychopharmacol 2008; 18: 860–870.
- Hernández ME, Mendieta D, Martínez-Fong D, Loría F, Moreno J, Estrada I, Bojalil r, Pavón L. Variations in circulating cytokine levels during 52 week course of treatment with SSRI for major depressive disorder. Eur Neuropsychopharmacol 2008; 18:917–924.
- 57. Himmerich H, Binder EB, Künzel HE, Schuld A, Lucae S, Uhr M, Pollmächer T, Holsboer F, Ising M. Successful antidepressant therapy restores the disturbed interplay between TNF-α system and HPA axis. Biol psychiatry 2006; 60: 882–888.
- 58. Ho LW, Furlong RA, Rubinsztein JS, Walsh C, Paykel ES, Rubinsztein DC. Genetic associations with clinical characteristics in bipolar affective disorder and recurrent unipolar depressive disorder. Am J Med Genet 2000; 96:36–42.
- 59. Holmans P, Zubenko GS, Crowe RR, DePaulo Jr JR, Scheftner WA, Weissman Mm, Zubenko WN, Boutelle S, Murphy-Eberenz K, MacKinnon D, McInnis MG, Marta DH, Adams P, Knowles JA, Gladis M, Thomas J, Chellis J, Miller E, and Levinson DF. Genomewide significant linkage to recurrent, early-onset major depressive disorder on chromosome 15q. Am J Hum Genet 2004; 74: 1154–1167.
- 60. Holsboer F. The corticosteroid receptor hypothesis of depression. Neuropsychopharmacology 2000; 23: 477–501. Review
- 61. Hong, CJ, Yu, YW, Chen, TJ, Tsai, SJ. Interleukin-6 genetic polymorphism and Chinese major depression. Neuropsychobiology 2005; 52: 202–205.
- 62. Horing M, Amsterdam JD, Kamoun M, Goodman DBP. Autoantibody disturbances in affective disorders: a function of age and gender. J Affect Disord 1999; 55: 29–37.

- 63. Ialongo N, McCreary BK, Pearson JL, Koenig AL, Schmidt NB, Poduska J, Kellam SG. Major depressive disorder in a population of urban, African-American young adults: Prevalence, correlates, comorbidity and unmet mental health service need. J Affect Disord 2004; 79: 127–136.
- Jacobi F, Rosi S, Faravelli C, Goodwin R, Arbabzaden-Bouchez S, Lépine JP. Epidemiology of mood disorders. In: Griez EJL, Faravelli C, Nutt DJ, Zohar J. (Eds.) Mood disorders: clinical management and research issues. John Wiley & Sons Ltd, England, 2005; pp. 8–12.
- Janeway CA, Travers P, Walport M, Shlomchik MJ. Immunobiology: the immune system in health and disease 6th ed. Garland science Publishing, Taylor & Francis Group, Oxon, UK; 2005; pp 337–338.
- 66. Joffe RT, Levitt AJ. Major depression and subclinical (grade 2) hypothyroidism. Psychoneuroendocrinology 1992; 17:215–221.
- 67. Joffe RT, Marriott M. Thyroid hormone levels and recurrence of major depression. Am J Psychiatry 2000; 157: 1689–1691.
- Johansson C, Jansson M, Linnér L, Yuan QP, Pedersen ML, Blackwood D, Barden N, Kelsoe J, Schalling M. Genetics of affective disorders. Eur Neuropsychiatr 2001; 11: 385–394.
- 69. Jun, TY, Pae, CU, Chae, JH, Bahk, WM, Kim, KS, Han, H. Report on IL-10 gene polymorphism at position –819 for major depression and schizophrenia in Korean population. Psychiatry Clin Neurosci 2002; 56: 177–180.
- Jun, TY, Pae, CU, Hoon, H, Chae, JH, Bahk, WM, Kim, KS, Serretti, A. Possible association between -G308A tumour necrosis factor-alpha gene polymorphism and major depressive disorder in the Korean population. Psychiatr Genet 2003; 13: 179–181.
- Kaestner F, Hettich M, Peters M, Sibrowaki W, Hetzel G, Ponath G, Arolt V, Cassens U, Rothermundt M. Different activation patterns of proinflammatory cytokines in melancholic and non-melancholic major depression are associated with HPA axis activity. J Affect Disord 2005; 87: 305–311.
- Kagaya A, Kugaya A, Takebayashi M, Fukue-Saeki M, Saeki T, Yamawaki S, Uchitomi Y. Plasma concentrations of interleukin-1beta, interleukin-6, soluble interleukin-2 receptor and tumor necrosis factor alpha of depressed patients in Japan. Neuropsychobiology. 2001; 43: 59–62.
- Kast RE. Anti- and pro-inflammatry considerations in antidepressant use during medical illness: bupropion lowers and mirtazapine increases circulating tumor necrosis factor-alpha levels. Gen Hosp Psychiatry 2003; 25: 495–496.
- Kessler RC, Berglund P, Demler O, Jin R, Koretz D, Merikangas KR, Rush AJ, Walters EE, Wang PS. The Epidemiology of Major Depressive Disorder. Results from the National Comorbidity Survey Replication (NCS-R). JAMA 2003; 289: 3095–3105.
- Kessler RC, McGonagle KA, Swartz M, Blazer DG, Nelson CB. Sex and depression in the National Comorbidity Survey I: Lifetime prevalence, chronicity and recurrence. J Affect Disord 1993; 29: 85–96.
- Kim YK, Na KS, Shin KH, Jung HY, Choi SH, Kim JB. Cytokine imbalance in the pathophysiology of major depressive disorder. Prog in Neuro-Psychopharmacol & Biol Psychiatry 2007; 31: 1044–1053.
- Kim YK, Lee SW, Kim SH, Shim SH, Han SW, Choi SH, Lee BH. Differences in cytokines between non-suicidal patients and suicidal patients in major depression. Prog in Neuro-Psychopharmacol & Biol Psychoatry 2008; 32: 356–361.

- Klabusay M, Kohutova V, Coupek P, Nenickova M, Tesarova E. Simultaneous analysis of cytokines and co-stimulatory molecules concentrations by ELISA technique and of probabilities of measurable concentrations of interleukins IL-2, IL-4, IL-5, IL-6, CXCL8 (IL-8), IL-10, IL-13 occurring in plasma of healthy blood donors. Mediators Inflamm. 2006: 1–7.
- 79. Kronfol Z, Remick DG. Cytokines and the brain: implications for clinical psychiatry. Am J Psychiatry 2006; 157: 683–694.
- Kubera M, Kenis G, Bosmans E, Kajta M, Basta-Kaim A, Scharpe S, Budziszewska B, Maes, M. Stimulatory effect of antidepressants on the production of IL-6. Int Immunopharmacol 2004; 4:185–192.
- 81. Kubera M, Kenis G, Bosmans E, Scharpe S, Maes M. Effects of serotonin and serotonergic agonists and antagonists on the production of interferon-gamma and interleukin-10. Neuropsychopharmacol 2000a; 23: 89–98.
- Kubera M, Kenis G, Bosmans E, Zieba A, Dubek D, Nowak G, Maes M. Plasma levels of interleukin-6, interleukin-10, and interleukin-1 receptor antagonist in depression. Comparison between the acute state and after remission. Pol J Pharmacol 2000b; 52: 237–241 (abstract).
- Kubera M, Lin AH, Kenis G, Bosmans E, van Bockstaele D, Maes M. Antiinflammatory effects of antidepressants through supression of the interferongamma/interleukin-10 production ratio. J Clin Psychopharmacol 2001a; 21: 199– 206.
- Kubera M, Maes M, Holan V, Basta-Kaim A, Roman A, Shani J. Prolonged desipramine treatment increase the production of interlekin-10, an antiinflammatory cytokine, in C57BL/6 mice subjected to the chronic mild stress model of depression. J Affecy Disord 2001b; 63: 171–178.
- Kubera M, Symbirtsev A, Basta-Kaim A, Roman A, Papp M, Claesson M. Effect on chronic treatment with imipramine on IL-1 and IL-2 production by splenocytes obtained from rats subjected to chronic mild stress model of depression. Pol J Pharmacol 1996; 48: 503–506 (abstract).
- 86. Thyroid peroxidase antibodies during gestation are a marker for subsequent depression postpartum. Kuijpens JL, Vader HL, Drexhage HA, Wiersinga WM, van Son MJ, Pop VJ. Eur J Endocrinol. 2001; 145: 579–584.
- Kupka RW, Nolen WA, Post RM, McElroy SL, Altshuler LL, Denicoff KD, Frye MA, Keck, Jr., PE, Leverich GS, Rush AJ, Suppes T, Pollio C, Drexhage HA. High rate of autoimmune thyroiditis in bipolar disorder: lack of association with lithium exposure. Biol Psychoatry 2002; 51:305–311.
- Kõks S, Nikopensius T, Koido K, Maron M, Altmäe S, Heinaste E, Vabrit K, Tammekivi V, Hallast P, Kurg A, Shlik J, Vasar V, Metspalu A, Vasar E. Analysis of SNP profiles in patients with major depressive disorder. Int J Neuropsychopharmacol 2006; 9:167–74.
- 89. Lanquillon S, Krieg JC, Bening-Abu-Shach U, Vedder H. Cutokine production and treatment response in major depressive disorder. Neuropsychopharmacol 2000; 22: 370–379.
- 90. Lee KM, Kim YK. The role of IL-12 and TGF-β1 in the pathophysiology of major depressive disorder. Int Immunopharmacol 2006; 6: 1298–1304.
- 91. Lekman M, Laje G, Charney D, Rush AJ, Wilson AF, Soranet AJM, Lipsky R, Wisnewski SR, Manji H, McMahon FJ, Paddock S. The FKBP5-gene in depression and Treatment response an association study in the sequenced

treatment alternatives to relieve depression (STAR*D) cohort. Biol Psychiatry 2008; 63:1103–1110.

- 92. Lenczowski MJ, Bluthé RM, Roth J, Rees GS, Rushforth DA, van Dam AM, Tilders FJ, Dantzer R, Rothwell NJ, Luheshi GN. Central administration of rat IL-6 induces HPA activation and fever but not sickness behavior in rats. Am J Physiol. 1999; 276: R652–658.
- 93. Leo R, Di Lorenzo G, Tesauro M, Razzini C, Forleo GB, Chiricolo G, Cola C, Zanasi M, Troisi A, Siracusano A, Lauro R, Romeo F. Association between enhanced soluble CD40 ligand ans proinflammatory and prothtombotic states in major depressive disorder. Pilot observation on the effects of selective serotonin reuptake inhibitor therapy. J Clin Psychiatry 2006; 67: 1760–1766.
- 94. Leonard BE. Changes in the immune system in depression and dementia: causal or co-incidental effects? Int J Devl Neuroscience 2001; 19: 305–312.
- 95. Levine J, Chengappa KNR, Gershon S, Drevets W. Differentiating primary pathophysiologic from secondary adaptational processes. Depress Anxiety 2001; 14: 105–111.
- Levitan RD, Vaccarino FJ, Brown GM, Kennedy SH. Low-dose dexamethasone challenge in women with atypical major depression: pilot study. J Psychiatry Neurosci 2002; 27: 47–51.
- Lin A, Song C, Kenis G, Bosmans E, De Jongh R, Scharpé S, Maes M. The in vitro immunosuppressive effects of moclobemide in healthy volunteers. J Affect Disord 2000; 58: 69–74.
- Maes M, Bosmans E, De Jongh R, Kenis G, Vandoolaeghe E, Neels H. Increased serum IL-6 and IL-1 receptor antagonists in major depression and treatment resistany depression. Cytokine 1997; 11: 853–858.
- Maes M, Bosmans E, Meltzer HY, Scharpé S, Suy E. Interleukin-1β: a putative mediator of HPA axis hyperactivity in major depression? Am J Psychiatry 1993; 150: 1189–1193.
- 100. Maes M, Kenis G, Kubera M, De Baets M, Steinbusch H, Bosmans E. The negative immunoregulatory effects of fluoxetine in relation to the cAMP-dependent PKA pathway. Int Immunopharmacol 2005; 5: 609–618.
- 101. Maes M, Meltzer HY, Bosmans E, Bergmans R, Vandoolaeghe E, Ranjan R, Desnyder R. Increased plasma concentrations of interleukin-6, soluble interleukin-6, soluble interleukin-2 and transferrin receptor in major depression. J Affect Disord 1995; 18: 301–309.
- 102. Maes M, Song C, Lin AH, Bonaccorso S, Kenis G, De Jongh R, Bosmans E, Scharpé S. Negative immunoregulatory effects of antidepressants: inhibition of interferon- γ and stimulation of interleukin-10 secretion. Neuropsychopharmacol 1999; 20: 370–379.
- 103. Maes M, Song C, Lin A, Gabriels L, De Jongh R, Van Gastel A, Kenis D, Bosmans E, De Meester I, Benoyt I, Neels H, Janca A, Scharpé S, Smith RS. The effects of psychological stress on the immune system in humans. Biol Psychiatry 1997a; 42:1S–297S.
- 104. Majde JA, Krueger JM. Links between the innate immune system and sleep. J Allergy Clin Immunol. 2005; 116: 1188–1198.
- Marques-Deak AH, Neto FL, Dominguez WV, Solis AC, Kurgant D, Sato F, Ross JM, Prado EBA. Cytokine profiles in women with different subtypes of major depressive disorder. J Psychiatr Res 2007; 41: 152–159.

- 106. Marsland AL, Sathanoori R, Muldoon MF, Manuck SB. Stimulated production of interleukin-8 covaries with psychosocial risk factors for inflammatory disease among middle-aged community volunteers. Brain Behav Immun 2007; 21: 218– 228.
- 107. McCulley, MC, Day, IN, Holmes, C. Association between interleukin 1-beta promoter (-511) polymorphism and depressive symptoms in Alzheimer's disease. Am J Med Genet B Neuropsychiatr Genet 2004; 124: 50–53.
- Mendlovic S, Mozes E, Eilat E, Doron A, Lereya J, Zakuth V, Spirer Z: Immune activation in non-treated suicidal major depression. Immunol Lett 1999; 67: 105– 108.
- 109. Merikangas KR, Chakravarti A, Moldin SO, Araj H, Blangero JC, Burmeister M, Crabbe J Jr, Depaulo JR Jr, Foulks E, Freimer NB, Koretz DS, Lichtenstein W, Mignot E, Reiss AL, Risch NJ, Takahashi JS. Future of genetics of mood disorders research. Biol Psychiatry 2002; 52: 457–77.
- 110. Mikova O, Yakimova R, Bosmans E, Kenis G, Maes M. Increased serum tumor necrosis factor alpha concentrations in major depression and multiple sclerosis. Eur Neuropsychopharmacol 2001; 11: 203–208.
- 111. Miller AH, Capuron L, Raison CL. Immunologic influence on emotion regulation. Clin Neurosci Research 2005; 4: 325–333.
- 112. Miller A, Raison CL. Cytokines, p38 MAP kinase and the pathophysiology of depression. Neuropsychopharmacology 2006; 31: 2089–2090.
- 113. Mittwoch-Jaffe T, Shalit F, Srendi B, Yehuda S: Modification of cytokine secretion following mild emotional stimuli. Neuroreport 1995; 6: 789–792.
- 114. Murphy GE. Why women are less likely than men to commit suicide. Comprehensive Psychiatry 1998; 39: 165–175.
- 115. Mössner R, Daniel S, Schmitt A, Albert D, Lesch KP. Modulation of serotonin transporter function by interleukin-4. Life Sci. 2001; 68: 873–880.
- Müller N, Ackenheil M. Psychoneuroimmunology and the cytokine action in the CNS: implications for psychiatric disorders. Progress Neuro-Psychopharmacology & Biological Psychiatry 1998; 22: 1–33.
- 117. Myint AM, Kim YK. Cytokine-serotonin interaction through IDO: a neurodegeneration hypothesis of depression. Med Hypotheses 2003; 61: 519–525.
- 118. Myint AM, Leonard BE, Steinbusch HWM, Kim YK. Th1, Th2, and Th3 cytokine alterations in major depression. J Affect Disord 2005; 88: 167–173.
- Nakano Y, Nakamura S, Hirata M, Harada K, Ando K, Tabuchi T, Matunaga I, Oda H: Immune function and lifestyle of taxi drivers in Japan. Ind Health 1998; 36: 32–39.
- 120. Narita K, Murata T, Takahashi T, Kosaka H, Omata N, Wada Y. Plasma levels of adiponectin and tumor necrosis factor-alpha in patients with remitted major depression receiving long-term maintenance antidepressant therapy. Prog in Neuropsychopharmacol & Biol Psychiatry 2006; 30: 1159–1162.
- 121. Neff CD, Abkevich V, Packer JCL, Chen Y, Potter J, Riley R, Davenport C, Warren JDeG, Jammulapati S, Bhathena A, Choi WS, Kroeger PE, Metzger RE, GutinA, Skolnick MH, Shattuck D, Katz DA. Evidence for *HTR1A* and *LHPP* as interacting genetic risk factors in major depression. Mol Psychiatry 2009; 14:621– 630.
- 122. Nemeroff CB, Vale W. The neurobiology of depression: inroads to treatment and new drug discovery. J Clin Psychiatry 2005; 66:5–13.

- 123. Nunes SOV, Reiche EMV, Morimoto HK, Matsuo T, Itano EN, Xavier ECD, Yamashita CM, Vieira VR, Menoli AV, Vilva SS, Costa FB, Reiche FV, Silva FLV, Kaminami MS. Immune and hormonal activity in adults suffering from depression. Braz J Med Biol Res 2002; 35: 581–587.
- 124. Nutt D, Demyttenaere K, Janka Z, Aarre T, Bourin M, Canonico PL, Carrasco JL, Stahl S. The other face of depression reduced positive affect: the role of catecholamines in causation and cure. J Psychopharmacol 2007; 21: 461–471.
- O'Brien SM, Scott LV, Dinan TG. Cytokines: abnormalities in major depression and implications for pharmacological treatment. Hum Psychopharmacol Clin Exp 2004; 19: 397–403.
- 126. O'Brien SM, Scully P, Fitzerald P, Scott LV, Dinan TG. Plasma cytokine profiles in depressed patients who fail to respond to selective serotonine reuptake inhibitor therapy. J Psychiatr Res 2007; 14: 84–90.
- 127. Obuchowicz E, Kowalski J, Labuzek K, Krysiak R, Pendzich J, Herman ZS. Amitriptyline and nortriptyline inhibit interleukin-1β and tumour necrosis factor-α release by rat mixed glial and microglial cell cultures. Int J Neuro-psychopharmacol 2005; 8: 1–9.
- 128. Pace TWW, Hu F, Miller AH. Cytokine-effects on glucocorticoid receptor function: relevance to glucocorticoid resistance and the pathophysiology and treatment of major depression. Brain Behav Immun 2007; 21:9–19.
- 129. Pae, CU, Yu, HS, Kim, TS, Lee, CU, Lee, SJ, Jun, TY, Lee, C, Serretti, A, Paik, IH. Monocyte chemoattractant protein-1 (MCP1) promoter –2518 polymorphism may confer a susceptibility to major depressive disorder in the Korean population. Psychiatry Res 2004; 127: 279–281.
- Pavon L, Sandoval-López G, Hernández ME, Loría F, Estrada I, Pérez M, Moreno J, Ávila U, Leff P, Antón B, Heinze G: Th2 cytokine response in major depressive disorder patients before treatment. J Neuroimmunol 2006; 172: 156–165.
- 131. Paykel ES, Brugha T, Fryers T. Size and burden of depressive disorders in Europe. Eur Neuropsychopharmacol 2005; 15: 411–423.
- 132. Pike JL, Irwin MR. Dissociation of inflammatory markers and natural killer cell activity in major depressive disorder. Brain Behav Immun 2006; 20: 169–174.
- 133. Pop VJ, Maartens JH, Leusink G, van Son MJ, Knottnerus AA, Ward AM; Metcalfe R, Weetman AP. Are autoimmune thyroid dysfunction and depression related? J Clin Endocrinol metab 1998; 83: 3194–3197.
- 134. Raison CL, Demetrashvili M, Capuron L, Miller AH. Neuropsychiatric adverse effects of interferon-alpha: recognition and management. CNS Drugs 2005; 19: 105–123.
- 135. Rausch JL. Initial conditions of psychotropic drug response: studies of serotonin transporter long promoter region (5-HTTLPR), serotonin transporter efficiency, cytokine and kinase gene expression relevant to depression and antidepressant outcome. Prog neuro-Psychiatr & Biol Psychiatr 2005; 29: 1046–1061.
- 136. Regier DA, Farmer ME, Rae DS, Myers JK, Kramer m, Robins LN, George LK, Karno M, Lake BZ. One month prevalence of mental disorders in the United States and sociodemographic characteristics: the Epidemiologic Catchment Area study. Acta Psychiatr Scand 1993; 88: 35–47.
- 137. Regier DA, Boyd JH, Burke JD, Rae DS, Myers JK, Kramer M, Robins LN, George LK, Karno M, Locke BZ. One-month prevalence of mental disorders in the United States. Arch Gen Psychiatry 1988; 45: 977–986.

- 138. Reichenberg A, Kraus T, Haack M, Schuld A, Pollmächer T, Yirmiya R. Endotoxin-induced changes in food consumption in healthy volunteers are associated with TNF- α and IL-6 secretion. Psychoneuroendocrinol 2002; 27: 945–956.
- Reynolds JL, Ignatowski TA, Sud R, Spengler RN. An natidepressant mechanisn of desipramine is to decrease tumor necrosis factor-α production culminating in increases in noradrenergic neurotransmission. Neuroscience 2005; 133: 519–531.
- 140. Rief W, Pilger F, Ihle D, Bosmans E, Egyed B, Maes M. Immunological differences between patients with major depression and somatisation syndrome. Psychiatry Res 2001; 105: 165–174.
- 141. Rohde P, Lewinsohn PM, Klein DN, Seeley JR. Association of parental depression with psychiatric course from adolescence to young adulthood among formerly depressed individuals. J Abnorm Psychol 2005; 114: 409–420.
- 142. Rosa, A, Peralta, V, Papiol, S, Cuesta, MJ, Serrano, F, Martinez-Larrea, A, Fananas, L. Interleukin-1beta (IL-1beta) gene and increased risk for the depressive symptom-dimension in schizophrenia spectrum disorders. Am J Med Genet B Neuropsychiatr Genet. 2004; 124: 10–14.
- 143. Rothermundt M, Arolt V, Peters M, Gutbrodt H, Fenker J, Kersting A, Kirchner H. Inflammatory markers in major depression and melancholia. J Affect Disord 2001; 63: 93–102.
- 144. Roumestan C, Michel A, Bichon F, Portet K, Detoc M, Henriquet C, Jaffuel D, Mathieu M. Anti-inflammatory properties of desipramine and fluoxetine. Respir Res 2007; 8:35–47.
- 145. Rush AJ, Zimmerman M, Wisniewski SR, Fava M, Hollon SD, Warden D, Biggs MM, Shores-Wilson K, Shelton RC, Luther JF, Thomas B, Trivedi MH. Comorbid psychiatric disorders in depressed outpatients: Demographic and clinical features. J Affect Disord 2005; 87: 43–55.
- 146. Sagud M, Effects of sertraline treatment on plasma cortisol, prolactin and thyroid hormones in female depressed patients. Sagud M, Pivac N, Mück-Seler D, Jakovljević M, Mihaljević-Peles A, Korsić M. Neuropsychobiology. 2002; 45:139–143.
- 147. Schiepers OJG, Wichers MC, Maes M. Cytokines and major depression. Progress in Neuro-Psychopharmacology & Biological Psychiatry 2005; 29: 201–217.
- 148. Schlatter J, Ortuňo F, Cervera-Enguix S. Lymphacyte subsets and lymphokine production in patients with melancholic versus nonmelancholic depression. Psychiatry Res 2004; 128: 259–65.
- 149. Schumacher J, Jamra RA, Becker T, Ohlraun S, Klopp N, Binder EB, Schulze TG, Deschner M, Schmäl C, Höfels S, Zobel A, Illig T, Propping P, Holsboer F, Rietschel M, Nöthen MM, Cichon S. Evidence for a relationship between genetic variants at the brain-derived neurotrophic factor (BDNF) locus and major depression. Biol Psychiatr 2005; 58: 307–314.
- 150. Schwarz MJ, Chiang S, Müller N, Ackenheil M. T-helper-1 and T-helper-2 responses in psychiatric disorders. Brain Behav Immun 2001; 15: 340–370.
- Seidel A, Arolt V, Hunstiger M, Rink L, Behnisch A, Kirchner H. Cytokine production and serum proteins in depression. Scand J Immunol 1995; 41: 534– 538.
- Serretti A. Genetics of Mood Disorders. In: Griez EJL, Faravelli C, Nutt DJ, Zohar J. (Eds.) Mood disorders: clinical management and research issues. John Wiley & Sons Ltd, England, 2005; pp. 35–75.

- 153. Simon NM, McNamara K, Chow CW, Maser RS, Papakostas GI, Pollack MH, Nierenberg AA, Fava M, Wong KK. A detailed examination of cytokine abnormalities in major depressive disorder. Eur Neuropsychopharmacol 2008; 18: 230–233.
- 154. Smit F, Beekman A, Cuijpers P, de Graaf R, Vollebergh W. Selecting key variables for depression prevention: results from a population-based prospective epidemiological study. J Affect Disord 2004; 81: 241–249.
- 155. Song C, Lin A, Bonaccorso S, Heide C, Verkerk R, Kenis G, Bosmans E, Scharpe S, Whelan A, Cosyns P, de Jongh R, Maes M. The inflammatory response system and the availability of plasma tryptophan in patients with primary sleep disorders and major depression. J Affect Disord 1998; 49: 211–219.
- 156. Steffens DC, Potter GG. Geriatric depression and cognitive impairment. Psychol Med 2008; 38: 163–175.
- 157. Suarez EC, Krishnan RR, Lewis JG. The relation of severity of depressive symptoms to monocyte-associated proinflammatory cytokines and chemokines in apparently healthy men. Psychosom Med 2003; 65: 362–368.
- 158. Suarez EC, Lewis JG, Krishnan RR, Young KH. Enhanced expression of cytokines and chemokines by blood monocytes toi n vitro lipopolysaccharide stimulation are associated with hostility and severity of depressive symptoms in healthy women. Psychoneuroendocrinol 2004; 29: 1119–1128.
- 159. Sullivan LE, Fiellin DA, O'Connor PG. The prevalence and impact of alcohol problems in major depression: a systematic review. Am J Med 2005; 118:330–341.
- 160. Sullivan PF, Neale MC, Kendler KS. Genetic epidemiology of major depression: review and meta-analysis. Am J Psychiatry 2000; 157: 1552–1562.
- Sutcigil L, Oktenli C, Musabak U, Bozkurt A, Cansever A, Uzun O, Sanisoglu SY, Yesilova N, Ozsahin A, Sengul A. Pro- and anti-inflammatory Cytokine balance in major depression: effect of sertraline therapy. Clin Dev Immunol 2007; 2007:1–6.
- 162. Szuster-Ciesielska A, Tustanowska-Stachura A, Słotwińska M, Marmurowska-Michałń M. In vitro immunoregulatory effects of antidepressants in healthy volunteers. Pol J Pharmacol 2003; 55. 353–362.
- Tayal V, Kalra BS. Cytokines and anti-cytokines as therapeutics An update. Eur J Psychopharmacol 2008; 579: 1–12.
- 164. Taylor WD, Steffens DC, MacFall JR, McQuoid DR, Payne ME, Provenzale JM, Krishnan RR. White matter hyperintensity progression and late-life depression outcomes. Arch Gen Psychiatry 2003; 60: 1090–1096.
- 165. Traks T, Koido K, Eller T, Maron E, Kingo K, Vasar V, Vasar E, Koks S. Polymorphisms in the interleukin-10 gene cluster are possibly involved in the increased risk for major depressive disorder. BMC Med Genet 2008; 16; 9:111.
- Tsao CW, Lin YS, Chen CC, Bai CH, Wu SR. Cytokines and serotonin transporte rin patients with major depression. Prog Neuro-Psychiatr & Biol Psychiatr 2006; 30: 899–905.
- 167. Tucker P, Ruwe WD, Masters B, Parker DE, Hossain A, Trautman RP, Wyatt DB. Neuroimmune and cortisol changes in selective serotonin reuptake inhibitor and placeba treatment of chronic posttraumatic stress disorder. Biol Psychiatry 2004; 56: 121–128.

- Tuglu C, Kara SH, Caliyurt O, Vardar E, Abay E. Increased serum tumor necrosis factor-alpha levels abd treatrment response in major depressive disorder. Psychopharmacol 2003; 170: 429–433.
- 169. Van West D, Kenis G, Maes M. Stress and depression. In: Griez EJL, Faravelli C, Nutt DJ, Zohar J. (Eds.) Mood disorders: clinical management and research issues. John Wiley & Sons Ltd, England, 2005; pp. 210–228.
- 170. Verma R, Holmans P, Knowles JA, Grover D, Evgrafov OV, Crowe RR, Scheftner WA, Weissman MM, DePaulo Jr JR, Potash JB, Levinson DF. Linkage disequilibrium mapping of a chromosome 15q25–26 major depression linkage region and sequencing of NTRK3. Biol Psychiatry 2008; 63:1185–1189.
- 171. Wang H, Buchner M, Moser MT, Daniel V, Schiltenwolf M. The role of IL-8 in patients with fibromyalgia. A prospective longitudinal study of 6 months. Clin J Pain 2009; 25: 1–4.
- 172. Weissman MM, Bland RC, Canino GJ, Faravelli C, Greenwald S, Hwu HG, Joyce PR, Karam EG, Lee CK, Lellouch J, Lépine JP, Newman SC, Rubio-Stipec M, Wells JE, Wickramaratne PJ, Wittchen H, Yeh EK. Cross-national epidemiology of major depression and bipolar disorder. JAMA 1996; 276: 293–299.
- 173. Weizman R, Laor N, Podliszewski E, Notti I, Djaldetti M, Bessler H. Cytokine production in major depressed patients before and after clomipramine treatment. Biol Psychiatry 1994; 35: 42–47 (abstract).
- 174. Wichers M, Maes M. The psychoneuroimmuno-pathophysiologyof cytokine induced depression in humans. Int J Neuropsychopharmacol 2002; 5: 375–388.
- 175. Wichers MC, Maes M. The role of indoleamine 2,3-dioxygenase (IDO) in the pathophysiology of interferon-α-induced depression. J Psychiatry Neurosci 2004; 29:11–17.
- 176. Willeit M, Praschak-Rieder N, Neumeister A, Pirker W, Asenbaum S, Vitouch O, Tauscher J, Hilger E, Stastny J, Brücke T, Kasper S. [1231]-beta-CIT SPECT imaging shows reduced brain serotonin transporter availability in drug-free depressed patients with seasonal affective disorder. Biol Psychiatry 2000; 47: 482– 489.
- Wittchen HU, Jacobi F. Size and burden of mental disorders in Europe a critical review and appraisal of 27 studies. Eur Neuropsychopharmacol 2005; 15: 357– 376.
- 178. Wood JG, Joyce PR, Miller AL, Mulder RT, Kennedy MA. A polymorphism in the Dopamine β -hydroxylase gene is associated with "paranoid ideation" in patients with major depression. Biol Psychiatry 2002; 51: 365–369.
- 179. Xia Z, DePierre JW, Nässberger L. Tricyclic antidepressants inhibit IL-6, IL-1 beta and TNF-alpha release in human blood monocytes and IL-2 and interferon-gamma in T cells.Immunopharmacology 1996; 34: 27–37.
- Yu YWY, Chen TJ, Hong CJ, Chen HM, Tsai SJ. Association study of the interleukin-1beta (C-511T) genetic polymorphism with major depressive disorder, associated symptomatology, and antidepressant response. Neuropsychopharmacol 2003; 28:1182–1185.
- 181. Zhu CB, Blakely RD, Hewlett WA. The proinflammatory cytokines interleukin-1beta and tumor necrosis factor-alpha activate serotonin transporters. Neuropsychopharmacol 2006; 31: 2121–2131.
- 182. Zubenko GS, Hughes, HB 3rd, Maher BS, Stiffler JS, Zubenko WN, Marazita ML. Genetic linkage of region containing the *CREB1* gene to depressive disorders

in women from families with recurrent, early-onset, major depression. Am J Med Genet B Neuropsychiatr Genet 2002; 114:980–987.

- 183. Zubenko GS, Maher BS, Hughes, HB 3rd, Zubenko WN, Stiffler JS, Kaplan BB, Marazita ML. Genome-wide linkage survey for genetic loci that influence the development of depressive disorders in families with recurrent, early-onset, major depression. Am J Med Genet B Neuropsychiatr Genet 2003; 123:1–18.
- 184. Zubenko GS, Maher BS, Hughes HB 3rd, Zubenko WN, Stiffler JS, Marazita ML. Genome-wide linkage survey for genetic loci that affect the risk of suicide attempts in families with recurrent, early-onset, major depression. Am J Med Genet B Neuropsychiatr Genet 2004; 129:47–54.

SUMMARY IN ESTONIAN

Immunoloogilised muutused unipolaarse depressiooni korral ja antidepressiivse ravi käigus

Sissejuhatus

Unipolaarne depressioon on ühiskonnas sage häire, mis põhjustab olulist elukvaliteedi ja sotsiaalse funktsioneerimise langust. Depressioonil on märkimisväärset mõju ka kaasnevate psüühika- ja kehaliste häirete kulule ja ravitulemustele.

Depressioon on polüfaktoriaalne haigus, mille tekkes osalevad koosmõjuna nii geneetilised kui ka keskkonnategurid. Oluliseks on osutunud vanus, sugu, partnerlus, etniline kuuluvus, haridus, immigrandistaatus, linna-maapiirkonna erinevused, eluolukorrad, lapsepõlve traumad, kaasnevad haigused. Depressiooni patogeneesis on teadaolevalt roll monoamiinidel: serotoniinil, noradrenaliinil ja dopamiinil. Samuti on leitud, et umbes pooltel depressioonihaigetest on häirunud HPA-telje talitlus, esinevad häired glükokortikoid- ja mineralokortikoidretseptorite süsteemis. On alust arvata, et HPA-telje üleaktiivsus stressolukordade tagajärjel tekib neil isikuil, kellel on predispositsioon HPA-telje regulatsioonihäirele. Aju visualiseerimisuuringutel on püsivamateks leidudeks aju külgvatsakeste laienemine, otsmiku- ja temporaalsagarate, samuti hipokampuse ja mõnede basaaltuumade mahu vähenemine. Ühe hüpoteesi kohaselt võiks see olla kroonilise hüperkortisoleemia tulemus, kuid on ka arvamus, et muutused esinevad juba haiguse varases staadiumis, olles depressiooni teket ennustavaks ilminguks. Tulemused geneetiliste tegurite osas on seni veel lõplikult kindlaks tegemata, ilmselt nii depressiooni fenotüübiliste erinevuste kui ka geneetilise keerukuse tõttu. On leitud mitmeid kromosoomipiirkondi, mis seonduvad depressiooniga, näiteks 1p, 1q, 2q, 4q, 5q, 8p, 10p, 11pter, 11g, 15g, 18g, 19p, Xg. Samas on eraldi piirkonnad, mida seostatakse käitumise ja depressiooni varase algusega. Geneetilised suitsidaalse assotsiatsiooniuuringud keskenduvad geenidele, mida seostatakse põhiliste neuromediaatoritega või teiste patogeneesimehanismidega, näiteks HPA-teljega. Üks perspektiivseid suundi on depressiooni farmakogeneetika, mis uurib geneetilise variatsiooni mõju ravivastusele.

Viimaste aastate jooksul on enam tähelepanu hakatud pöörama ka immuunsüsteemi osatähtsusele depressiooni korral. Immuunsüsteemi primaarne roll on organismi kaitsmine viiruste, bakterite ja kasvajarakkude eest. Erinevad immuunrakud suhtlevad omavahel tsütokiinide abil. Laias laastus võib tsütokiinid jaotada kaheks: põletikutsütokiinideks (näiteks interleukiin-1, interleukiin-6, tuumor-nekroosfaktor alfa (TNF- α) ning põletikuvastasteks tsütokiinideks (interleukiin-10, interleukiin-1 retseptori antagonist). Lisaks sellele, et tsütokiinid toimivad virgatsainetena immuunsüsteemi rakkude vahel, on neil roll ka ajus, kus nad osalevad neurokeemiliste, neuroendokriinsete, neuroimmuunsete ja käitumuslike muutuste tekkes. On leitud näiteks, et põletikutsütokiinid tõstavad HPA-telje aktiivsust, mõjutavad monoamiinide ja peptiidide taset hüpotaalamuse välistes piirkondades. Samuti on tõendeid, et põletikutsütokiinid aktiveerivad ensüüm indoleamiin-2,3-dioksügenaasi (IDO), mis konverteerib trüptofaanist kinureiini, mitte vajaliku serotoniini; samuti on leitud, et tõuseb serotoniin-transporteri aktiivsus, mis omakorda vähendab ekstratsellulaarse serotoniini taset.

Patsientidel, kes saavad tsütokiin-immuunteraapiat, tekivad kõrvaltoimena sageli depressiooni sümptomid, mis antidepressantide abil taanduvad. Depressioonihaigetel on korduvalt näidatud kõrgemaid põletikutsütokiinide tasemeid kui tervetel kontrollisikutel, samas on erinevad uuringud olnud vastuoluliste tulemustega. Uurimise all on olnud seosed tsütokiinide ja depressiooni raskusastme, melanhooliasümptomite esinemise või unemustritega. Lahknevaid tulemusi on saadud erinevate antidepressantide kasutamisega. Üldine seisukoht on, et antidepressantidel on leitud depressiooni korral immunomoduleeriv toime. Kuigi enam rõhutatakse põletikutsütokiinide taseme langust ja põletikuvastaste tsütokiinide taseme tõusu antidepressantravi foonil, on ka siin tulemused vastuolulised, sõltudes kasutatavast ravimist, uuringu kestvusest ning mitmetest muudest teguritest.

Ka kilpnäärme talitlushäireid on seostatud meeleoluhäiretega, rõhuasetusega subkliinilisele alatalitlusele, mille korral on leitud ka halvemat ravivastust antidepressantravile. Oluliselt vähem on uuringuid, mis keskenduvad kilpnäärme vastaste antikehade rollile meeleoluhäirete korral. Vaid üks uuring toob välja, et kilpnäärme vastaste antikehade esinemine on seotud halvema ravile reageerimisega depressioonihaigetel.

Uurimuse põhieesmärgid

Uurimuse põhieesmärgiks oli selgitada välja võimalik interleukiin-8 (IL-8), TNF- α ja lahustuva interleukiin-2 retseptori (sIL-2R) haaratus unipolaarse depressiooni patogeneesis ning tuvastada nende põletikutsütokiinide taseme muutused antidepressiivse ravi käigus.

Uurimuse täpsemad eesmärgid olid järgnevad:

- 1. Võrrelda TNF- α ja sIL-2R seerumkontsentratsioone depressioonihaigetel ja tervetel vabatahtlikel.
- 2. Leida võimalike seoseid uuritavate tsütokiinide seerumkontsentratsioone ja depressiooni raskuse vahe, mida määrati HAM-D skaalaga.
- 3. Leida võimalikke seoseis uuritavate tsütokiinide ja depressiooni üksiksümptomite vahel.
- 4. Leida estsitalopraamravi mõju IL-8, TNF- α ja sIL-2R tasemetele depressioonihaigetel.
- 5. Selgitada välja, kas bupropioonaugmentatsioon põhjustab muutusi IL-8, TNF-α ja sIL-2R produktsioonis estsitalopraamresistentsetel depressioonihaigetel.

- 6. Leida võimalikud seosed IL-8, TNF- α ja sIL-2R tasemete ja antidepressantravile reageerimise vahel depressioonihaigetel.
- 7. Selgitada välja, kas kilpnäärme peroksidaasi vastased antikehad või kilpnäärmehormoonid (täpsemalt TSH, T3, T4) mõjutavad patsientide estsitalopraamravile reageerimist.

Materjal ja meetodid

Kõik uuringusse kaasatud isikud olid Eesti elanikud. Depressioonihaiged vanusepiirides 15–65 aastat selekteeriti Tartu Ülikooli Kliinikumi Psühhiaatriakliinikusse pöördunud patsientide seast. Depressiooni diagnoos määrati DSM-IV kriteeriumite järgi, kasutades diagnostilist struktureeritud intervjuud M.I.N.I.5.0.0 ja psühhiaatrilist intervjuud. Raskusastme hindamiseks olid kasutusel Hamiltoni depressiooni skaala (HAMD), Becki depressiooni hindamise küsimustik (BDI) ning ravifaasis osalenutel ka Montgomeri-Asbergeri depressiooniskaala (MADRS). Tervetel isikutel välistasime depressiooniepisoodid minevikus, samuti esimese astme sugulastel. Raskemad kehaliselt haigused olid uurimuses osalemise välistavateks teguriteks.

Kokku osales uurimuses 247 depressioonihaiget ning 94 tervet kontrollisikut. Neist 166 depressioonihaiget osales ravifaasis, mille käigus uuritavad said antidepressantravi estsitalopraamiga. Esimese 4 nädala jooksul oli kõigil osalenutel raviannus 10 mg, neil patsientidel, kel selle aja jooksul depressiooniskoor MADRS skaalal langes vähem kui 50%, oli edasine raviannus 20 mg. Raviannust tõstsime ka neil patsientidel, kel hilisemas uuringufaasis tekkis tagasilöök. Kokku kestis esimene raviperiood 12 nädalat. Raviperioodi lõpus analüüsisime eraldi paranenuid (MADRS skoor alla 12) ja mitte ravile reageerinud patsiente (skoori langus alla 50%). Kuna patsientide arv, kes ravile reageerisid skoori alanemisega enam kui 50%, kuid kelle lõppskoor oli siiski üle 12 punkti, oli väga väike, analüüsisime neid ühiselt esimeses rühmas. Ravi teise faasi kaasasime 28 estsitalopraamravile mitte reageerinud isikut, kes said järgneva 6 nädala jooksul lisaks 20 mg estsitalopraamile 150-300 mg bupropiooni (noradernaliini ja dopamiini tranporteri blokaator). Sarnaselt esimese faasiga, analüüsisime raviperioodi lõpus eraldi ravile reageerinud ja mittereageerinud patsiente.

Tsütokiinide määramisel kasutati IMMULITE süsteemi, kilpnäärmehormoonid määrati kemiluminestsentsmeetodil ja anti-TPO määramisel kasutati ImmunoCAP süsteemi.

Peamised tulemused

Esimeses uurimuses võrreldi TNF- α ja sIL-2R tasemeid käesolevalt esmase ja korduva depressiooniga patsientidel, remissioonis patsientidel ning tervetel kontrollisikutel. Uurimuses selgus, et sIL-2R tase seerumis oli remissioonis patsientidel oluliselt madalam võrreldes nii tervete kontrollisikute kui ka korduvate depressioonihaigetega võrreldes. Esmaste depressioonihaigete suhtes täheldati samasuunalist tendentsi. Samuti selgus, et eelnev antidepressantravi kasutamine tulemust ei mõjutanud. TNF-α tase nende gruppide vahel ei erinenud, kuid käesolevalt depressiivsete haigetel oli TNF-α tase madalam kui eutüümsetel isikutel (remissioonis haiged koos tervete kontrollisikutega). Ilmnes, et eelnevalt antidepressantidega ravitud isikutel oli TNF-α tase madalam kui ravinaiivsetel haigetel ja tervetel kontrollisikutel. Ainult ravinaiivseid isikuid analüüsides leidsime, et korduva depressiooni korral oli sIL-2R tase kõrgem kui ülejäänud gruppides. Depressioonihaigetel korreleerusid HAMD skoorid positiivselt TNF-α tasemega. Seoseid depressioonepisoodi kestvusega, eelnevate episoodide arvuga, suitsetamisharjumuste, melanhoolsete sümptomite ja määratud tsütokiinide vahel esile ei tulnud. Üksiksümptomitest seostusid TNF-α väärtustega alanenud aktiivsustase ning agiteeritus; sIL-2R väärtustega alanenud aktiivsustase ning suitsidaalsus.

Teine uurimus keskendus estsitalopraamravi mõjule tsütokiinide tasemele. Selgus, et ravile reageerinud patsientidel alanes sIL-2R seerumkontsentratsioon ravi esimese 4 nädala jooksul, samas raviresistentsetel patsientidel sIL-2R tase esialgu kergelt tõusis, alanedes alles 4 ja 12 ravinädala vahel. IL-8 ja TNF- α osas kahe grupi vahel erinevusi ei täheldatud. Sama uuring näitas, et ravile reageerinud patsientidel oli ravieelne TNF- α tase madalam kui raviresistentsetel haigetel ja tervetel kontrollisikutel.

Bupropioonaugmentatsiooni faasis ei esinenud olulisi grupi- ja ajagrupi seoseid, kuid ilmnes ajaefekt IL-8 osas: 6 nädala jooksul tõusis IL-8 tase kogu grupi lõikes. Augmentatsioonifaasi ravitulemust ei õnnestunud ühegi tsütokiini taseme abil ette ennustada.

Kuigi anti-TPO esines vaid kaheksal naispatsiendil ja kahel tervel naisel, tuli uurimuses esile tendents, et estsitalopraamile raviresistentsetel naistel on enam anti-TPO positiivsust kui ravile reageerinud naistel. Olulisi seoseid kilpnäärmehormoonide, depressiooni parameetrite ja ravitulemuste vahel esile ei tulnud.

Järeldused

- 1. Depressiivsetel patsientidel oli TNF- α tase madalam kui eutüümsetel isikutel. Depressioonihaigete eelnev antidepressantravi oli seotud madalamate TNF- α väärtustega võrreldes ravinaiivsete haigete ja tervete kontrollisikutega.
- 2. Depressioonihaigetel korreleerus TNF-α tase positiivselt HAMD skooridega. sIL-2R taseme ja depressiooni raskusastmega seoseid esile ei tulnud.
- 3. TNF-α väärtused korreleerusid alanenud aktiivsuse ja agiteeritusega ja sIL-2R väärtused alanenud aktiivsustaseme ning suitsidaalsusega.
- Immuunsüsteemi vastus, mõõdetuna sIL-2R produktsioonis, erines estsitalopraamravi resistentsete ja ravile reageerinud patsientide vahel. sIL-2R väärtused näitasid raviresistentsetel isikutel langustendentsi ajaliselt hiljem. IL-8 ja TNF-α osas erinevusi esile ei tulnud.

- 5. Estsitalopraamravi augmenteerimine bupropiooniga põhjustas olulise IL-8 produktsiooni tõusu 6 nädalase perioodi vältel. Ravi ei mõjutanud TNF-α ja sIL-2R taset.
- 6. Estsitalopraamravile reageerinud patsientidel esinesid madalamad TNF- α ravieelsed väärtused võrreldes raviresistentsete haigete ja tervete kontrollisikutega. Oluline oli siin ka sooefekt: meestel, kes ravile ei reageerinud, esinesid kõrgemad TNF- α ravieelsed väärtused võrreldes ravile reageerinud meestega, samuti nii raviresistentsete kui ka ravile reageerinud naistega.
- 7. Raviresistentsete naiste hulgas oli enam anti-TPO positiivseid isikuid kui ravile reageerinud naiste hulgas. TSH, T3 ja T4 ravieelne tase ei seostunud raviefekti ega ka depressiooni parameetritega.

ACKNOWLEDGEMENTS

This work was carried out at the Department of Psychiatry, University of Tartu. The financial support was received from the target grant 0423 from the Ministry of Education of Estonia (Prof Veiko Vasar) and from Estonian Scientific Foundation (grant no 7034; Eduard Maron) and from Center of Molecular and Clinical Medicine (Prof. Raivo Uibo). Cipralex® was courtesy of Lundbeck Eesti AS and Wellbutrin® of GlaxoSmithKline Eesti OÜ.

I wish to express my gratidude to:

- Professor Veiko Vasar, Eduard Maron and Jakov Šlik for support and supervision
- Associate professor Anu Aluoja for support, help in English language and statistics
- Professor Raivo Uibo, Kaja Metsküla and Ija Talja for collaboration
- The nurses Merle Talvik, Birgit Aumeste, Ketlin Veeväli and Jane Puusepp for assistance
- All collegues who contributed their time and efforts to this work: Ülle Iher, psychiatrists who send their patients to the studies
- My family for care and support

PUBLICATONS

CURRICULUM VITAE

TRIIN ELLER

Citizenship: Estonian Date and place of birth: December 19, 1970, Tartu, Estonia Address: 31 Raja Street, Tartu 50417; Estonia Phone: +372 731 8835 Fax: +372 731 8801 E-mail: triin.eller@kliinikum.ee

Education

1978–1989	Nyo Secondary School
1989–1995	University of Tartu, Faculty of Medicine (MD)
1995–1997	Internship in general medicine (Estonian Seamen's Hospital)
1997–2001	Residency training in psychiatry (Tartu University Hospital)
2003-2009	University of Tartu, Faculty of Medicine, PhD studies in
	psychiatry

Professional employment

- 1993–1995 Tartu University Hospital, Nurse
- 1995–1997 Estonian Seamen's Hospital, Intern
- 1997–2001 Tartu University Hospital, Resident in Psychiatry
- 2004– Tartu University Hospital, Psychiatrist
- 2008– Tartu University, Department of Psychiatry, Teaching Assistant

Special Courses

Family psychology and psychotherapy training. Tartu, Estonia

- Intensive teaching programme on mood disorders. University of Maastricht, the Netherlands
- Intensive teaching programme on anxiety disorders. University of Maastricht, the Netherlands

Master's seminar in affective neurosciences. University of Bristol, UK

Core Maudsley Forum, Institute of Psychiatry, King's College, London, UK

Research

My research has primarily focused on the changes in the immune system in mood disorders. I have participated in a number of studies on the genetics of mental disorders and in research into sleep habits in medical students.

CURRICULUM VITAE

TRIIN ELLER

Kodakondsus: Eesti Sünniaeg ja koht: 19. detsember 1970, Tartu, Eesti Aadress: TÜ Psühhiaatriakliinik, Raja 31; Tartu 50417, Eesti Telefon: +372 731 8835 Faks: +372 731 8801 E-post: triin.eller@kliinikum.ee

Haridus

1978–1989	Nõo Keskkool
1989–1995	Tartu Ülikooli arstiteaduskond, arstiteadus
1995–1997	internatuur (Eesti Meremeeste Haigla)
1997-2001	Tartu Ülikooli Kliinikum, psühhiaatria residentuur
2003-2009	Tartu Ülikooli arstiteaduskond, doktoriõpe

Teenistuskäik

1993–1995	Tartu Ülikooli Kliinikum, õde
1995–1997	Eesti Meremeeste Haigla, intern
1997-2001	Tartu Ülikooli Kliinikum, arst-resident psühhiaatria erialal
2004–	Tartu Ülikooli Kliinikum, arst-õppejõud psühhiaatria erialal
2008-	Tartu Ülikool, psühhiaatria assistent

Täiendus

Perekonna psühholoogia ja psühhoteraapia, Tartu, Eesti Seminar-töötuba meeleoluhäiretest. Maastrichti Ülikool Seminar-töötuba ärevushäiretest. Maastrichti Ülikool Maudsley Foorum, Psühhiaatria Instituut, King's College, London, UK Talveseminar afektiivsest neuroteadusest. Bristoli Ülikool

Teadustöö

Teadustöö on keskendunud peamiselt immuunsüsteemi iseärasustele meeleoluhäirete korral. Olen osalenud ka mõnes psühhiaatrilise geneetika valdkonna uurimuses, samuti meditsiinitudengite uneharjumusi käsitlevas töös.

List of Publications

- 1. Eller T, Aluoja A, Maron E, Vasar V. Soluble interleukin 2 receptor and tumour necrosis factor in depressed patients in Estonia. Medicina, 2009 (accepted).
- Eller T, Aluoja A, Vasar V, Veldi M. Symptoms of anxiety and depression in Estonian medical students with sleep problems. Depress Anxiety. 2006; 23 (4): 250–6.
- 3. Eller T, Metsküla K, Talja I, Maron E, Uibo R, Vasar V. Thyroid autoimmunity and treatment response to escitalopram in major depression (submitted to Nord J Psychiatry, 2008).
- 4. Eller T, Vasar V, Shlik J, Maron E. Pro-inflammatory cytokines and treatment response to escitaloprsam in major depressive disorder. Prog Neuropsychopharmacol Biol Psychiatry. 2008; 15; 32 (2): 445–50.
- 5. Eller T, Vasar V, Shlik J, Maron E. Effects of bupropion augmentation on proinflammatory cytokines in escitalopram-resistant patients with major depressive disorder. J Psychopharmacol. 2009; 23: 854–858.
- Eller T, Vasar, V, Shlik J, Maron M. The role of IL-2 and sIL-2R in depression and antidepressant response. Current Opinion in Investigational Drugs 2009; 10: 638–643.
- Koido, K, Eller T, Kingo K, Kõks S, Traks T, Shlik J, Vasar V, Vasar E, Maron E. Interleukin 10 family gene polymorphisms are not associated with major depressive disorder and panic disorder phenotypes. J Psychoatric Research 2009 (in press).
- 8. Maron E, Eller T, Vasar V, Nutt DJ. Effects of bupropion augmentation in escitalopram-resistant patients with major depressive disorder: an open-label, naturalistic study J Clin Psychiatry 2009; 70(7):1054–1056.
- Maron E, Eller T, Vasar V, Nutt DJ. Bupropiooni augmentatsiooni toime estsitalopraamravi suhtes resistentsetel depressiivsetel patsientidel. Eesti Arst 2009; 88(2): 82–90.
- Maron E, Tammiste A, Kallassalu K, Eller T, Vasar V, Nutt DJ, Metspalu A. Serotonin transporter promoter region polymorphisms do not influence treatment response to escitalopram in patients with major depression. Eur Neuropsychopharmacol 2009; 19(6); 451–456.
- 11. Traks T, Koido K, **Eller T**, Maron E, Kingo K, Vasar V, Vasar E, Koks S. Polymorphisms in the interleukin-10 gene cluster are possibly involved in the increased risk for major depressive disorder. BMC Med Genet. 9:111. (in press)
- 12. Veldi M, Vasar V, Hion T, Eller T, Shlik J, Kull M. Vanus ja uneapnoehaigus riskitegurid ja päevased kaebused. Eesti Arst 2001; 8: 325–328.

DISSERTATIONES MEDICINAE UNIVERSITATIS TARTUENSIS

- 1. **Heidi-Ingrid Maaroos.** The natural course of gastric ulcer in connection with chronic gastritis and *Helicobacter pylori*. Tartu, 1991.
- 2. **Mihkel Zilmer.** Na-pump in normal and tumorous brain tissues: Structural, functional and tumorigenesis aspects. Tartu, 1991.
- 3. **Eero Vasar.** Role of cholecystokinin receptors in the regulation of behaviour and in the action of haloperidol and diazepam. Tartu, 1992.
- 4. **Tiina Talvik.** Hypoxic-ischaemic brain damage in neonates (clinical, biochemical and brain computed tomographical investigation). Tartu, 1992.
- 5. Ants Peetsalu. Vagotomy in duodenal ulcer disease: A study of gastric acidity, serum pepsinogen I, gastric mucosal histology and *Helicobacter pylori*. Tartu, 1992.
- 6. **Marika Mikelsaar.** Evaluation of the gastrointestinal microbial ecosystem in health and disease. Tartu, 1992.
- 7. Hele Everaus. Immuno-hormonal interactions in chronic lymphocytic leukaemia and multiple myeloma. Tartu, 1993.
- 8. **Ruth Mikelsaar.** Etiological factors of diseases in genetically consulted children and newborn screening: dissertation for the commencement of the degree of doctor of medical sciences. Tartu, 1993.
- 9. Agu Tamm. On metabolic action of intestinal microflora: clinical aspects. Tartu, 1993.
- 10. Katrin Gross. Multiple sclerosis in South-Estonia (epidemiological and computed tomographical investigations). Tartu, 1993.
- 11. **Oivi Uibo.** Childhood coeliac disease in Estonia: occurrence, screening, diagnosis and clinical characterization. Tartu, 1994.
- 12. Viiu Tuulik. The functional disorders of central nervous system of chemistry workers. Tartu, 1994.
- 13. **Margus Viigimaa.** Primary haemostasis, antiaggregative and anticoagulant treatment of acute myocardial infarction. Tartu, 1994.
- 14. **Rein Kolk.** Atrial versus ventricular pacing in patients with sick sinus syndrome. Tartu, 1994.
- 15. **Toomas Podar.** Incidence of childhood onset type 1 diabetes mellitus in Estonia. Tartu, 1994.
- 16. **Kiira Subi.** The laboratory surveillance of the acute respiratory viral infections in Estonia. Tartu, 1995.
- 17. **Irja Lutsar.** Infections of the central nervous system in children (epidemiologic, diagnostic and therapeutic aspects, long term outcome). Tartu, 1995.
- 18. **Aavo Lang.** The role of dopamine, 5-hydroxytryptamine, sigma and NMDA receptors in the action of antipsychotic drugs. Tartu, 1995.
- 19. Andrus Arak. Factors influencing the survival of patients after radical surgery for gastric cancer. Tartu, 1996.

- 20. **Tõnis Karki.** Quantitative composition of the human lactoflora and method for its examination. Tartu, 1996.
- 21. **Reet Mändar.** Vaginal microflora during pregnancy and its transmission to newborn. Tartu, 1996.
- 22. **Triin Remmel.** Primary biliary cirrhosis in Estonia: epidemiology, clinical characterization and prognostication of the course of the disease. Tartu, 1996.
- 23. **Toomas Kivastik.** Mechanisms of drug addiction: focus on positive reinforcing properties of morphine. Tartu, 1996.
- 24. **Paavo Pokk.** Stress due to sleep deprivation: focus on GABA_A receptorchloride ionophore complex. Tartu, 1996.
- 25. **Kristina Allikmets.** Renin system activity in essential hypertension. Associations with atherothrombogenic cardiovascular risk factors and with the efficacy of calcium antagonist treatment. Tartu, 1996.
- 26. **Triin Parik.** Oxidative stress in essential hypertension: Associations with metabolic disturbances and the effects of calcium antagonist treatment. Tartu, 1996.
- 27. Svetlana Päi. Factors promoting heterogeneity of the course of rheumatoid arthritis. Tartu, 1997.
- 28. **Maarike Sallo.** Studies on habitual physical activity and aerobic fitness in 4 to 10 years old children. Tartu, 1997.
- 29. Paul Naaber. *Clostridium difficile* infection and intestinal microbial ecology. Tartu, 1997.
- 30. Rein Pähkla. Studies in pinoline pharmacology. Tartu, 1997.
- 31. Andrus Juhan Voitk. Outpatient laparoscopic cholecystectomy. Tartu, 1997.
- 32. Joel Starkopf. Oxidative stress and ischaemia-reperfusion of the heart. Tartu, 1997.
- 33. Janika Kõrv. Incidence, case-fatality and outcome of stroke. Tartu, 1998.
- 34. Ülla Linnamägi. Changes in local cerebral blood flow and lipid peroxidation following lead exposure in experiment. Tartu, 1998.
- 35. Ave Minajeva. Sarcoplasmic reticulum function: comparison of atrial and ventricular myocardium. Tartu, 1998.
- 36. **Oleg Milenin.** Reconstruction of cervical part of esophagus by revascularised ileal autografts in dogs. A new complex multistage method. Tartu, 1998.
- 37. **Sergei Pakriev.** Prevalence of depression, harmful use of alcohol and alcohol dependence among rural population in Udmurtia. Tartu, 1998.
- 38. Allen Kaasik. Thyroid hormone control over β -adrenergic signalling system in rat atria. Tartu, 1998.
- 39. Vallo Matto. Pharmacological studies on anxiogenic and antiaggressive properties of antidepressants. Tartu, 1998.
- 40. **Maire Vasar.** Allergic diseases and bronchial hyperreactivity in Estonian children in relation to environmental influences. Tartu, 1998.

- 41. **Kaja Julge.** Humoral immune responses to allergens in early childhood. Tartu, 1998.
- 42. **Heli Grünberg.** The cardiovascular risk of Estonian schoolchildren. A cross-sectional study of 9-, 12- and 15-year-old children. Tartu, 1998.
- 43. **Epp Sepp.** Formation of intestinal microbial ecosystem in children. Tartu, 1998.
- 44. **Mai Ots.** Characteristics of the progression of human and experimental glomerulopathies. Tartu, 1998.
- 45. Tiina Ristimäe. Heart rate variability in patients with coronary artery disease. Tartu, 1998.
- 46. Leho Kõiv. Reaction of the sympatho-adrenal and hypothalamo-pituitaryadrenocortical system in the acute stage of head injury. Tartu, 1998.
- 47. Bela Adojaan. Immune and genetic factors of childhood onset IDDM in Estonia. An epidemiological study. Tartu, 1999.
- 48. Jakov Shlik. Psychophysiological effects of cholecystokinin in humans. Tartu, 1999.
- 49. **Kai Kisand.** Autoantibodies against dehydrogenases of α -ketoacids. Tartu, 1999.
- 50. Toomas Marandi. Drug treatment of depression in Estonia. Tartu, 1999.
- 51. Ants Kask. Behavioural studies on neuropeptide Y. Tartu, 1999.
- 52. Ello-Rahel Karelson. Modulation of adenylate cyclase activity in the rat hippocampus by neuropeptide galanin and its chimeric analogs. Tartu, 1999.
- 53. **Tanel Laisaar.** Treatment of pleural empyema special reference to intrapleural therapy with streptokinase and surgical treatment modalities. Tartu, 1999.
- 54. Eve Pihl. Cardiovascular risk factors in middle-aged former athletes. Tartu, 1999.
- 55. **Katrin Õunap.** Phenylketonuria in Estonia: incidence, newborn screening, diagnosis, clinical characterization and genotype/phenotype correlation. Tartu, 1999.
- 56. Siiri Kõljalg. Acinetobacter an important nosocomial pathogen. Tartu, 1999.
- 57. Helle Karro. Reproductive health and pregnancy outcome in Estonia: association with different factors. Tartu, 1999.
- 58. **Heili Varendi.** Behavioral effects observed in human newborns during exposure to naturally occurring odors. Tartu, 1999.
- 59. Anneli Beilmann. Epidemiology of epilepsy in children and adolescents in Estonia. Prevalence, incidence, and clinical characteristics. Tartu, 1999.
- 60. Vallo Volke. Pharmacological and biochemical studies on nitric oxide in the regulation of behaviour. Tartu, 1999.
- 61. **Pilvi Ilves.** Hypoxic-ischaemic encephalopathy in asphyxiated term infants. A prospective clinical, biochemical, ultrasonographical study. Tartu, 1999.

- 62. Anti Kalda. Oxygen-glucose deprivation-induced neuronal death and its pharmacological prevention in cerebellar granule cells. Tartu, 1999.
- 63. **Eve-Irene Lepist.** Oral peptide prodrugs studies on stability and absorption. Tartu, 2000.
- 64. **Jana Kivastik.** Lung function in Estonian schoolchildren: relationship with anthropometric indices and respiratory symptomas, reference values for dynamic spirometry. Tartu, 2000.
- 65. **Karin Kull.** Inflammatory bowel disease: an immunogenetic study. Tartu, 2000.
- 66. **Kaire Innos.** Epidemiological resources in Estonia: data sources, their quality and feasibility of cohort studies. Tartu, 2000.
- 67. **Tamara Vorobjova.** Immune response to *Helicobacter pylori* and its association with dynamics of chronic gastritis and epithelial cell turnover in antrum and corpus. Tartu, 2001.
- 68. **Ruth Kalda.** Structure and outcome of family practice quality in the changing health care system of Estonia. Tartu, 2001.
- 69. Annika Krüüner. *Mycobacterium tuberculosis* spread and drug resistance in Estonia. Tartu, 2001.
- 70. **Marlit Veldi.** Obstructive Sleep Apnoea: Computerized Endopharyngeal Myotonometry of the Soft Palate and Lingual Musculature. Tartu, 2001.
- 71. Anneli Uusküla. Epidemiology of sexually transmitted diseases in Estonia in 1990–2000. Tartu, 2001.
- 72. Ade Kallas. Characterization of antibodies to coagulation factor VIII. Tartu, 2002.
- 73. **Heidi Annuk.** Selection of medicinal plants and intestinal lactobacilli as antimicrobil components for functional foods. Tartu, 2002.
- 74. Aet Lukmann. Early rehabilitation of patients with ischaemic heart disease after surgical revascularization of the myocardium: assessment of health-related quality of life, cardiopulmonary reserve and oxidative stress. A clinical study. Tartu, 2002.
- 75. **Maigi Eisen.** Pathogenesis of Contact Dermatitis: participation of Oxidative Stress. A clinical – biochemical study. Tartu, 2002.
- 76. **Piret Hussar.** Histology of the post-traumatic bone repair in rats. Elaboration and use of a new standardized experimental model bicortical perforation of tibia compared to internal fracture and resection osteotomy. Tartu, 2002.
- 77. **Tõnu Rätsep.** Aneurysmal subarachnoid haemorrhage: Noninvasive monitoring of cerebral haemodynamics. Tartu, 2002.
- 78. **Marju Herodes.** Quality of life of people with epilepsy in Estonia. Tartu, 2003.
- 79. **Katre Maasalu.** Changes in bone quality due to age and genetic disorders and their clinical expressions in Estonia. Tartu, 2003.

- 80. **Toomas Sillakivi.** Perforated peptic ulcer in Estonia: epidemiology, risk factors and relations with *Helicobacter pylori*. Tartu, 2003.
- 81. Leena Puksa. Late responses in motor nerve conduction studies. F and A waves in normal subjects and patients with neuropathies. Tartu, 2003.
- 82. Krista Lõivukene. *Helicobacter pylori* in gastric microbial ecology and its antimicrobial susceptibility pattern. Tartu, 2003.
- 83. **Helgi Kolk.** Dyspepsia and *Helicobacter pylori* infection: the diagnostic value of symptoms, treatment and follow-up of patients referred for upper gastrointestinal endoscopy by family physicians. Tartu, 2003.
- 84. **Helena Soomer.** Validation of identification and age estimation methods in forensic odontology. Tartu, 2003.
- 85. **Kersti Oselin.** Studies on the human MDR1, MRP1, and MRP2 ABC transporters: functional relevance of the genetic polymorphisms in the *MDR1* and *MRP1* gene. Tartu, 2003.
- 86. Jaan Soplepmann. Peptic ulcer haemorrhage in Estonia: epidemiology, prognostic factors, treatment and outcome. Tartu, 2003.
- 87. **Margot Peetsalu.** Long-term follow-up after vagotomy in duodenal ulcer disease: recurrent ulcer, changes in the function, morphology and *Helicobacter pylori* colonisation of the gastric mucosa. Tartu, 2003.
- 88. Kersti Klaamas. Humoral immune response to *Helicobacter pylori* a study of host-dependent and microbial factors. Tartu, 2003.
- 89. **Pille Taba.** Epidemiology of Parkinson's disease in Tartu, Estonia. Prevalence, incidence, clinical characteristics, and pharmacoepidemiology. Tartu, 2003.
- 90. Alar Veraksitš. Characterization of behavioural and biochemical phenotype of cholecystokinin-2 receptor deficient mice: changes in the function of the dopamine and endopioidergic system. Tartu, 2003.
- 91. **Ingrid Kalev.** CC-chemokine receptor 5 (CCR5) gene polymorphism in Estonians and in patients with Type I and Type II diabetes mellitus. Tartu, 2003.
- 92. Lumme Kadaja. Molecular approach to the regulation of mitochondrial function in oxidative muscle cells. Tartu, 2003.
- 93. Aive Liigant. Epidemiology of primary central nervous system tumours in Estonia from 1986 to 1996. Clinical characteristics, incidence, survival and prognostic factors. Tartu, 2004.
- 94. Andres, Kulla. Molecular characteristics of mesenchymal stroma in human astrocytic gliomas. Tartu, 2004.
- 95. Mari Järvelaid. Health damaging risk behaviours in adolescence. Tartu, 2004.
- 96. Ülle Pechter. Progression prevention strategies in chronic renal failure and hypertension. An experimental and clinical study. Tartu, 2004.

- 97. **Gunnar Tasa.** Polymorphic glutathione S-transferases biology and role in modifying genetic susceptibility to senile cataract and primary open angle glaucoma. Tartu, 2004.
- 98. **Tuuli Käämbre.** Intracellular energetic unit: structural and functional aspects. Tartu, 2004.
- 99. Vitali Vassiljev. Influence of nitric oxide syntase inhibitors on the effects of ethanol after acute and chronic ethanol administration and withdrawal. Tartu, 2004.
- 100. Aune Rehema. Assessment of nonhaem ferrous iron and glutathione redox ratio as markers of pathogeneticity of oxidative stress in different clinical groups. Tartu, 2004.
- 101. Evelin Seppet. Interaction of mitochondria and ATPases in oxidative muscle cells in normal and pathological conditions. Tartu, 2004.
- 102. Eduard Maron. Serotonin function in panic disorder: from clinical experiments to brain imaging and genetics. Tartu, 2004.
- 103. Marje Oona. *Helicobacter pylori* infection in children: epidemiological and therapeutic aspects. Tartu, 2004.
- 104. **Kersti Kokk.** Regulation of active and passive molecular transport in the testis. Tartu, 2005.
- 105. **Vladimir Järv.** Cross-sectional imaging for pretreatment evaluation and follow-up of pelvic malignant tumours. Tartu, 2005.
- 106. Andre Õun. Epidemiology of adult epilepsy in Tartu, Estonia. Incidence, prevalence and medical treatment. Tartu, 2005.
- 107. **Piibe Muda.** Homocysteine and hypertension: associations between homocysteine and essential hypertension in treated and untreated hypertensive patients with and without coronary artery disease. Tartu, 2005.
- 108. **Külli Kingo.** The interleukin-10 family cytokines gene polymorphisms in plaque psoriasis. Tartu, 2005.
- 109. **Mati Merila.** Anatomy and clinical relevance of the glenohumeral joint capsule and ligaments. Tartu, 2005.
- 110. **Epp Songisepp**. Evaluation of technological and functional properties of the new probiotic *Lactobacillus fermentum* ME-3. Tartu, 2005.
- 111. **Tiia Ainla.** Acute myocardial infarction in Estonia: clinical characteristics, management and outcome. Tartu, 2005.
- 112. Andres Sell. Determining the minimum local anaesthetic requirements for hip replacement surgery under spinal anaesthesia a study employing a spinal catheter. Tartu, 2005.
- 113. **Tiia Tamme.** Epidemiology of odontogenic tumours in Estonia. Pathogenesis and clinical behaviour of ameloblastoma. Tartu, 2005.
- 114. **Triine Annus**. Allergy in Estonian schoolchildren: time trends and characteristics. Tartu, 2005.
- 115. **Tiia Voor.** Microorganisms in infancy and development of allergy: comparison of Estonian and Swedish children. Tartu, 2005.

- 116. **Priit Kasenõmm.** Indicators for tonsillectomy in adults with recurrent tonsillitis clinical, microbiological and pathomorphological investigations. Tartu, 2005.
- 117. **Eva Zusinaite.** Hepatitis C virus: genotype identification and interactions between viral proteases. Tartu, 2005.
- 118. **Piret Kõll.** Oral lactoflora in chronic periodontitis and periodontal health. Tartu, 2006.
- 119. **Tiina Stelmach.** Epidemiology of cerebral palsy and unfavourable neurodevelopmental outcome in child population of Tartu city and county, Estonia Prevalence, clinical features and risk factors. Tartu, 2006.
- 120. **Katrin Pudersell.** Tropane alkaloid production and riboflavine excretion in the field and tissue cultures of henbane (*Hyoscyamus niger* L.). Tartu, 2006.
- 121. **Külli Jaako.** Studies on the role of neurogenesis in brain plasticity. Tartu, 2006.
- 122. Aare Märtson. Lower limb lengthening: experimental studies of bone regeneration and long-term clinical results. Tartu, 2006.
- 123. Heli Tähepõld. Patient consultation in family medicine. Tartu, 2006.
- 124. **Stanislav Liskmann.** Peri-implant disease: pathogenesis, diagnosis and treatment in view of both inflammation and oxidative stress profiling. Tartu, 2006.
- 125. **Ruth Rudissaar.** Neuropharmacology of atypical antipsychotics and an animal model of psychosis. Tartu, 2006.
- 126. **Helena Andreson.** Diversity of *Helicobacter pylori* genotypes in Estonian patients with chronic inflammatory gastric diseases. Tartu, 2006.
- 127. Katrin Pruus. Mechanism of action of antidepressants: aspects of serotoninergic system and its interaction with glutamate. Tartu, 2006.
- 128. **Priit Põder.** Clinical and experimental investigation: relationship of ischaemia/reperfusion injury with oxidative stress in abdominal aortic aneurysm repair and in extracranial brain artery endarterectomy and possibilities of protection against ischaemia using a glutathione analogue in a rat model of global brain ischaemia. Tartu, 2006.
- 129. Marika Tammaru. Patient-reported outcome measurement in rheumatoid arthritis. Tartu, 2006.
- 130. Tiia Reimand. Down syndrome in Estonia. Tartu, 2006.
- 131. **Diva Eensoo.** Risk-taking in traffic and Markers of Risk-Taking Behaviour in Schoolchildren and Car Drivers. Tartu, 2007.
- 132. **Riina Vibo.** The third stroke registry in Tartu, Estonia from 2001 to 2003: incidence, case-fatality, risk factors and long-term outcome. Tartu, 2007.
- 133. Chris Pruunsild. Juvenile idiopathic arthritis in children in Estonia. Tartu, 2007.
- 134. Eve Õiglane-Šlik. Angelman and Prader-Willi syndromes in Estonia. Tartu, 2007.

- 135. **Kadri Haller.** Antibodies to follicle stimulating hormone. Significance in female infertility. Tartu, 2007.
- 136. Pille Ööpik. Management of depression in family medicine. Tartu, 2007.
- 137. Jaak Kals. Endothelial function and arterial stiffness in patients with atherosclerosis and in healthy subjects. Tartu, 2007.
- 138. **Priit Kampus.** Impact of inflammation, oxidative stress and age on arterial stiffness and carotid artery intima-media thickness. Tartu, 2007.
- 139. Margus Punab. Male fertility and its risk factors in Estonia. Tartu, 2007.
- 140. Alar Toom. Heterotopic ossification after total hip arthroplasty: clinical and pathogenetic investigation. Tartu, 2007.
- 141. Lea Pehme. Epidemiology of tuberculosis in Estonia 1991–2003 with special regard to extrapulmonary tuberculosis and delay in diagnosis of pulmonary tuberculosis. Tartu, 2007.
- 142. Juri Karjagin. The pharmacokinetics of metronidazole and meropenem in septic shock. Tartu, 2007.
- 143. **Inga Talvik.** Inflicted traumatic brain injury shaken baby syndrome in Estonia epidemiology and outcome. Tartu, 2007.
- 144. **Tarvo Rajasalu.** Autoimmune diabetes: an immunological study of type 1 diabetes in humans and in a model of experimental diabetes (in RIP-B7.1 mice). Tartu, 2007.
- 145. **Inga Karu.** Ischaemia-reperfusion injury of the heart during coronary surgery: a clinical study investigating the effect of hyperoxia. Tartu, 2007.
- 146. **Peeter Padrik.** Renal cell carcinoma: Changes in natural history and treatment of metastatic disease. Tartu, 2007.
- 147. Neve Vendt. Iron deficiency and iron deficiency anaemia in infants aged 9 to 12 months in Estonia. Tartu, 2008.
- 148. Lenne-Triin Heidmets. The effects of neurotoxins on brain plasticity: focus on neural Cell Adhesion Molecule. Tartu, 2008.
- 149. **Paul Korrovits.** Asymptomatic inflammatory prostatitis: prevalence, etiological factors, diagnostic tools. Tartu, 2008.
- 150. **Annika Reintam.** Gastrointestinal failure in intensive care patients. Tartu, 2008.
- 151. **Kristiina Roots.** Cationic regulation of Na-pump in the normal, Alzheimer's and CCK₂ receptor-deficient brain. Tartu, 2008.
- 152. **Helen Puusepp.** The genetic causes of mental retardation in Estonia: fragile X syndrome and creatine transporter defect. Tartu, 2009.
- 153. Kristiina Rull. Human chorionic gonadotropin beta genes and recurrent miscarriage: expression and variation study. Tartu, 2009.
- 154. Margus Eimre. Organization of energy transfer and feedback regulation in oxidative muscle cells. Tartu, 2009.
- 155. Maire Link. Transcription factors FoxP3 and AIRE: autoantibody associations. Tartu, 2009.

- 156. Kai Haldre. Sexual health and behaviour of young women in Estonia. Tartu, 2009.
- 157. **Kaur Liivak.** Classical form of congenital adrenal hyperplasia due to 21-hydroxylase deficiency in Estonia: incidence, genotype and phenotype with special attention to short-term growth and 24-hour blood pressure. Tartu, 2009.
- 158. **Kersti Ehrlich.** Antioxidative glutathione analogues (UPF peptides) molecular design, structure-activity relationships and testing the protective properties. Tartu, 2009.
- 159. Anneli Rätsep. Type 2 diabetes care in family medicine. Tartu, 2009.
- 160. Silver Türk. Etiopathogenetic aspects of chronic prostatitis: role of mycoplasmas, coryneform bacteria and oxidative stress. Tartu, 2009.
- 161. **Kaire Heilman.** Risk markers for cardiovascular disease and low bone mineral density in children with type 1 diabetes. Tartu, 2009.
- 162. Kristi Rüütel. HIV-epidemic in Estonia: injecting drug use and quality of life of people living with HIV. Tartu, 2009.