LIISA KUHI

A contribution of biomarker collagen type II neoepitope C2C in urine to the diagnosis and prognosis of knee osteoarthritis





LIISA KUHI

A contribution of biomarker collagen type II neoepitope C2C in urine to the diagnosis and prognosis of knee osteoarthritis



Department of Internal Medicine, Institute of Clinical Medicine, University of Tartu, Estonia

Dissertation accepted for the commencement of the degree of Doctor of Philosophy in Medicine on June 15th, 2022 by the Council of the Faculty of Medicine, University of Tartu, Estonia.

Supervisors: Kalle Kisand, MD, PhD, Professor

Department of Internal Medicine, Institute of Clinical

Medicine, University of Tartu, Estonia

Agu Tamm, MD, PhD, DMSci, Professor emeritus Department of Internal Medicine, Institute of Clinical

Medicine, University of Tartu, Estonia

Reviewers: Katre Maasalu, MD, PhD, Associate Professor

Department of Traumatology and Orthopaedics, Institute of Clinical Medicine, University of Tartu,

Estonia

Raili Müller, MD, PhD, Lecturer

Department of Internal Medicine, Institute of Clinical

Medicine, University of Tartu, Estonia

Opponent: Stefan Lohmander, MD, PhD, Senior Professor

Orthopedics, Deptarment of Clinical Sciences Lund,

Lund University, Sweden

Commencement: August 29th, 2022

ISSN 1024-395X ISBN 978-9949-03-923-4 (print) ISBN 978-9949-03-924-1 (pdf)

Copyright: Liisa Kuhi, 2022

University of Tartu Press www.tyk.ee

TABLE OF CONTENTS

LIST OF PUBLICATIONS 8				
Αŀ	BBREVIATIONS	9		
1.	INTRODUCTION	11		
2.	REVIEW OF THE LITERATURE	14		
	2.1. Epidemiology of OA	14		
	2.2. Pathogenesis of OA	15		
	2.2.1. Role of obesity and meta-inflammation	17		
	2.2.2. Role of biomechanics	18		
	2.2.3. Role of aging	19		
	2.3. Pathological changes in the joint tissues	20		
	2.3.1. Articular cartilage	20		
	2.3.2. Subchondral bone	22		
	2.3.3. Synovium	23		
	2.3.4. Menisci and ligaments	23		
	2.3.5. Cross-talk between the joint tissues in OA	24		
	2.4. Phenotypes of OA	24		
	2.5. Classification criteria of kOA	25		
	2.5.1. Imaging in the diagnosis of kOA	26		
	2.5.2. Patient-reported outcome measures in kOA	29		
	2.6. Course of kOA	29		
	2.6.1. Pre-kOA	30		
	2.6.2. Early-stage kOA	30		
	2.6.3. Established kOA and progression of the disease	31		
	2.6.4. End-stage kOA	32		
	2.7. OA biomarkers: renewed approach	33		
	2.7.1. Inflammatory markers	36		
	2.7.2. Bone remodelling markers	37		
	2.7.3. Cartilage markers	37		
	2.8. Summary of the literature review	41		
3	AIM AND HYPOTHESES	43		
		44		
4.	MATERIALS AND METHODS	44 44		
	4.1. Study subjects	44		
	4.2. Urinary collagen type-II C-terminal cleavage neoepitope	47		
	(uC2C) measurement	47		
	4.3. Standardized radiographic investigation	48		
	4.3.1. Definitions of radiographic progression and distribution of	<i>E</i> 0		
	the progression groups (Paper II)	50		
	4.4. Evaluation of articular cartilage lesions (Paper I)	52		
	4.5. Patient-relevant outcome questionnaires	52		
	4.5.1. Knee Injury and Osteoarthritis Outcome Score (KOOS)	52		

		4.5.2. Visual analog scale for pain (VASpain) in joints of different
		skeletal areas
		Statistical analysis
	4.7.	Ethics
5.	RES	ULTS
		Association of uC2C and kOA radiographic features (Paper I
		and III)
		5.1.1. Prevalence of radiographic features according to clinical
		study groups
		5.1.2. Association of uC2C level with gOA
		5.1.3. Association of uC2C level with TFOA and PFOA
		5.1.4. Association of uC2C level with Ophs, JSN, and the SFA
		score
		5.1.5. Sex-related differences in associations with uC2C and kOA features
	5.2	uC2C level as a marker of progression of kOA (Paper II)
	٠.۷.	5.2.1. Clinical characteristics of the longitudinal study group at
		baselinebaseline baseline
		5.2.2. Association of uC2C at baseline level with the three-year
		follow-up of kOA progression
		5.2.3. A prognostic value of uC2C for kOA progressors in distinct
		gOA
		5.2.4. Sex-related differences between associations of uC2C and
		kOA progression
	5 3	uC2C and total knee replacement (Paper III)
	5.5.	5.3.1. The dynamics of uC2C levels after TKR
		5.3.2. Sex-related differences in the dynamics of uC2C levels after
		•
	5 1	TKRuC2C level and patient's self-assessment outcomes (Paper III)
	J.4.	5.4.1. Associations between uC2C and pain
		•
		5.4.2. An association of uC2C levels with the improvement of
		KOOS subscales in the post-TKR
6.	DISC	CUSSION
	6.1.	uC2C: a risk and an early diagnostic biomarker of
		radiographic kOA
	6.2.	uC2C: an early integrative biomarker of different kOA
		features in distinct joint compartments
	6.3.	uC2C: a biomarker to monitor a severity of disease
		uC2C: a prognostic biomarker of kOA radiographic progression
		uC2C: a biomarker to monitor pre-TKR and post-TKR
		response
	6.6.	uC2C: a predictive biomarker for clinical outcome of TKR
		Sex-related differences of uC2C level in the course of kOA
		Limitations of the study

7. CONCLUSIONS	83
8. REFERENCES	85
9. SUMMARY IN ESTONIAN	114
10. ACKNOWLEDGEMENTS	119
PUBLICATIONS	121
CURRICULUM VITAE	172
ELULOOKIRJELDUS	175

LIST OF PUBLICATIONS

- I Kuhi, Liisa; Tamm, Ann E.; Tamm, Agu O.; Kisand, Kalle (2020). Cartilage collagen neoepitope C2C in urine as an integrative diagnostic marker for early knee osteoarthritis. Osteoarthritis and Cartilage Open, Volume 2, Issue 4, December 2020, 100096. DOI: 10.1016/j.ocarto.2020.100096.
- II Kuhi, L.; Tamm, A.E.; Tamm, A.O.; Kisand, K. (2021). Risk Assessment of the Progression of Early Knee Osteoarthritis by Collagen Neoepitope C2C: A Longitudinal Study of an Estonian Middle-Aged Cohort. Diagnostics, 11 (7), ARTN 1236. doi:10.3390/diagnostics11071236
- III Kuhi, Liisa; Tamm, Ann E.; Jaanika Kumm J.; Kristel Järv, K., Märtson, A.; Tamm, Agu O.; Kisand, Kalle (2022). Associations of urinary collagen II neoepitope C2C with total knee replacement outcomes: is OA a systemic disease in rapidly progressive cases? Applied Sciences, 12, 164. doi: 10.3390/app12010164.

Author's contribution:

- Paper 1 and 2: conceptualization, subject and sample selection, performing of statistical analysis, visualization, writing the paper
- Paper 3: conceptualization, subject and sample selection, performing the ELISA tests, performing of statistical analysis, visualization, writing the paper

ABBREVIATIONS

ACR the American College of Rheumatology

AKOA accelerated knee osteoarthritis

AUC area under the curve

BL baseline

BMI body mass index CI confidence interval Col2 type II collagen

COMP cartilage oligomeric matrix protein

Crea creatinine in urine

DMOAD disease modifying OA drug EKOA early knee osteoarthritis

EULAR European League against Rheumatism

GAG glycosaminoglycan GLM generalized linear model GP general practitioners

gOA global grade of radiographic kOA by Nottingham system for

classification

IL interleukin

JSN joint space narrowing JSW joint space width

KL radiographic knee osteoarthritis by Kellgren and Lawrence

scoring system for classification

kOA knee osteoarthritis

KOOS The Knee injury and Osteoarthritis Outcome Score

KOOSsympt KOOS subscale of knee symptoms KOOSpain KOOS subscale of knee pain

KOOSadl KOOS subscale of activities of daily living KOOSsp/recr KOOS subscale of sport and recreation

KOOSqol KOOS subscale of knee-related quality of life

LM linear regression model
MJOA multiple joint osteoarthritides
MMP matrix metalloproteinase
MRI magnet resonance imagine

n numbers

NSy radiographic knee osteoarthritis by Nottingham scoring system

for classification

OA osteoarthritis

OARSI Osteoarthritis Research Society International

Oph osteophyte
OR odds ratio
PF patellofemoral

PROM patient-reported outcome measure

rkOA radiographic knee osteoarthritis SCB substantial clinical benefit

SD standard deviation SF synovial fluid

SFA (medial) Société Française d'Arthroscopie score

sHA serum hyaluronic acid

sumVAS summary VAS pain of different joints

TF tibiofemoral

TKR total knee replacement TNF-α tumor necrosis factor alpha

uC2C collagen type II C-terminal cleavage neoepitope C2C in urine

uCTX-II C-terminal telopeptide of type II collagen in urine

VAS pain Visual Analog Scale for Pain

WOMAC Western Ontario and McMaster Universities osteoarthritis index

X-ray radiographic examinatiom

Yrs years

1. INTRODUCTION

Osteoarthritis (OA) is the most frequent musculoskeletal disorder and the single most common cause of disability in elderly people; therefore, the global impact of this chronic disease is a major worldwide challenge for healthcare systems in the 21st century (Allen *et al.* 2021). In 2019, OA affected population was ~500 million people worldwide, which was about 7% of the global population. (Vos *et al.* 2020), and the incidence of the disease is increasing annually by approximately 0.3% (Jin *et al.* 2020). OA amounts a tremendous burden globally, especially in aging and increasingly obese populations (Hunter *et al.* 2020). In 2015, the overall cost of OA represents 1.1% of the United States' gross domestic product (GDP), making it the second most expensive disease (Zhao *et al.* 2019). It should be underlined that OA affects not only pain and physical function but also has many other outcomes such as a decline in mental health, quality of life, and even mortality from cardiovascular disease (CVD) (Veronese *et al.* 2016).

Although OA was first discovered among Palaeolithic hunter-gatherers, an approximate doubling of knee OA (kOA) prevalence has occurred since the mid-20th century (Berenbaum *et al.* 2018; Wallace *et al.* 2017). This tremendous rise in the prevalence of OA can be attributed at least in part to the increase in known OA risk factors, including longevity, obesity, physical inactivity, and joint injury. From 2000 to 2013, global all-age obesity increased by 26%, and the proportion of adults with low physical activity grew by 20%. However, these factors are insufficient to explain the increase in OA prevalence indicating that the additional, yet unexplained, risk factors are contributing (Hawker, 2019; Berenbaum *et al.* 2018). The analysis has shown that by 2032, more than 26,000 new OA consultations per 1,000,000 people aged ≥45 will take place mainly in the primary care system, therefore a potential "tsunami" in OA cases is expected (Turkiewicz *et al.* 2014; Mobasheri *et al.* 2015).

OA may affect any synovial joint; however, knees, hips, hands (carpometacarpal joints, distal or proximal interphalangeal joints), and spine are most commonly involved. A prevalence of radiographic OA in the knee joint ranges from 15 to 76% depending on investigated population (D. T. Felson *et al.* 1987; Dillon *et al.* 2006; Jordan *et al.* 2007; Turkiewicz *et al.* 2014; Blanco *et al.* 2021). Worldwide, a pooled global prevalence of kOA is 22.9% in people aged 40 years and over, while a pooled global incidence was 203 per 10000 personyears in a recent review (Cui *et al.* 2020). In Estonia, the prevalence of kOA was 6.6% in 2019 (Vos *et al.* 2020). However, the prevalence of kOA is most likely underestimated worldwide, including in Estonia, as the disease in the patellofemoral (PF) joint is usually overlooked. Moreover, there is an evidence that knee pain and severity of radiographic OA features do not correlate (Bedson & Croft, 2008). Therefore, the assessment of OA prevalence depends significantly on the used diagnostic criteria. All these facts reveal that OA is a comprehensive and expensive disorder that puts stress on the health and social

care systems across the globe. Despite its considerable social-economic toll, OA was generally neglected until the year 2000 (Veronese *et al.* 2016). There is an enormous unmet need to avoid this disability. The Bone and Joint Decade 2000–2010, a global campaign, was arranged to achieve a greater awareness of the growing burden of musculoskeletal disorders, including OA, supported by the United Nations and WHO (Woolf, 2000). In 2016, the Osteoarthritis Research Society International (OARSI), the leading global organization in OA research, submitted a White Paper, Osteoarthritis: A serious disease (https://oarsi.org/education/oarsi-resources/oarsi-white-paper-oa-serious-disease), to raise an understanding of the importance of OA.

Since the year 2000, there has been an extensive research in the field of knee OA in Estonia. Notably, the Estonian middle-age population kOA cohort was created. These subjects were investigated longitudinally using novel methods of radiographic evaluation of kOA in different joint compartments by Nothingam system (Nagaosa et al. 2000), for the duration of three-years, and follow-up of a relevant proportion of subjects lasted as long as 12 years. This large kOA repository has provided an opportunity for extensive research and involvement in several international consortia (e.g. TREAT-OA, NanoDiaRA) to discover the causes and solutions to the complexity of this serious disease (Kerkhof et al. 2011). Several papers have been published based on the data of Estonian cohorts, including the studies on the prevalence and progression of radiographic kOA (rkOA) (Tamm et al. 2008b; Kumm et al. 2012; Kumm et al. 2013a), the studies on rkOA in association with cartilage (Kumm et al. 2006; Kerna et al. 2012; Kumm et al. 2013b), bone (Kumm et al. 2008; Kumm et al. 2013c), genetic markers (Kerna et al. 2009; Valdes et al. 2010a; Valdes et al. 2010b; Kerna et al. 2010; Valdes et al. 2011; Kerna et al. 2013), and cytokines (Kisand et al. 2018), patient-reported assessment and performance-based knee function test (Tamm et al. 2008a; Tamm et al. 2011; Tamm et al. 2012; A.E. Tamm et al. 2014), and the studies on knee ultrasound (Kumm et al. 2009; Kumm et al. 2010).

Years ago, OA was characterized as a degenerative joint disease, or hypertrophic arthritis affecting an irrelevant proportion of the population (Katz *et al.* 2021; Mobasheri *et al.* 2021). However, the approach to OA as a single entity of cartilage damage changed greatly in recent decades. In 1986, the American of College Rheumatology (ACR), suggested the definition of OA as, 'a heterogeneous group of conditions that lead to the joint symptoms and signs which are associated with the defective integrity of articular cartilage, in addition to related changes in the underlying bone at the joint margins' (Hutton, 1987).

Nowadays, OA is considered to have a complex pathophysiology affecting multiple joint tissues from molecular to structural level (Katz *et al.* 2021; Lv & Shi, 2021). In 2015, OARSI proposed a new definition of OA as, 'the disease manifests first as a molecular derangement (abnormal joint tissue metabolism) followed by anatomic, and/or physiologic derangements (characterized by cartilage degradation, bone remodelling, osteophyte formation, joint inflammation

and loss of normal joint function), that can culminate in illness.' (Kraus et al. 2015).

Currently, there is no cure for OA; this means, we do not know how to prevent the disease or to reverse its pathogenic processes (Hunter & Bierma-Zeinstra, 2019). However, some progress in the development of disease-modifying OA drugs (DMOADs) can be noticed, for example, the preliminary evidence of substantial structure-protective action of fibroblast growth factor 18 has been established (FGF-18, named Sprifermin) (Oo et al. 2021). OA management is targeted to relieve the symptoms, primarily pain. Total knee replacement (TKR) at the terminal-stage of the disease is expensive and does not always give the expected results. Several important challenges such as a limited understanding of the OA joint biology, etc. have slowed the development of OA treatment, and as a result, it is considerably lagging as compared to the advances in other rheumatic diseases (Mobasheri et al. 2021). Moreover, recently it is recognized that OA can have several endotypes (subtypes) with certain differences in the pathogenesis, risk factors (including genetics), and the disease trajectory (Mobasheri et al. 2019). The three main molecular or mechanistic OA endotypes are supposed to be cartilage-driven, synovitis-driven, and bonedriven endotypes (Oo et al. 2021). It has been proposed that the best "window of opportunity" exists in the early phase of the disease before a serious remodelling of the joint tissues (Mahmoudian et al. 2021). However, reliable biomarkers for the early diagnosis of OA patients are needed for appropriate surrogate endpoints in clinical trials.

The new definition of OA highlights a molecular derangement as a primary disorder in the pathogenesis of this disease; therefore, the detection of early biomarkers is paramount in searching for more sensitive diagnostic possibilities in kOA. In the current study, we investigated a urinary biomarker for OA, type II collagen (Col2) cleavage neoepitope (uC2C) as a potential biomarker of kOA for diagnosis, prognosis, and outcome throughout the disease progression.

2. REVIEW OF THE LITERATURE

2.1. Epidemiology of OA

OA is one of the most frequent chronic progressive disorders in the world that causes a notable disease burden in middle-aged and elderly population (Turkiewicz et al. 2014; Jin et al. 2020; Allen et al. 2021). The incidence of OA rises precipitously with age and tends to be higher in females, especially after the age of 50 years (Cui et al. 2020; Prieto-Alhambra et al. 2014). However, the estimated OA prevalence varies depending on the definition (radiographic, symptomatic, etc.) and severity used to categorize the disease (Bedson & Croft, 2008), as well as the characteristics of study population taken into consideration (age, sex, ethnicity, genetics, diet, overweight/obesity, smoking, physical activity, and joint injuries) (Hunter & Bierma-Zeinstra, 2019), for example, in the Framingham Osteoarthritis Study, the prevalence of radiographic OA found elevated from 33% among the subjects aged 60–70 years to 44% among those aged more than 80 years (Felson et al. 1987). The prevalence of OA in Estonia as estimated in the year 2019 is 150 800 cases (12% of all causes), including the prevalence of kOA (88100 cases, 7% of all causes) (Vos et al. 2020).

There is limited data on the prevalence of kOA in a population younger than 50 years. In a Dutch study, rkOA (grade 2 or higher) was found in 8% of men and 12.5% of females aged 45–49 years (van Saase *et al.* 1989). Similarly, chronic knee pain (> 3 months) was reported by 15% of people aged 35–54 years in Sweden, however, rkOA was diagnosed in only about 10% of them (1.5% prevalence in the age group) (Petersson *et al.* 1997). A small population study in South Estonia demonstrated that knee problems are common among the Estonian middle-aged population (34–55 years of age). 60% of the participants reported knee pain and 40% other knee symptoms. However, early rkOA (grade 1 according to the Nottingham system (NSy) (Nagaosa *et al.* 2000)) was found in 55.6% and grade 2 or 3 in 8.1% of the subjects, resulting in the prevalence of 46% and 3.7%, respectively (Tamm *et al.* 2008b).

Functionally, the knee includes 2 articulations – tibiofemoral (TF) and patellofemoral (PF) joints which share the common capsule, making the knee the largest synovial joint (Brandt *et al.* 2003; Flandry & Hommel, 2011). Although the medial TF compartment is most frequently disturbed, OA is more common in PF joint and occurs even in the absence of the disease in TF joint (McAlindon *et al.* 1992). The meta-analysis of PF OA revealed that overall crude prevalence of OA in PF joint was 25% in the population-based cohorts (aged \geq 20 years) and 39% in the symptom-based cohorts (aged \geq 30 years) (Kobayashi *et al.* 2016). Unlike to the population-based cohorts, women appeared to have a higher prevalence of PF OA (41%) than men (23%) in the symptom-based cohorts. The prevalence of PF OA classified by osteophytes was 48%. A study on the Estonian middle-aged people population-based cohort (438 participants: aged 35–57 years) (Kerna *et al.* 2013) was comprised in this meta-analysis and reported the lowest prevalence estimate (8%) (Kobayashi *et*

al. 2016). Younger cohort could explain the lower prevalence of kOA in the Northern European studies.

The kOA could be unilateral or bilateral, with or without multi-joint involvement (Sharma, 2021). Established and symptomatic kOA is often radiographically bilateral (Ledingham *et al.* 1993; Bihlet *et al.* 2019). The analysis of a 12-year prospective cohort study revealed that 26% of patients had bilateral disease at baseline, whereas this percentage was found increased to 52% at the 5th year and 70% at the 12th year follow-up (Metcalfe *et al.* 2012). The most common pattern was an involvement of medial compartment in both knees (Metcalfe *et al.* 2012). Unilateral and isolated medial TF kOA was more common in men (Ledingham *et al.* 1993). The examination of both knees (and not restricted to only the symptomatic knee) is always indicated, and the different conditions of knees could influence the results of biomarkers.

Involvement of multiple joints in OA is frequent (Nelson *et al.* 2014; Kraus *et al.* 2007). Generalized or multiple joint OA (MJOA), in particular MJOA of hand joints, is more prevalent in women than men (Nelson *et al.* 2014). However, men as compared to women are more likely to have lumbosacral spine OA as reported (Nelson *et al.* 2013). A higher frequency of MJOA or elevated risk of MJOA progression is associated with age (Nelson *et al.* 2014).

Therefore, the analyses of kOA biomarkers should take into account the different knee compartments and possible MJOA.

2.2. Pathogenesis of OA

OA is a heterogeneous disease that results from a combination of mechanical, inflammatory, genetic, and metabolic factors (Johnson & Hunter, 2014; Hunter & Bierma-Zeinstra, 2019). This multifactorial pathogenesis involves a wide range of underlying pathways leading to a common final pathway of joint destruction (Martel-Pelletier *et al.* 2016; Deveza & Loeser, 2018). OA is previously referred to as passive degenerative cartilage-limited disease or so-called 'wear-and-tear' disease (Loeser *et al.* 2012). However, the modern concept describes OA as an active dynamic alteration arising from an imbalance between the repair and destruction of joint tissues (Hunter & Bierma-Zeinstra, 2019). Initially considered cartilage-driven, OA is a much more complex disease with different inflammatory and tissue mediators released by the joint tissues (cartilage, bone, synovium, etc.; Fig. 1) (Berenbaum *et al.* 2017).

There is now a wealth of evidence that inflammation plays an important role in OA pathogenesis (Scanzello, 2017) and the involvement of all branches of immune system is demonstrated (Haseeb & Haqqi, 2013). Inflammation can be triggered within the joint because of tissue damage and stress responses, and obesity-related systemic inflammation might enhance these local responses. Generated low-grade inflammation disturbs tightly regulated anabolic and catabolic processes responsible for the maintenance of cartilage homeostasis (Wojdasiewicz *et al.* 2014).

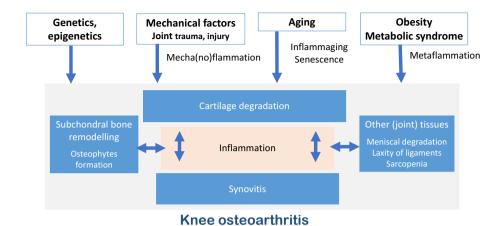


Figure 1. Pathogenic interactions involving different joint tissues in knee osteoarthritis.

Dysregulation of several cytokines and chemokines has been described in the blood and joint tissues of OA patients (Haseeb & Hagqi, 2013; Primorac et al. 2020; Endres et al. 2020). Interleukin-1 beta (IL-1β) and tumor necrosis factor alpha (TNF- α) are the two major synergistically acting pro-inflammatory cytokines in OA causing degradation of the cartilage (Haseeb & Haqqi, 2013). They are also involved in pain generation. IL-1 and TNF-α upregulate the expression of inducible nitric oxide synthase (iNOS), soluble phospholipase A2, cyclooxygenase 2 (COX-2), and microsomal prostaglandin E synthase 1, and also stimulate the release of nitric oxide (NO) and prostaglandin E2 (PGE2) (Kapoor et al. 2011). Moreover, these cytokines increase the expression of metalloproteases (collagenases and aggrecanases) in chondrocytes and synovial fibroblasts and inhibit the synthesis of proteoglycan and Col2 in chondrocytes (Saklatvala, 1986; Goldring et al. 1988; Kobayashi et al. 2005; Fan et al. 2005). In response to IL-1 β and TNF- α , the joint cells release IL-6, which leads to amplifying the catabolic effect (Goldring & Goldring, 2007; Mabey & Honsawek, 2015). In OA, also the other pro-inflammatory cytokines, including IL-15, IL-17, IL-18, IL-21, and leukaemia inhibitory factor (LIF) has been implicated (Kapoor et al. 2011).

Several chemokines are also involved in the OA pathology and were reported to be expressed in synovium or chondrocytes including IL-8/ chemokine (C-X-C motif) ligand 8 (CXCL-8), growth-regulated oncogene α (GRO α)/CXCL-1, monocyte chemoattractant protein-1 (MCP-1)/ chemokine (CC-motif) ligand 2 (CCL-2), regulated upon activation, normal T cell expressed and presumably secreted (RANTES)/CCL-5, macrophage inflammatory protein-1 α (MIP-1 α)/CCL-3, and MIP-1 β /CCL-4 (Borzi et al. 1999). Chondrocytes also express chemokine receptors including C-X-C chemokine receptor 3 (CXCR3), CXCR4, CXCR5, CC chemokine receptor 1 (CCR1), CCR3, CCR5, and CCR6

(Houard et al. 2013). Pro-inflammatory cytokine IL-1 β was reported to stimulate the expression of the chemokines in OA chondrocytes (Akhtar & Haqqi, 2011). In addition, several pro-inflammatory cytokines and adipokines (like leptin and adiponectin) are produced by adipocytes and tissue-infiltrating macrophages in overweight people (Lago *et al.* 2007). Recently, it was shown in the Estonian cohort that an association between angiogenic cytokines and kOA severity exists since a very early phase of the disease. Interestingly, this important pathogenic aspect was predominantly expressed in females (Kisand *et al.* 2018).

2.2.1. Role of obesity and meta-inflammation

Obesity is one of the most clinically significant and modifiable risk factors for OA (Felson *et al.* 1997; Muthuri *et al.* 2011). The mechanisms by which obesity leads to the onset and progression of OA are debatable due to the complex interactions among metabolic, biomechanical, and inflammatory factors that accompany overweight and adiposity (Collins *et al.* 2021).

Traditionally, it has been suggested that weight gain predisposes people to OA simply because of mechanical loading. However, obesity-associated metabolic factors have been found to be associated with a higher incidence of OA in non-weight bearing joints as well (Yusuf *et al.* 2010; Gandhi *et al.* 2012; Visser *et al.* 2014). Therefore, obesity and overweight are now regarded as low-grade systemic inflammatory states, named as 'metaflammation' with elevated inflammatory markers (Lago *et al.* 2007; Choi *et al.* 2013).

During obesity, tissue macrophages adopt a metabolically activated phenotype in response to the altered metabolic environment of white adipose tissue (WAT) in this setting, particularly to an increased concentration of free fatty acids, and high insulin and glucose concentration during an emerging state of insulin resistance. WAT is considered as an endocrine organ secreting a large variety of peptides named as 'adipokines', including leptin, visfatin, adiponectin, resistin, and others (Wang et al. 2015). Adipokines can also be produced by the joint cells including chondrocytes when induced by inflammatory stimuli (Conde et al. 2011). Leptin seems to be a possible link between obesity and OA; high leptin levels are found in the synovial fluid of obese patients. Also, leptin receptor is expressed in cartilage and its sensitivity was found enhanced. Leptin induces the production of matrix metalloproteinases (MMPs), proinflammatory mediators, and nitric oxide (NO) in chondrocytes (Vuolteenaho et al. 2014). A recent study on lipodystrophic mice demonstrated a direct relationship between adipose tissue and cartilage damage, independent of the mechanical aspect of joint loading - implantation of fat tissue to fat-free animals restores the susceptibility to OA (Collins et al. 2021).

In addition to WAT, an infrapatellar (Hoffa) fat pad (IPFP) may act as a modulator in OA (X. Wang *et al.* 2015). IPFP is a very sensitive tissue composed of adipocytes, immune cells (primarily macrophages and lymphocytes), fibroblasts, blood vessels, and collagen matrix (Tu *et al.* 2019). Thus, IPFP

serves both as a local source of adipokine and a local modulator of inflammatory responses contributing to initiation and progression of knee OA (Belluzzi *et al.* 2017). IPFP-derived adipokines exhibit unique patterns of secretion and distribution, imparting a direct impact on articular cartilage degeneration (Richter *et al.* 2015). For instance, except leptin and resistin, the other adipokines are more actively secreted by IPFP than by subcutaneous WAT in the same OA individual (Ioan-Facsinay & Kloppenburg, 2013).

Obesity also has an effect on synovium and it causes synovial adipocyte hypertrophy, macrophage accumulation, fibrosis, and increased expression of TNF-α and toll-like receptor 4 (TLR4) (Hamada et al. 2016; Harasymowicz et al. 2017; Eymard et al. 2017) supporting innate immune signalling via TLR4 in obesity-induced OA (Kalaitzoglou et al. 2019). Some levels of lipopolysaccharides (LPS) are routinely detectable even in the absence of infection, presumably due to disturbances in the gut microbiome of obese patients. However, whether the relationship between LPS and OA severity is directly mediated by TLR4 interaction is unclear (Scanzello, 2017). A number of questions remain about which factors are associated with obesity and directly modulate synovial inflammation. Studies indicate that multiple factors could be involved, including synovial insulin resistance (Hamada et al. 2016), dietary fatty acid composition (Sekar et al. 2017), and gut microbiome composition, which is related to circulating LPS levels, body fat percentage, fat cell apoptosis, and OA manifestations (Collins et al. 2015). To sum up the aspects of fatty tissue, obesity is an important confounding factor in OA biomarker research and should be taken into account in the analysis of clinical groups.

2.2.2. Role of biomechanics

High risk of OA after knee injury demonstrates a crucial role of biomechanical factors in the initiation of the disease in susceptible individuals (Englund, 2010). Although all joint tissues are affected to some degree as a result of joint trauma, injury of articular cartilage appears substantial, as it is largely irreversible and may initiate a subsequent development of OA. The injury drives active mechanosensitive intracellular signalling which modulates the biochemical activity of chondrocytes by the process called 'mechanotransduction' (Primorac et al. 2020). Two principal pathways are proposed – the one that can lead to the release of growth factors from the matrix and stimulate repair; the another can trigger an inflammatory response to mechanical injury, referred also as 'mechanoflammation' (Vincent, 2019). The upstream activator of mechanoflammation remains unknown, but it involves the activation of several transcription factors controlling the expression of target genes such as MMP13, NOS2, COX2, ADAMTS, and IL1B (Houard et al. 2013). Repair-promoting pathways appear to be largely driven by the release of growth factors such as transforming growth factor β (TGFβ) and fibroblast growth factor 2. Both the factors are sequestered in the pericellular matrix of cartilage and released immediately in response to injury (Vincent, 2013).

A precise relationship between mechanoflammation and cartilage repair is currently unclear but it is likely that chronic mechanoflammation contributes to the disease by suppressing intrinsic tissue repair (Vincent, 2019) and changing a balance between pro-degenerative and pro-repair pathways.

2.2.3. Role of aging

Aging of the joint tissues increases the incidence of OA substantially but aging and OA are two independent processes (Loeser et al. 2016). While OA is not an inevitable consequence of aging, aging-related changes in the joint tissues contribute to OA development along with some other risk factors (Anderson & Loeser, 2010). Aging is characterized by changes in metabolic and mitotic activity and decreased sensitivity of chondrocytes to growth factors such as TGF-β signalling (van der Kraan & van den Berg, 2008). Common biological changes seen in senescence are genomic instability, telomere attrition, epigenetic alterations, loss of proteostasis, deregulated nutrient-sensing, mitochondrial dysfunction, cellular senescence associated with cell cycle arrest, and unresponsiveness to mitogenic stimuli, stem cell exhaustion, and altered intercellular communication (López-Otín et al. 2013). 'Chondrosenescence' is defined as an age-dependent deterioration of chondrocyte function (Mobasheri et al. 2015). Aging-related changes in the cartilage matrix, including formation of advanced glycation end-products (AGEs) and development of the senescence-associated secretory phenotype (SAPS) of chondrocytes with the production of plethora of soluble signalling factors (inflammatory cytokines and chemokines, growth and angiogenic factors, proteases, bioactive lipids, matrix metalloproteinases) are considered to be involved in OA pathogenesis (Anderson & Loeser, 2010). Aging and inflammation are major contributing factors to the development and progression of arthritic diseases (Mobasheri et al. 2015). 'Inflammaging' is a term for describing low-grade chronic inflammation that occurs during physiological aging as a consequence of the adaption and counteraction to the different stressors, i.e, lifelong antigenic burden (Franceschi & Bonafè, 2003). An increase in fat mass, aging-related metabolic changes, and epigenetic regulation of age-dependent gene expression can also result in inflammaging (Loeser et al. 2016; M. Zhang et al. 2019). This condition is suggested to be a result of an imbalance between inflammatory and anti-inflammatory networks (Franceschi et al. 2007). In chondrosenescence a proliferative and synthetic capacity of cells decreases, while a production of pro-inflammatory mediators and matrix-degrading enzymes is maintained (Loeser, 2009). Inflammaging and chondrosenescence are intimately linked with each other in the osteoarthritic joint contributing to a decrease in the efficacy of articular cartilage repair (Mobasheri et al. 2015). Non-enzymatic crosslinking of collagen by AGEs that occurs with aging alters mechanical properties of cartilage, and the resulting changes to mechanotransduction pathways reduce extracellular matrix synthesis by chondrocytes (Loeser et al. 2016). With normal joint aging, articular cartilage remains intact but loses its

thickness and glycosaminoglycan (GAG) content, however in OA, fibrillation of the cartilage surface occurs in focal areas and can be associated with a complete loss of GAG (Lotz & Loeser, 2012).

In conclusion, several hallmarks of senescence are associated with OA. However, it remains uncertain that which factors and mechanisms contribute to the distinct OA phenotypes and in particular to the disease progression (Coryell *et al.* 2021).

2.3. Pathological changes in the joint tissues

OA is a whole joint disease, involving structural alterations in the hyaline articular cartilage, subchondral bone, ligaments, capsule, synovium, and periarticular muscles (Hunter & Bierma-Zeinstra, 2019; Martel-Pelletier *et al.* 2016). Despite the multifactorial nature of OA, the pathologic changes seen in osteoarthritic joints have common features that affect the entire joint structure, resulting in pain, deformity, and loss of function (Loeser *et al.* 2012).

2.3.1. Articular cartilage

Normal articular cartilage is avascular, alymphatic, and aneural connective tissue which is composed of chondrocytes and extracellular matrix (ECM) (Martel-Pelletier *et al.* 2008). ECM consists of water (65–80 % of the wet mass), inorganic salts, and organic components in which collagens (15–22% of the cartilage wet weight) and proteoglycans (aggrecans) are the main macromolecules (Martel-Pelletier *et al.* 2008; Xia *et al.* 2014). All cartilage components and proteolytic enzymes are synthesized by chondrocytes, providing a minimal and balanced cartilage turnover between anabolic and catabolic processes during the life (Goldring & Marcu, 2009). In recent years, it has been reported that mature articular cartilage contains a small population of mesenchymal stem cells (MSC)-like progenitors that are capable of differentiating into mature chondrocytes. Furthermore, these cells exist in greater numbers in OA cartilage than in normal cartilage (Jayasuriya *et al.* 2018).

The unique properties of cartilage are related to the composition and structure of its components, which are highly ordered from the cartilage surface to its deepest layers. Cartilage consists of four zones with different architecture and functions – superficial, middle, deep, and calcified cartilage zones (Martel-Pelletier *et al.* 2008). The calcified zone is separated from the unmineralized upper cartilage layers by a histologically defined zone called 'tidemark', which separates the cartilage from the underlying subchondral bone (Martel-Pelletier *et al.* 2016; Goldring & Goldring, 2016). The layers are characterized by chondrocyte shape and positioning, as well as collagen fibrils' orientation (Korhonen *et al.* 2008).

Collagen fibrils compose a network providing a shape and form that provides tensile stiffness and strength to the cartilage. Fibril diameters vary from

20 nm in the superficial zone to 70–120 nm in the deep zone. Col2(90–98% of the total tissue collagen) is specific to cartilage (Martel-Pelletier *et al.* 2008). Majority of the cartilage proteoglycans are in complex supramolecular aggregates (aggregans), which consist of a centrally placed/ satiated hyaluronic acid (HA) filament and multiple monomers are attached to it. The monomers are made up of negatively charged GAG molecules (polymeric saccharides chondroitin-4-sulfate, chondroitin-6-sulfate, and keratan sulfate) bound to the aggrecan core protein (Goldring & Goldring, 2016). Aggregans, together with other matrix components, are entrapped in a collage network. GAG molecules are responsible for osmotic properties of the cartilage matrix.

In OA, the earliest changes in the cartilage appear at the joint surface areas, where mechanical forces, in particular shear stress, are greatest (Andriacchi et al. 2004). One of the initial changes is an increase in water content in the superficial zone of articular cartilage, with aggrecan degradation, presenting as a superficial cartilage fibrillation (Pratta et al. 2006). As OA advances, a process known as 'matrix swelling' expands to the deep zone (Goldring & Goldring, 2010). Also, chondrocytes become 'activated', characterized by cell proliferation, cluster formation, and increased production of both matrix proteins and matrix-degrading enzymes (Goldring & Marcu, 2009; Houard et al. 2013). The matrix-degrading enzymes found in OA joints include members of ADAMTSs (a disintegrin and metalloproteinase with thrombospondin motifs) and MMP families (Loeser, 2006; Troeberg & Nagase, 2012). Matrix degradation in early OA may be due to ADAMTS-5 (aggrecanase) which degrades aggrecan (Stanton et al. 2005), followed by increased activity of collagenases, MMP-1, MMP-8, MMP-13 (Loeser et al. 2012). MMP-13 (collagenase 3) is highly efficient in the cleavage of Col2 participating in cartilage degradation (Mitchell et al. 1996; Houard et al. 2013). The collagen fragments (gelatins) can be denatured by further cleavage by gelatinases (MMP-2 and MMP-9) (Martel-Pelletier et al. 2008).

Once the collagen network is degraded, it reaches the irreversible state (Loeser et al. 2012). Degenerative changes lead to a diminished cartilage thickness and fissuring, which could be observed by arthroscopy. On the other side, transformation in the cellular composition along with the cartilage calcification takes place. This process involves osteochondral angiogenesis, the penetration of calcified cartilage by vascular elements and sensory nerve fibres that extend from the subchondral bone (Walsh et al. 2007). These processes could be associated with the development of chronic pain. All these tissue transformations result in duplication of the tidemark and advancement of the calcified cartilage into the deep zone of articular cartilage, leading to local cartilage thinning (Loeser et al. 2012). These changes become more pronounced with time, when articular cartilage reaches total destruction, eventually leaving the underlying subchondral bone plate completely exposed (Goldring & Goldring, 2007). The advanced signs of cartilage damage are seen by imaging and arthroscopic methods but identification of early signs and sensitive monitoring of

dynamics can be provided by soluble biomarkers (see section 2.7. OA biomarkers).

2.3.2. Subchondral bone

Subchondral bone is divided into two layers: a plate-like layer of cortical bone beneath the calcified cartilage, also known as a 'subchondral bone plate', and a deeper layer of subchondral trabecular or cancellous bone with bone marrow space (Li et al. 2013; Funck-Brentano & Cohen-Solal, 2015). The structure of subchondral bone is mostly dependent on two types of cells —osteoblasts and osteoclasts. Osteoblasts synthesize new bone, while osteoclasts resorb the old (Funck-Brentano & Cohen-Solal, 2015). The structural and functional properties of subchondral bone are defined by a composition of an organic bone matrix and mineral content that represent a dynamic adaptation to biomechanical factors as well as the effects of biochemical factors. A state of bone mineralization is highly dependent on the rate of bone remodeling (Day et al. 2001; Donnelly et al. 2010). Higher rate of bone remodelling is associated with a state of relative hypomineralization. Subchondral bone adapts its architecture and structure more rapidly than the cartilage in order to respond to the changes in the mechanical environment via cell-mediated processes of modelling (Goldring & Goldring, 2007).

The OA related changes in the volume and density of subchondral bone are caused by previous trauma or excessive load (Frost, 2003). Bone remodelling may be initiated at the sites of local bone damage resulting from excessive repetitive loading. In early OA, alterations of bone turnover initiate a deterioration of subchondral trabecular bone such as bone marrow lesions (BML), cysts, and osteophyte (Oph) formation (Goldring, 2008; Burr & Gallant, 2012). BML represents a micro-damage to subchondral bone and is characterized by marrow fibrosis at various stages of healing, fat necrosis, and a local increase in bone remodelling that results in microfractures of the trabecular bone (Taljanovic et al. 2008; Driban et al. 2012). As a putative additional skeletal adaptive mechanism, Ophs are formed at the joint margins via endochondral ossification (Loeser et al. 2012). Oph formation is also indicated as a result of abnormal healing response of subchondral trabeculae, or blood vessels and nerve fibres' penetration into the degrading cartilage (Gilbertson, 1975). A growth factor TGF-β is involved in the Oph formation (Uchino et al. 2000). Although Oph remains controversial regarding their functional role; they may serve to stabilize the joint rather than to contribute to OA progression (van der Kraan & van den Berg, 2007). Anyway, Ophs are simply recognizable features by imaging methods that can help to identify OA progression in the studies of biomarkers' classification.

Subchondral bone cysts development, a hallmark of the advanced OA, depends on osteoclast-mediated bone resorption, a process initiated by bone damage and necrosis at sites of former BMLs (Crema *et al.* 2010). In established OA, the subchondral bone plate increases in volume and thickness. The

sclerosis of subchondral trabecular bone is a characteristic of terminal OA (Burr & Gallant, 2012).

2.3.3. Synovium

Synovium is a specialized connective tissue that lines diarthrodial joints, surrounds tendons, and forms a lining of bursae and fat pads (Mathiessen & Conaghan, 2017). It is responsible for the maintenance of synovial fluid (SF), which is a source of nutrients for cartilage. SF is also a reservoir for cartilage degrading products and is a valuable material for OA studies (Sellam & Berenbaum, 2010).

Synovial inflammatory infiltrates are found in many OA patients in ultrasound or magnet resonance imagine (MRI) investigations (Loeser *et al.* 2012). Macrophages and T-cells are the most predominant immune cells in OA synovium, whereas mast cells, B cells, and plasma cells are also noticed (Haseeb & Haqqi, 2013). Histological pattern of the synovium in OA patients is characterized by hyperplasia, sub-lining fibrosis, and stromal vascularization (Scanzello & Goldring, 2012). Angiogenesis in the synovium is closely associated with chronic synovitis and is observed at all stages of OA. Synovitis can occur even in early OA. Specific aspects of inflammation, such as a higher number of infiltrating macrophages (Benito *et al.* 2005) and co-location of inflamed synovium and cartilage degradation areas, were also noticed (Ayral *et al.* 2005). However, the prevalence of synovitis increases with the disease progression (Scanzello *et al.* 2011; Krasnokutsky *et al.* 2011), and synovial inflammation is diffuse in the late stage (Ene *et al.* 2015).

2.3.4. Menisci and ligaments

Pathologic changes in the menisci include matrix disruption, fibrillation, cell clusters, calcification, and cell death (Katsuragawa et al. 2010; Pauli et al. 2011). Degeneration of menisci is initiated within the tissue substance rather than the surface. Tissue fibrillation and disruption are first seen at the inner rim, which spreads into articular surfaces of the meniscus over time and progresses to a total disruption or loss of meniscus tissue mainly in the avascular zone (Pauli et al. 2011). Col1 content decreases gradually from the surface to the middle and the deep zone of osteoarthritic meniscus (Sun et al. 2012). Unlike Col1, a decrease in Col2 content is severe in the surface zone, and also prominent in the middle and deep zones of osteoarthritic meniscus (Sun et al. 2012). In turn, proteoglycan content increase in osteoarthritic menisci, when compared to normal menisci (Sun et al. 2012). All these intrameniscal changes correlated with perimeniscal synovitis contributing to the degeneration and reduction in the tensile strength of the meniscus (Grainger et al. 2007). The meniscus is rarely able to withstand loading and force transmission during normal movements of the joint, further leading to degenerative tear in it (Bhattacharyya et al. 2003). Moreover, a meniscal tear can be considered as the

first feature of emerging OA (Englund *et al.* 2012). An increased vascular penetration accompanied by increased sensory nerve densities has been noted in OA menisci and could be associated with pain (Ashraf *et al.* 2011).

Degenerative changes are commonly seen in the ligaments in the late stage of OA. Similar to the meniscus, histologic changes include matrix disruption, collagen fibres disorganization, and mucoid degeneration (Hasegawa *et al.* 2012).

2.3.5. Cross-talk between the joint tissues in OA

Synovial joint acts as a functional unit, where all parts support each other (Goldring & Goldring, 2016). The synovium and subchondral bone help chondrocytes with nutrition, as articular cartilage has no direct supply (Scanzello & Goldring, 2012). Because of relatively slower turnover rate (as compared to articular cartilage), subchondral bone undergoes more rapid modelling and remodelling to respond to the changes in the mechanical environment (Goldring, 2012). Furthermore, in the knee joint, meniscal lesions and injuries decrease resistance to mechanical forces, often leading to the structural progression of the disease (Englund *et al.* 2012). A molecular crosstalk between cartilage and bone increases with OA progression and newly formed vessels together with nerves infiltrate the subchondral bone and invade the overlying cartilage tissue, creating a communication channel for an exchange of biologic factors such as cytokines (Funck-Brentano & Cohen-Solal, 2015).

A fact remains controversial if initial structural and composition alterations related to OA, first take place in the bone or articular cartilage (Goldring, 2012). The study by Yang et al. involving a rat model found that the changes in the subchondral bone precedes cartilage degeneration, while cartilage changes were noted before the subchondral bone changes in the collagenases induced model (Yang et al. 2020). It is also unclear whether the morphological changes that occur in osteoarthritic synovial membrane are primary or secondary (Sutton et al. 2009). Synovitis is believed to be induced at first by the ECM degradation products and soluble cartilage-specific neo-antigens, as well as other factors including microcrystals and abnormal mechanical stress (Sellam & Berenbaum, 2010). These components are released into synovial fluid and phagocyted by synovial lining macrophages, triggering an immune response with the synthesis of mediators. The whole process creates a vicious circle, with increased cartilage degradation, subsequently producing more inflammation and pain (Berenbaum, 2013). Although inflammation may not be the initiator of disease, at some point it becomes a driver of disease progression.

2.4. Phenotypes of OA

OA is well accepted as a multifaceted disease and of heterogeneous nature; therefore, it can be considered a complex syndrome rather than a single disease (Deveza & Loeser, 2018; van Spil *et al.* 2020). However, there are many

discrepancies about OA phenotypes as they could be created for different purposes, such as treatment choice or prognosis prediction (Bierma-Zeinstra & van Middelkoop, 2017). Another proposal is to differentiate clinical phenotypes and molecular/mechanistic endotypes (Mobasheri et al. 2019). However, the task is rather complicated as the same patient may have overlapping phenotypes and endotypes of OA (Mobasheri et al. 2019a; Mobasheri et al. 2019b). Clinical phenotype could be defined as a subgroup of patients with similar clinical characteristics (Mobasheri et al. 2019a; Dell'Isola et al. 2016). The systematic review by Dell'Isola *et al.* (2016) proposed six clinical phenotypes of OA as – inflammatory, metabolic syndrome, bone and cartilage metabolism, malaligned biomechanical, minimal joint disease, and chronic pain (Dell'Isola et al. 2016). Molecular endotypes congregate cases with distinct pathophysiological mechanisms and/or molecular signalling pathways (Mobasheri et al. 2019a). Also, different mechanistic osteoarthritis endotypes have been described, for example, inflammatory (local and systemic), metabolic syndrome driven, aging driven, mechanical injury driven, endocrine (oestrogen deficiency), etc. (Mobasheri et al. 2019b; Henrotin, 2021). Recently, in an international project the provision of consensus-based definitions and recommendations was proposed (van Spil et al. 2020). In this project, OA phenotypes were defined as subtypes of OA that share distinct underlying pathobiological, pain mechanisms, and their structural and functional consequences. OA phenotypes are a very promising but challenging area of research that should determine the underlying mechanisms and discover a combination of sensitive biomarkers for differentiation of distinct OA phenotypes already in the molecular stage of disease (Mobasheri et al. 2019a; Van Spil et al. 2019).

2.5. Classification criteria of kOA

The most often used criteria for established kOA are the American College of Rheumatology (ACR) criteria for the classification of OA (Altman *et al.* 1986) and the European League against Rheumatism (EULAR) recommendations for diagnosis (W. Zhang *et al.* 2010). Both the criteria define kOA by a combination of clinical symptoms and radiographic findings. It is important because of the discordance between symptoms and radiography, although these discrepancies decrease with more severe radiographic disease (Duncan *et al.* 2007). Classification criteria aim for differentiating kOA patients from those with other arthritic diseases to achieve a homogeneous kOA patient group for research to assess the effect of novel treatment or appropriateness of biomarkers (Aggarwal *et al.* 2015).

Due to the frequent discrepancies between symptoms and radiographic findings in kOA, a combined use of both, imaging and clinical findings in the diagnosis of kOA is justified to achieve higher specificity for kOA and to get uniform kOA severity groups (Anderson & Loeser, 2010; Kraus *et al.* 2011). Moreover, imaging features and patient-reported outcome measures (PROMs)

are proposed as reliable outcome measures for OA patients, wherein: PROMs are used for the assessment of knee complaints and physical function, while imaging can be used for the detection of the structural progression of kOA (Emery *et al.* 2019).

2.5.1. Imaging in the diagnosis of kOA

Radiography (X-ray) is a gold standard for the morphological assessment of OA. Major OA-associated radiographic hallmarks include a progressive loss of articular cartilage and bony features, such as Oph-s, subchondral sclerosis, and subchondral cysts (Altman & Gold, 2007). Joint space narrowing (JSN) is still the only structural end point currently approved by the U.S. Food and Drug Administration (FDA) to demonstrate an efficacy of disease-modifying OA drugs (DMOAD) in phase-III clinical trials (Roemer *et al.* 2014). However, a precise determination of cartilage is not possible by radiography; joint space width (JSW) is an indirect measure, which depends on cartilage thickness and meniscal integrity, extrusion, or subluxation (Gale *et al.* 1999; Hunter *et al.* 2006). Therefore, Oph on knee radiographs are both sensitive (91%) and fairly specific (83%) (Altman *et al.* 1986; Katz *et al.* 2021).

While the knee joint is a multicompartmental, a weight-bearing posteroanterior view to visualize the TF joint and skyline view for axial projection or lateral view for the PF joint is used to confirm all the relevant OA-related radiographic changes which may not be parallel in each compartment (Fig. 2) (Sharma, 2021). However, the skyline view has a better reproducibly assessing PF compartment than the lateral view (Jones *et al.* 1993).

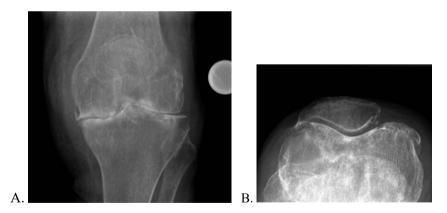


Figure 2. Knee radiographs for examination of osteoarthritis-related changes (from an unpublished personal archive of professor Aare Märtson). A. Weight-bearing posteroanterior view to visualize the tibiofemoral (TF) joint; B. Skyline view of axial projection of the patellofemoral (PF) joint. Radiographs are from a female (aged 82 years) with radiographic OA grade 4 according to Kellgren and Lawrence (KL) grading system.

Low sensitivity and specificity for the detection of OA-associated cartilage damage, a poor correlation between radiographic findings and symptoms of the disease (e.g. pain), lack of ability to detect synovitis and bone marrow lesions, variations in semi-flexed knee positioning are major limitations of radiography (Bedson & Croft, 2008; Schiphof *et al.* 2008; Roemer *et al.* 2014; Mobasheri *et al.* 2019b). Despite these limitations, radiography remains the most widely used, well accessible, and affordable imaging modality.

To standardize the evaluation of kOA severity, several semi-quantitative imaging-based scoring systems have been worked out (Roemer *et al.* 2014). The most widely used radiographic scoring system to define classification criteria for clinical trials is Kellgren and Lawrence (KL) grading system (Roemer *et al.* 2014; Hayashi *et al.* 2018). This system categorize kOA into five grades (0–4) (Grade 0 = normal; Grade 1 = presence of equivocal osteophyte; Grade 2 = presence of definite osteophyte without joint space narrowing (JSN), Grade 3 = presence of JSN; Grade 4 = complete loss of joint space). KL score 2 or more is traditionally accepted as a definitive criterion for kOA and the inclusion criterion for OA research (Kraus *et al.* 2011).

However, the KL grading has some limitations – the definition of OA is based on the TF joint, and does not estimate the status of the PF joint. Several knees with KL Grade 0 revealed OA features detectable by MRI (Guermazi *et al.* 2012). Although KL grade 1 is not commonly used in the definitive diagnosis of kOA, its inclusion in epidemiologic studies may be useful; this is because, KL 1 had a higher subsequent risk of OA progression than KL grade 0 knees (Hart & Spector, 2003). KL 3 expresses a large scale of JSN severity and is insensitive to longitudinal changes (Felson *et al.* 2011). KL Grade 4 is considered as the end-stage of kOA, but MRI can detect further progression in non-bony structures (Guermazi *et al.* 2015). Because the KL scoring system is based predominantly on the presence of Oph, an atrophic form of OA with JSN may remain underestimated (Kraus *et al.* 2011). Finally, the results of OA assessment of radiographic severity by the KL grading system are relatively variable among observers (Spector *et al.* 1993).

Therefore, complementary radiographic scoring systems for OA were developed. One of the most frequently used is, the OA Research Society International system, published as an OARSI photographic atlas of radiographs (Altman & Gold, 2007). In this scoring system, kOA can be assessed separately on JSN (graded as 0 (none), 1 (mild), 2 (moderate), and 3 (severe), and Oph-s (divided similarly into grades 0–3) both in TF and PF joint. As the severity of osteophytosis and JSN is not proportional (Jones *et al.* 2004), separate grading of the two illustrates the severity and progression of OA in a better way. Intra- and interobserver reproducibility are suitable for clinical practices (Lanyon *et al.* 1998).

However, in radiographs, the grades for JSN and Oph do not strictly increase geometrically, and unusual shapes are found. To overcome these disadvantages, a user-friendly atlas of line drawings for grading principal features of OA has been produced by Nottingham University scientists (Nagaosa *et al.* 2000). Dra-

wings were presented separately for JSN and Oph in the TF and PF compartments and separately for men and women (in the case of JSN). Grade 0 means normal joint widths, grade 1 and 2 were calculated as two-third and one-third of the widths of grade 0, respectively. Grade 3 being bone on bone and accords to the maximum sized osteophyte; grades 1 and 2 are two-thirds and one-third decrease of grade 3 Ophs. Nottingham atlas sustained the reproducibility with OARSI atlas but has more advantages in practical use.

Magnetic resonance imaging (MRI) is indicated only in special cases (e.g., in doubt of displaced meniscal tear, tumour) in routine diagnostics of kOA (Katz et al. 2021; Sharma, 2021). It is useful for research studies due to its ability to evaluate the knee as a whole organ, while multiple tissues can be visualized simultaneously over several time-points (Roemer et al. 2014). MRI can detect changes in other knee tissues such as cartilage, menisci, bone, synovium, capsular structures, and ligaments. Of note, sometimes MRI images may be affected by artifacts that mimic pathological findings (Roemer et al. 2014). The technique has its semiquantitative scoring systems, e.g. MRI OA Knee Score (MOAKS) (Hayashi et al. 2018). At present, MRI is not recommended as an aid to identify early kOA (EKOA) in routine clinical practice due to the absence of validated consensus criteria and the high frequency of MRI-detected structural joint changes in the population (Luyten et al. 2018).

Ultrasound imaging enables the detection of synovial pathology, which is a major advantage over radiography; it can visualize joint effusion, synovitis, Oph, cortical erosive changes, and other features (Katz *et al.* 2021; Roemer *et al.* 2014). It is less expensive and more portable than MRI but not as accurate as MRI in the assessment of JSN (Podlipská *et al.* 2016). Ultrasound is frequently used in imaging of hands in OA research (Hayashi *et al.* 2018), however, our research group has demonstrated valuable additional findings of this imaging technology in EKOA patients (Kumm *et al.* 2009).

Arthroscopy provides a direct visualization and palpation of intra-articular soft tissues (Chu *et al.* 2012). It enables to study radiographically invisible pathologies such as meniscal tears, articular cartilage lesions, and cruciate ligament tears. Moreover, articular cartilage softening, called also chondromalacia, detected by a surgeon through subjective palpation is the earliest detectable clinical sign of pre-OA (Outerbridge, 1961). A modified Outerbridge system for arthroscopy evaluates cartilage damage in five grades (0–4); Grade 0: smooth, firm articular cartilage, Grade 1: articular cartilage is surface-intact, but softened, Grade 2: articular cartilage with a damaged surface <50% of tissue depth, Grade 3: articular cartilage with a damaged surface >50% of tissue depth Grade 4: articular cartilage with full-thickness tissue disruption extending to the subchondral bone.

2.5.2. Patient-reported outcome measures in kOA

Patient-reported outcome measures (PROMs) are preferable methods for a regular assessment of health status and outcomes in OA to follow up the disease dynamics and even the response to treatment (O'Neill et al. 2018). A standard set of outcome measures includes joint pain, function, and quality of life scales (Rolfson et al. 2016). The most widely used disease-specific questionnaires in kOA are The Western Ontario and McMaster Universities osteoarthritis index (WOMAC) and Knee injury and Osteoarthritis Outcome Score (KOOS) (Bellamy et al. 1988; Roos et al. 1998). While WOMAC was developed focusing on the elderly people to assess OA status, KOOS was made as an extent of WOMAC for younger and more active patients with knee injuries or kOA (O'Neill et al. 2018). WOMAC consists of three domains - pain, stiffness, and physical function (activities of daily living [ADLs]) including 24 items with a recall period previous 48 hours (McAlindon et al. 2015). The scoring range in individual domains varies as -pain from 0 to 20, stiffness from 0 to 8, and physical function from 0 to 68. A total score is obtained by the summation of all the three individual scores. A higher score shows worse disease status.

On the other hand, KOOS involves five domains – pain, symptoms, ALD function, Sport and Recreation Function, and quality of life, including 42 items. The self-evaluation covers the past week. In contrast to WOMAC, the total KOOS score should not be calculated. 5-point Likert scale scores are transformed to a 0–100 scale for each domain, with 0 meaning extreme knee problems and 100 representing no knee problems. Similar to WOMAC scores, KOOS scores are influenced significantly by age, sex, and BMI in the general population (Marot *et al.* 2019).

2.6. Course of kOA

Process of OA development can be viewed as a multi-stage continuum from health to joint failure (Kraus *et al.* 2011). Staging of kOA development starts with pre-kOA, when the first molecular biomarkers appear in the absence of symptoms and signs, followed by symptomatic early-stage kOA, established kOA, and finally end-stage kOA (Mahmoudian *et al.* 2021) (Fig. 3). Spectrum of joint tissue damage levels can be divided into molecular, pre-radiographic, and radiographic stages, defined by the techniques' capabilities in distinguishing damaged from healthy tissue (Migliore & Massafra, 2014).

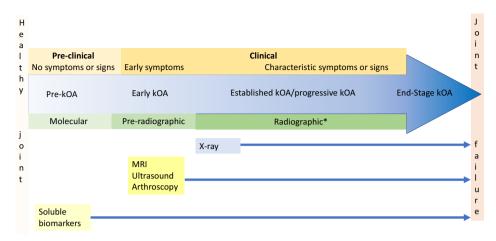


Figure 3. A course of knee osteoarthritis: staging and diagnostic capabilities according to the stage. *Radiographic kOA stage is defined as KL grade ≥2. The figure is inspired by the works of V. Kraus *et al.* (2011), A. Mahmoudian *et al.* (2021), and Z. Lv *et al.* (2021).

2.6.1. Pre-kOA

The novel, pre-kOA stage, was proposed for a characterization of the disease stage where cellular processes have been triggered but no structural changes can be detected by standard MRI, arthroscopy, or X-ray (Ryd *et al.* 2015). The risk factors-related molecules that contribute to a long transition from healthy to early symptomatic kOA, can act as biomarkers for the disease prediction and evaluation (Lv *et al.* 2021).

2.6.2. Early-stage kOA

Although a common understanding about EKOA involves the stage where the first relevant OA findings only emerge, the definite, harmonized definition is not established, and a validated diagnostic criteria are not available for EKOA at present (Mahmoudian et al. 2021). The study population in distinct EKOA groups includes either symptoms such as pain and stiffness (the CHECK study), or the presence of risk factors such as a previous knee surgery or injury and overweight (the OAI study) (Wesseling et al. 2009). The middle-aged individuals of the Estonian kOA cohort were recruited from the general list of family doctors. The identification of EKOA is thought to be problematic, because the characteristic clinical signs and symptoms of kOA may still be limited and occur sporadic (Luyten et al. 2012). The first symptoms and structural changes on imaging can appear often discordant (Roemer et al. 2015). The first symptoms could be vague knee pain on bending activities such as using stairs, twisting and pivoting, swelling of the knee, or difficulty to the knee in taking off socks/stockings (Hensor et al. 2015). Thus, early symptomatic kOA can be detected with reliable PROMs like KOOS. These prodromal symptoms can last for 2-3 years before the onset of radiographic kOA (Case et al.

2015). Patients could also experience stiffness after a period of inactivity (e.g. after awakening), which normally disappears with an exercise for a few minutes, known as 'gelling phenomenon' (Madry *et al.* 2016).

Clinical examination mostly reveals almost normal range of mobilization, joint-line tenderness, crepitus, or mild joint effusion (Mahmoudian *et al.* 2021). Radiographic evaluation is of limited value in EKOA and by the time the first definite Ophs as per the KL classification are visible by radiography, the loss of cartilage reaches more than 10% (Jones *et al.* 2004).

For clinical research studies, the reliable discrimination of the patients with symptomatic EKOA from the patients with knee symptoms due to other pathologies a set of classification criteria is proposed. These consensus-based criteria consist of three classes: (1) pain, symptoms/signs, self-reported function, and quality of life using PROMs (KOOS) (scoring ≤85% in at least 2 out of these 4 categories); (2) clinical examination – at least 1 criterion should be present out of joint line tenderness or crepitus; (3) knee radiographs – Kellgren & Lawrence (KL) grade 0 or 1 (Luyten *et al.* 2018). Biomarkers may have future utility in EKOA classification, but no individual or set of biomarkers is yet robust enough (Luyten *et al.* 2018).

2.6.3. Established kOA and progression of the disease

The next stage, established kOA, is characterized by typical kOA symptoms and/or signs, although a severity of the symptoms and structural damages as assessed on imaging are often discordant (Sharma, 2021). A majority (90%) of the subjects with chronic knee pain but without radiographic changes developed knee OA over a period of 12 years (Thorstensson *et al.* 2009).

Clinical progression was defined as an increase in pain (using scales such as WOMAC or KOOS), worsening in physical function by lower limb performance test results, or knee joint surgery as an outcome (Bastick *et al.* 2015a). Although large variation in definitions to characterize structural worsening of kOA is existing, the studies with structural progression have commonly used two criteria: decrease in joint space width (JSW) or increase in radiographic grade (generally KL grade) (Bastick *et al.* 2015b). However, the definitions using both Oph and JSN provide the most precise estimation of an association between the most known OA risk factors and kOA progression (LaValley *et al.* 2001).

Systematic reviews of kOA prognostic factors found that age, BMI, ethnicity, co-morbidity count, MRI-detected infrapatellar synovitis, joint effusion, and baseline OA severity (both radiographic and clinical) are associated with clinical kOA progression (Bastick, Runhaar, *et al.* 2015); whereas baseline knee pain, presence of Heberden nodes, varus alignment, and high levels of serum hyaluronic acid (sHA) and TNF-α were able to predict radiographic kOA progression (Bastick *et al.* 2015b).

Although OA is generally a slowly progressive disease, there are variabilities in trajectories of symptoms and structural changes, and patients with kOA (Katz *et al.* 2021). While some individuals have a trajectory of slowly worsening

symptoms and experience an intermittent pain over many years, whereas some undergo a rapid disease worsening (Katz et al. 2021; Mahmoudian et al. 2021). An overview of different trajectories of structural progression is rather complicated because there is no universal definition of kOA progression and drawing the trajectory is somewhat subjective (Collins et al. 2021). Felson at al. (2013) has proposed a concept that progression of kOA follows a state of inertia – the stable knees tend to remain stable, whereas knees with recent worsening would be expected to continue (Felson et al. 2013). However, 8-year longitudinal data in a patient with kOA structural changes, indicated that a stable trajectory is prevalent (over 85%) (Collins et al. 2021).

Moreover, at the stage of translation from EKOA to established kOA, a separate assessment of TF and PF joints is recommended. The result of our Estonian population-based cohort of middle-aged subjects presented that the radiographic course of kOA was non-consistent with intermittent periods of progression and stabilization. The follow-up study over 6 years showed that 40% of subjects had no kOA progression but 6% of subjects showed a continuous radiographic progression (Kumm *et al.* 2012). It turned out that the progression rate of radiographic kOA over 9 years was 69%, of which approximately one-third of progression was only in PF joint (Kumm *et al.* 2013a).

Accelerated kOA (AKOA) is defined as a process characterized by a transition between no radiographic kOA to advanced-stage kOA in less than 4 years (Driban *et al.* 2014). Approximately 3.4% of adults and at least 1 in 7 cases of emerging kOA were found to develop AKOA (Driban *et al.* 2014). Two out of three adults that develop AKOA will experience a sudden onset and progression within 12 months. To identify the patients who follow an accelerated track is a high priority in OA research because these patients are expected to experience the best effect of appropriate DMOAD therapy (Collins *et al.* 2021). Importantly, identifying risk factors/prognostic biomarkers for rapid progression could uncover the targets for preventive management (Conaghan *et al.* 2014).

2.6.4. End-stage kOA

End-stage kOA is an advanced stage of the disease with persistent severe pain and functional limitations, accompanied by complications, such as flexion contractures and joint laxity, restricting normal joint function (Driban *et al.* 2016). Radiographically, end-stage kOA generally corresponds to KL grade 4. Currently, as the licensed DMOADs are missing, the OA therapy focuses on pain management and improving disability and quality of life with non-pharmacologic methods such as physical activity/exercise and weight management coupled with self-management strategies (Hawker, 2019). At this stage, TKR surgery is the treatment option if non-surgical management fails to provide any relief (Hunter & Bierma-Zeinstra, 2019). However, TKR is an expensive management. The study by Losina *et al.* (2015) showed that the costs of primary TKR accounted for ~60% of OA-related total direct medical costs (Losina et al. 2015). If current eligibility criteria for TKR will continue, over

50% of symptomatic kOA patients in the US are expected to receive TKR in their lifetimes (Losina *et al.* 2015). Whereas a minor percentage of the typical kOA patient (0.3%) undergo TKR over 8 years, 1 in 7 AKOA cases (14%) are performed TKR during a span of 9-years. More seriously, 7% of AKOA patients may undergo TKR even within 2.3 years after the onset of radiographic progression (Davis *et al.* 2018). However, the expected outcome of TKR is not always achieved and 20–30% of TKR recipients have reported no benefit from TKR one year after surgery (Hawker, 2019). Therefore, a discovery of predictive biomarkers as a part of TKR appropriateness tool to identify the individuals with the best improvement after TKR is urgently needed.

Taking together, this kind of staging of OA would enable a discovery of the most appropriate biomarkers for each stage, and eventually an identification of diagnostic and management algorithms that could fit the right patient in the corresponding stage.

2.7. OA biomarkers: renewed approach

A biomarker is defined by the National Institutes of Health (NIH) Biomarkers Definitions Working Group as "a characteristic that is objectively measured and evaluated as an indicator of normal biological processes, pathogenic processes, or pharmacologic responses to a therapeutic intervention" (Biomarkers Definitions Working Group, 2001). There is a clear need for biomarkers that could select a patient for personalized and eventually a safer treatment (Karsdal *et al.* 2014; Bay-Jensen *et al.* 2016). However, it is important to understand that one biomarker may not fit for all purposes and one has to find the appropriate area of use for each biomarker (Kraus & Karsdal, 2021).

Biomarkers can be divided into two major groups- the non-soluble or 'dry' or 'in vivo' biomarkers (patient-reported outcome measures (PROMs), performed tasks, or imaging) and the soluble or 'wet' or 'in vitro' biomarkers, which are usually endogenous molecules measured in a selected body fluid such as blood (serum, plasma), urine, or synovial fluid (SF) (Kraus *et al.* 2011). Biomarker measurement in SF is a more direct study of joint tissue metabolism and structural changes than in serum or urine (Rousseau *et al.* 2021; Mobasheri *et al.* 2017; Elsaid & Chichester, 2006). However, it is not feasible in all patients because sampling involves an invasive procedure, and standardizing protocols for analysis are needed (Rousseau *et al.* 2021). On this background, urine values showed a better correlation with structural changes when compared to serum values (Elsaid & Chichester, 2006).

While OA may be long-time asymptomatic in its early stages, biomarkers that reflect a tissue turnover and preclinical disease activity provide an early warning of the onset of tissue damage, enabling earlier diagnosis of OA (Kraus *et al.* 2011; Kraus *et al.* 2015; Kraus & Karsdal, 2021). These biomarkers may allow earlier management to prevent cartilage and bone destruction that leads to disability (Kraus *et al.* 2011). Instead of commonly used PROMs, the develop-

ment of disease-modifying OA drugs (DMOADs) requires objective endpoints such as measures of joint structure (imaging biomarkers), joint tissue homeostasis (molecular biomarkers), and/or joint survival (joint replacement frequency) (Kraus & Karsdal, 2021).

Imaging biomarkers such as radiographs or MRI may provide a sensitive measure of knee joint status, whereas molecular biomarkers are often produced during the pathophysiological process (Kraus & Karsdal, 2021). Thus, molecular biomarkers, especially neo-epitopes, are sensitive and dynamic markers for joint tissues' turnover, reflecting more disease activity and thereby rate of disease progression, than the current status (Karsdal et al. 2010; Siebuhr et al. 2014a). They are potentially more useful treatment efficacy markers as compared to conventional imaging methods because of their fast reaction to the pathological changes (Karsdal et al. 2019). They have a unique property to detect changes within a few weeks (Rousseau et al. 2021). However, the early and end-stage disease may be associated with different activity periods, therefore imaging may not correlate to molecular biomarkers (Kraus & Karsdal, 2021). OA is a slowly progressive disease for which molecular biomarkers could provide a more rapid indication of an active period of the disease and therapeutic response than other biomarkers (Kraus et al. 2011). Identification of prognostic markers that predict rapid progression, is necessary for patient stratification, while the preventive management and early interventional treatment are justified to target a high-risk group (Favero et al. 2015). In summary, molecular and imaging markers complement each other and their combined utility may be necessary (Dam et al. 2009; Kraus & Karsdal, 2021).

Biomarkers require a validation as well as a qualification for their use as a surrogate endpoint (Wagner, 2002). The NIH-funded OA Biomarkers Network in 2006 proposed a classification of molecular biomarkers, named by the acronym BIPED (Bauer *et al.* 2006). This classification scheme includes five categories based on the key parameters of utility – Burden of Disease (B), Investigative (I), Prognostic (P), Efficacy of intervention (E), and Diagnostic (D). In 2011, the OARSI FDA Osteoarthritis Biomarkers Working Group added "S" into the categories, which stands for safety (BIPEDS) (Kraus *et al.* 2011). In 2016, the FDA-NIH Biomarkers Working Group published the BEST (Biomarkers, EndpointS, and other Tools) glossary which includes more detailed descriptions of biomarker functions and surrogate endpoints (FDA-NIH Biomarker Working Group, 2016). While BIPED criteria are more suitable for early biomarker development, the BEST criteria may be valuable at the phase of approval, qualification, or labelling of that biomarker for clinical usage (Mobasheri *et al.* 2017). An overview of both criteria is presented in Table 1.

Many of the existing OA-related biomarkers are associated with radiographic severity, progression, and pharmacodynamics (Bay-Jensen *et al.* 2022). However, biomarkers at the level of *in vitro* diagnostics (IVD) in all categories are lacking (Bay-Jensen *et al.* 2022). A potential use of biomarkers in personalized medicine and their role in monitoring drug efficacy are most promising (Mobasheri *et al.* 2017; Rousseau *et al.* 2021). A very few bio-

markers have been tested to identify and define specific endotypes of OA and should be a focus of future research efforts (Mobasheri *et al.* 2017; Bay-Jensen *et al.* 2022).

Table 1. BIPEDS and BEST biomarker classification criteria (Mobasheri et al, 2017).

	BIPEDS		BEST
Diagnostic Biomarker	classifies subjects as – diseased or non- diseased.	Diagnostic Biomarker	detects or confirms the presence of a disease or condition or identifies subjects with a subtype of the disease.
Prognostic Biomarker	predicts a future onset of OA among those without OA at baseline or OA progression among those with the existing disease.	Susceptibility/ Risk Biomarker Prognostic	indicates a potential of developing a disease or condition in a subject, who does not currently have a clinically apparent disease or condition. identifies a likelihood of a
		Biomarker	clinical event, disease recurrence, or progression in patients who have the disease or condition.
Burden of Disease Biomarker	assesses the severity or extent of disease, typically at a single point in time, among OA subjects.	Monitoring Biomarker	is measured serially for assessing the status of a disease or condition or as evidence of exposure to (or effect of) a medical product or an environmental agent.
Efficacy of Intervention Biomarker	provides information about the efficacy of treatment among OA patients or those at high risk of developing OA.	Pharmaco- dynamic/ Response Biomarker Predictive Biomarker	shows that a biological response occurred in a subject who has been exposed to a medical product or an environmental agent. identifies subjects who are more likely to experience a favourable effect from exposure to a medical product or an environmental agent than similar subjects without the
Safety Biomarker	monitors the health status of the joint tissue or general cytotoxic status in response to treatment.	Safety Biomarker	biomarker. measured before or after exposure to a medical product or an environmental agent, indicates the likelihood, presence, or extent of toxicity as an adverse effect.
Investigative Biomarker	insufficient information to allow inclusion into one of the existing categories.		

2.7.1. Inflammatory markers

Although synovitis may not initiate OA, it may become the driver of the disease at some later phase (Karsdal *et al.* 2014; Kraus *et al.* 2015). Majority of subjects with radiographic kOA have inflammation, including effusion in 70–81% of patients and synovial thickening in 34–50% (Tarhan & Unlu, 2003). The cytokines levels in SF, such as IL-1α, IL-18, and TNF-α, were associated with OA severity grades (Daghestani & Kraus, 2015). Moreover, a baseline level of IL-18 can predict OA progression. Recently, the association of synovial MMP-3, soluble vascular cell adhesion molecule-1 (sVCAM-1), soluble intercellular adhesion molecule-1 (sICAM-1), vascular endothelial growth factor (VEGF), tissue inhibitor of metalloproteinase-1 (TIMP-1), and monocyte chemoattractant protein-1 (MCP-1) with synovial inflammation, clinical symptoms, and radiographic severity of kOA has been shown (Haraden *et al.* 2019).

Identifying systemic biomarkers that can reflect localized joint inflammation in OA is a rather complicated task (Kraus & Karsdal, 2021). C-reactive protein (CRP) is a commonly used inflammatory marker, but it does not specify a location of the tissue (Skjøt-Arkil *et al.* 2012). Erythrocyte sedimentation rate, IL-1β, IL-6, TNF-α, and fibrinogen have the same disadvantages in addition to high variation (Siebuhr *et al.* 2016). Moreover, a meta-analysis of 32 studies on CRP revealed its significant association with OA pain and decreased physical function, but not radiographic OA (X. Jin *et al.* 2015). Interestingly, the higher CRP was associated with greater kOA pain in women, but not in men (Perruccio *et al.* 2017).

Another opportunity to identify a status of joint inflammation is to study the tissue turnover biomarkers, which are influenced by inflammation. MMP-mediated type I and III collagen degradation markers C1M and C3M, the markers of connective tissue destruction, are highly correlated with CRP (Siebuhr *et al.* 2013; Siebuhr *et al.* 2016). CRPM is a metabolite of CRP that is generated in joint tissues by MMP (Siebuhr *et al.* 2014) and serve as a promising marker of multi-joint inflammation (Alexander *et al.* 2021). High level of C1M is positively associated with KL score, CRP, and CRPM, (Siebuhr *et al.* 2014; Alexander *et al.* 2021).

A growing body of evidence indicates that a subset of OA patients with inflammation has the highest risk of progression and symptoms (Daghestani & Kraus, 2015). Although the role of inflammation in OA pathogenesis is strongly recognized, clinical trials with several anti-inflammatory agents have failed (McAlindon *et al.* 2017; Deyle *et al.* 2020). One of the promising ways could be an identification and classification of molecular inflammatory markers for better subdivision of heterogeneous groups of patients into the homogeneous subset of inflammatory phenotype for effective clinical trials testing of anti-inflammatory therapeutics (Lv *et al.* 2021).

2.7.2. Bone remodelling markers

Although alterations in subchondral bone are not evident in all patients with kOA, they can be the earliest pathological changes in a fraction of patients (Hu *et al.* 2021). OA changes in bone occur more rapidly and therefore are recognizable at initial stage as compared to cartilage abnormalities (Goldring & Goldring, 2010). So, early-stage kOA displays a typical activation of bone resorption, since an increased formation and decreased mineralization of subchondral bone occurs in late-stage kOA (Funck-Brentano & Cohen-Solal, 2011).

Type I collagen (Col1) is the major protein in bone and its fragments can be used for the characterization of bone turnover (Siebuhr *et al.* 2014b; Karsdal *et al.* 2019). N-terminal propeptide of type I procollagen (P1NP), the product of posttranslational cleavage of Col1, is a marker of bone formation (Eastell & Szulc, 2017). In the Estonian middle-aged cohort study, the authors reported that higher baseline values of serum P1NP are associated with kOA progression, especially the progression of osteophytosis (Kumm *et al.* 2013c).

C-telopeptide of Col1 (CTX-I) and N-telopeptide of Col1 (NTX-I) are degradation products of Col1 and indicate osteoclast activity (Siebuhr *et al.* 2014b). They can be assessed as markers of bone resorption in serum (s) and urine (u). uNTX-I and uCTX- are significantly increased in the patients with progressive kOA as compared to controls (Bettica *et al.* 2002). CTX-I epitope exists in two forms – non-isomerized form of CTX-Iα is an indicator of newly formed bone, while isomerized CTX-Iβ is a consequence of aging (Huebner *et al.* 2014). Data from the FNIH OA Biomarkers Consortium study showed that time-integrated concentrations over 24 months of sCTX-I, sNTX-I, uNTX-I, uCTX-Iα, and uCTX-Iβ can predict clinically relevant kOA progression (Kraus *et al.* 2017). In summary, bone remodelling markers can reflect high bone turnover suggesting the kOA progression.

2.7.3. Cartilage markers

While Col2 is the main structural component of cartilage, numerous tests of several distinct degradation fragments of Col2 have been developed for non-invasive and objective assessment of OA. In addition to aggrecan which is another main component (next to Col2), cartilage consists of minor collagens and non-collagenous proteins such as cartilage oligomeric matrix protein (COMP), fibulin, cartilage intermediate layer protein (CILP), follistatin-like protein 1 (FSTIL-1), etc. (Karsdal *et al.* 2019; Kumavat *et al.* 2021). These proteins are targeted as OA potential biomarkers in a range of studies.

2.7.3.1. Aggrecans

The structure of aggrecan is extremely complex and multiple cleavage sites create neoepitopes, for example, peptides with amino acid sequences of FFGV (AGNx-2), NITEGE (AGNx-1), and ARGS (Bay-Jensen *et al.* 2022). ARGS is the most robust in this group of biomarkers and commonly considered as an

indicator of cartilage degradation. However, it also reveals cartilage remodelling (Bay-Jensen *et al.* 2022)., ARGS levels in SF are reported to be associated with WOMAC stiffness scores in end-stage OA as well as improved KOOS pain and symptoms scores in knee trauma (Struglics *et al.* 2015; Wasilko *et al.* 2016). Moreover, low serum levels of ARGS were reported to predict fast radiographic kOA progression over 2 years (He *et al.* 2021). Chondroitin sulfate 846 (CS846), the fragment of aggrecan, is considered to be a marker of cartilage turnover, although proposed to use as a marker of aggrecan formation (Mazzuca *et al.* 2006). The potential of CS846 as an independent diagnostic marker of radiographic OA was shown in the OAI-FNIH cohort study (Liem *et al.* 2020a).

2.7.3.2. Cartilage oligomeric protein (COMP)

Meta-analyses confirmed that elevated serum COMP was indicative of kOA severity and predicted the kOA progression (Hoch *et al.* 2011; Hao *et al.* 2019). A high level of serum COMP was detected after traumatic injury (Bjerre-Bastos *et al.* 2021; Bay-Jensen *et al.* 2022). In an Estonian middle-aged cohort study, serum COMP was associated with meniscal changes and progressive osteophytosis in early-stage kOA (Kumm *et al.* 2013b). However, this level was rather associated with clinical parameters of kOA than the radiographic severity in late-stage kOA (Riegger *et al.* 2020). Another study also supported the finding that the COMP level was increased in the early stages but declined in later stages of kOA. The same study demonstrated a positive correlation of COMP with pain and no correlation with radiographic grading (Verma & Dalal, 2013).

2.7.3.3. Type II collagen (Col2)

Col2, a major structural and characteristic protein of articular cartilage is composed of three identical alpha 1 chains, arranged in a triple helix (Poole *et al.* 2003). It creates a meshwork that receives stabilization from other proteins, providing cartilage with a tensile strength (Gly-Jones *et al.* 2015). In normal cartilage, there is a strict regulation of matrix turnover; this means, there is a delicate balance between synthesis and degradation of matrix proteins. In OA, this balance is disturbed; both degradation and synthesis of Col2 are usually increased (Sandell & Aigner, 2001), followed by a loss of articular cartilage matrix. The breakdown of the Col2 meshwork is considered a crucial point in the pathogenesis of OA (Henrotin *et al.* 2007). Proteolytic enzymes, including collagenases and gelatinases, participate in this breakdown process. MMP-13 may be a key enzyme in OA (Abramson & Attur, 2009). A cleavage of the triple helix produces two fragments that are 3/4 and 1/4 the length of the mature Col2 (Aurich *et al.* 2017).

Col2 is a substantial source of epitopes (Henrotin *et al.* 2007). Several potential biomarkers of Col2 fragments that reflect a cartilage turnover are developed and intensively studied (Bay-Jensen et al. 2022). Col2 biomarkers may be divided into four main groups, according to the localization and expres-

sion of epitopes (Henrotin *et al.* 2007; Fig. 4). The first group of epitopes of Col2 propeptide protein fragments that are released during collagen synthesis represents cartilage formation markers (e.g., PIIANP, PRO-C2). The other three groups are markers of cartilage degradation. The second group includes epitopes localized at the telopeptides of Col2 (like uCTX-II). The third group consists of the denaturation epitopes, which are localized in the N-terminal triple-helical region (for example, Coll2-1, Coll2-1-NO2). The final group includes the markers formed by processes of protease cleavage, denoted as cleavage neoepitopes, and localized at the cleavage site (for example, uC2C, C1,2C, TIINE) (Henrotin *et al.* 2007; Karsdal *et al.* 2010).

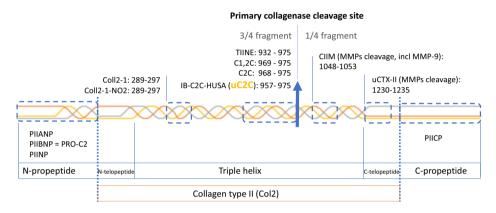


Figure 4. A schematic localization of epitopes of human type II procollagen (UniProtKB n^o P02458). The arrow indicates a primary site of cleavage leading to the neoepitopes (The design is based on the work by Rousseau *et al.* 2021).

Biomarkers of type II collagen formation

Serum levels of propeptides at the C- and N-termini (PIICP and PIINP, respectively, Fig. 4) were used for the evaluation of Col2 formation (Rousseau *et al.* 2021). These propeptides are cleaved by specific proteases during a maturation of Col2. Two main splice variants, N-propeptide of collagen IIA (PIIANP), and N-propeptide of collagen IIB (PIIBNP or termed also PRO-C2) are generated and their measurement in serum was developed (Luo et al. 2018), (Bay-Jensen *et al.* 2022). Both PIIANP and PRO-C2 levels were lower in the subjects with established kOA than in the controls (Luo *et al.* 2018). Considering a fact that high serum PIIANP predicts decreased odds of clinically relevant kOA progression PIIANP may own some prognostic value for kOA, (Kraus, Collins, *et al.* 2017). Very recently, a study revealed that low levels of baseline serum PRO-C2 were associated with a higher (3.4-fold) possibility of rkOA progression as compared to high PRO-C2 (Luo *et al.* 2021). Thus, PRO-C2 seems to be an indicator of low cartilage repair endotype in OA (Bay-Jensen *et al.* 2022).

Biomarkers of type II collagen degradation

Urinary C-terminal cross-linked telopeptide of type II collagen (uCTX-II)

uCTX-II, currently one of the most evaluated OA biomarkers, is a neoepitope generated by MMP-9 and MMP-13 (Bay-Jensen et al. 2016). Besides the damaged articular cartilage, CTX-II is also present in the tidemark and calcified cartilage at the bone interface (Huebner et al. 2014; Bay-Jensen et al. 2016). In addition to being a marker of cartilage degradation, uCTX-II is supposed to be a marker of subchondral bone degradation (Spil et al. 2013). It is a proven promising diagnostic and prognostic biomarker for OA in multiple clinical studies as summarized in the meta-analyses (Valdes et al. 2014; Huang et al. 2017). Elevated baseline uCTX-II predicts an increased risk of both symptomatic and radiographic kOA progression over 4 years, as well as a higher risk of undergoing a total knee replacement (TKR) in several OA studies (Kraus, Collins, et al. 2017; Garnero et al. 2020; Bihlet et al. 2020). Levels of CTX-II were reported to be associated with radiographic severity of OA and the number of affected skeletal sites (knees, hip, spine, and hands), and to be able to differentiate between slow and rapid OA progressors (Reijman et al. 2004; Meulenbelt et al. 2006; Dam et al. 2009). A recent meta-analysis demonstrated that uCTX-II levels were higher in the patients with severe kOA than in moderate kOA (Cheng et al. 2020). Moreover, this study revealed that uCTX-II had a better diagnostic performance in females than in males. In addition, uCTX-II has been applied in clinical trials for testing the efficacy of DMOADs such as risedronate, oral salmon calcitonin (sCT), and strontium ranelate (Rousseau et al. 2021). Despite a lack of visible treatment effect on X-ray progression, an early decline in the levels of uCTX-II was reported along with a significant response to the treatment (Bingham III et al. 2006; Karsdal et al. 2015; Alexandersen et al. 2011). However, prospective studies could show a true role of uCTX-II in evaluating the treatment effect (Rousseau et al. 2021).

Coll2-1 and Coll2-1-NO2

The assays for measuring Coll2-1 and its nitrated form, Coll2-1-NO2, were developed as another version of markers of cartilage degradation (Deberg *et al.* 2005). Coll2-1 is susceptible to nitration resulting in the production of Coll2-1-NO2, thereby reflecting an oxidative-related Col2 degradation due to local inflammation (Bay-Jensen *et al.* 2022), (Mobasheri *et al.* 2019). Serum levels of both Coll2-1 and Coll2-1-NO2 were found significantly elevated in OA patients (Deberg *et al.* 2005). Moreover, baseline levels of both markers in urine were associated with the clinical activity of kOA. Furthermore, the one-year increase in uColl2-1 levels was predictive of kOA radiological progression (Deberg *et al.* 2005). A study with the FNIH cohort reported that Coll2-1-NO2 was independently associated with radiographic OA severity (Liem *et al.* 2020b).

Cleavage neoepitopes

A cleavage at the primary cleavage site in Col2 creates a variety of neoepitope that is located on the carboxy (C)-terminus of the ³/₄ fragment and several assays

were developed to measure these neoeptiopes (Billinghurst et al. 1997) (Fig. 4). A 45-mer peptide fragment of Col2 is the most abundant necepitope peptide in urine (Nemirovskiy et al., 2007). The level of C1,2C, the utmost necepitope in the C-terminus of the 3/4 fragment, was higher in OA cartilage than in the healthy samples; however, the C1,2C assay was relatively unspecific due to cross-reactivity of Col1 fragments (Billinghurst et al. 1997). The type II collagen neoepitope (TIINE) assay targets a Col2 neoepitope upstream of the 3/4 cleavage sites, but monoclonal 9A4 antibodies in TIINE assay can recognize both type I and II C-terminal cleavage neoepitope-containing 45-mer collagen fragments. Nevertheless, uTIINE assay targets a Col2 epitope upstream of the 3/4 cleavage sites and was shown to distinguish between OA patients and healthy subjects, as well as between symptomatic and asymptomatic patients with radiographic OA (Nemirovskiy et al. 2007). C2C assay measures an elongated version of C1,2C in serum and urine, using specific monoclonal C2C antibodies. However, the old C2C assay can detect smaller fragments than the dominant 45-mer fragment and has a limited capability in detecting an initiation and progression of OA (Poole et al. 2004; Cahue et al. 2007; Cibere et al. 2009). Therefore, a new C2C assay, denoted also as C2C-HUSA, was designed to detect urinary C-terminal 45-mer fragment and any larger fragments of Col2 that contain both necepitope C2C and intrachain epitope (Poole et al. 2016). Thus, the new uC2C assay is more specific and can identify a subpopulation of fragments that are associated with cartilage degradation. Previously our study group has shown that an increased output of uC2C correlates with knee pain, a decline in the functional abilities of a lower limb, and results of lower limb performance tests (A.O. Tamm et al. 2014). A recent study the patients with knee pain showed that uC2C levels were increased in subjects with KL grade 2, a definitive for the radiographic diagnosis of OA. A baseline urinary uC2C level was significantly elevated in kOA progressors in comparison with nonprogressors and was associated with an increased risk of progression of knee cartilage degradation during the next 3 years (Poole et al. 2016). The OAI-FNIH study endorsed that uC2C was associated with kOA progression (Kraus et al. 2017). However, a role of uC2C as an OA biomarker still remains unclear because of very scarce clinical data. Quite a small number of studies have used a new, more specific uC2C method, IB-C2C-HUSA, and a complete potential of this biomarker as an indicator of distinct aspects of kOA is undiscovered and unconfirmed yet.

2.8. Summary of the literature review

There is a need to a great extent for biomarkers that could support a personalized management in kOA patients to achieve the best outcome at each stage of kOA. Currently, the development of efficient and safe OA treatment faces several important challenges such as a limited understanding of the OA joint biology, a limited biomarker toolbox for characterization of molecular processions.

ses in different joint tissues, and a lack of tools for the faster prediction of disease progression in high-risk individuals, gender differences, etc. Despite the above-described intensive research in the field of OA biomarkers, there are a few very promising candidates and none of the best candidate biomarkers has entered into clinical use as in vitro diagnostic tests (IVD). Published reviews underlined a typical shortage – a lack of consistent evidence due to differences in sample collection, implemented diagnostic criteria, investigated populations, and the used methods. Thus, there remains a clear need for more research in the field in terms of validation of existing markers and identifying new candidates applying the BIPED and BEST classifications. Moreover, such research should involve further exploration of the underlying mechanisms of OA. Since a breakdown of Col2, the major structural cartilage protein, is considered a crucial point in the pathogenesis of OA and a substantial source of epitopes, several biomarkers of Col2 fragments have been developed. The assay of uC2C detects the most abundant Col2 cleavage fragments in urine. Thus, uC2C is a potential, yet incompletely studied biomarker of kOA.

3. AIM AND HYPOTHESES

Overall aim:

To evaluate collagen type II C-terminal cleavage neoepitope C2C in urine (uC2C) as a potential biomarker for diagnosis, prognosis, and outcome of knee osteoarthritis (kOA) through its course.

Hypotheses:

- I. uC2C reflects an involvement of osteophytes (Ophs) and joint space narrowing (JSN) in the different compartments of the knee joint.
- II. uC2C is an early-stage biomarker of radiographic kOA.
- III. uC2C is higher in progressors than in non-progressors in the same stage of radiographic kOA.
- IV. Total knee replacement (TKR) reduces the level of uC2C in the 12th month after the surgery. The change in uC2C level and the Knee injury and Osteoarthritis Outcome Score (KOOS) scores after the surgery depends on its preoperative level.
- V. In presence of kOA, uC2C level can behave differently in men and women.

Specific tasks:

- I. To measure uC2C levels in healthy subjects (Paper I), early-grade (preradiographic) kOA (Paper I), advanced kOA (Paper I, III), and to specify associations of uC2C with Ophs and JSN in the tibiofemoral (TF) and the patellofemoral (PF) joint (Paper I).
- II. To compare uC2C baseline values in different groups subjects with emerging of kOA and without emerging of kOA in 3 years and controls for a prediction of onset of kOA (Paper II).
- III. To compare uC2C baseline values at distinct kOA stages in progressors and non-progressors of the disease (Paper II).
- IV. To assess preoperative uC2C and dynamics of uC2C levels after TKR during 12 months period (Paper III).
- V. To compare uC2C baseline values in the subjects with and without postoperative improvement in KOOS scores (in12 months) for a prediction of a subjective outcome of TKR (Paper III).
- VI. To compare the results of the above-mentioned tasks in different genders (Paper I–III).

4. MATERIALS AND METHODS

4.1. Study subjects

We used the subsets from three different study cohorts: Estonian Early Knee OA Study, The Arthroscopy, and Total Knee Replacement Cohort. First two cohorts consisted of a middle-aged population; this age-group could presumably be in an early stage of kOA and can provide an opportunity for a better description of soluble biomarkers to find high-risk persons for kOA and to ensure the expected DMOAD treatment outcome for the future.

- A. Estonian Early Knee OA Study Cohort. A population-based cohort was recruited by three general practitioners (GP) in southern Estonia (Kumm 2012, Kerna 2013). The relevant questionnaire was sent to randomly selected 1793 subjects and the response was received from 964 of them (54%). Out of 964 responders, 506 reported kOA complaints (knee pain in 65%, stiffness, and crepitus in 35%), and the remaining 458 had no kOA problems. Out of the 964 responders, 475 (67% females) agreed to in-depth investigations. Among them, 308 study patients indicated some subjective knee complaints, while the remaining 167 were without any complaints. Two subgroups were formed as follows:
 - a. Longitudinal group (n=388; 69% females). In addition to the subjects with knee complaints, this group consisted of knee complaints-free subjects with radiographic kOA signs or KOOS scores <85%. They were examined at least two different time points, at baseline and three years after. In 62 subjects, some data were missing and were excluded from futher analysis.
 - b. Long-term control group (n=25; 44% females). Among the study subjects without knee complaints, 35 individuals (57% females) had KOOS ≥85% and no radiographic signs of kOA at baseline. After 12 years of follow-up of these persons, 25 subjects did not develop radiographic signs of kOA and they formed a long-term control group.
- B. The Arthroscopy Cohort consisted of 109 patients (50% females), who were examined and treated during this visit at the Department of Traumatology and Orthopedics, Tartu University Hospital (Estonia) in 2007–2010. This cohort consistently recruited all subjects aged <65 years, who were indicated for arthroscopic surgery due to chronic knee complaints (duration several months to years). Men and women were recruited separately; therefore, the gender groups were similar in size. The types of knee impairments were different in the arthroscopy cohort but the clinical data were available only for 71 cases. Twenty-six subjects from 71 cases (36%) in the arthroscopy cohort had isolated degeneration of the meniscus, 23% had isolated knee trauma and 41% had a combination of degeneration and knee trauma. 84 subjects agreed

to attend the longitudinal study and were examined at the second timepoint three years later.

In both cohorts, demographic, clinical data (including KOOS questionnaire and a visual analog scale for pain (VAS Pain)), and radiographs of both knees were obtained for each subject at every time point. The subjects with radiographic evidence of rheumatoid arthritis or other inflammatory arthropathies in the knees, history of knee arthroplasty, or technically unsuitable radiographs were excluded. The knee injury was not an exclusion criterion.

C. Total Knee Replacement (TKR) Cohort. A prospective study cohort consisted of 105 patients with end-stage kOA (KL 3–4) and undergoing primary unilateral TKR between January 2017 and October 2019 at the Department of Orthopedics, Tartu University Hospital (Estonia). All the subjects under 70 years of age (mostly <65 years) targeted for knee arthroplasty were sequentially enrolled in the TKR cohort, as the progression of kOA was expected to be particularly rapid in this age group. The records of men and women were kept separately so that the groups of men and women were equal.

Paper I. A cross-sectional study involved 302 subjects from the Estonian Early Knee OA Study Cohort and The Arthroscopy Cohort, from whom the urine sample was collected and were available for assessment (Fig. 5). In a small number of subjects from the arthroscopy cohort [n = 14 (5 males and 9 females)], orthopedic surgeons performed a direct visual assessment of knee articular cartilage according to the evaluation system of *Société Francaise d'Arthroscopie* (SFA score).

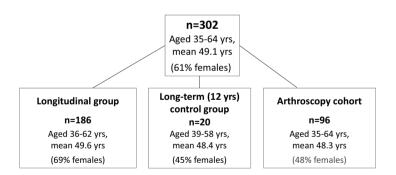


Figure 5. A distribution and characteristics of the subjects from different cohorts in Paper I. Abbreviation: yrs – years.

Paper II. In this longitudinal study, a total of 330 subjects from the Estonian Early Knee OA Study Cohort and The Arthroscopy Cohort (Fig. 6) were investigated at two time points – at baseline (T0) and a follow-up visit three years later (T3) (mean follow-up period 38 ± 5 months). Urine samples were collected and assessed at T0.

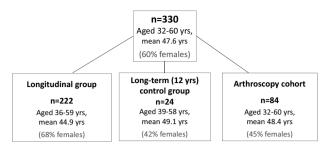


Figure 6. A distribution and characteristics of the subjects from different cohorts in Paper II. Abbreviation: yrs – years.

Paper III. A subset of the Total Knee Replacement (TKR) Cohort consisted of 86 patients. The exclusion criteria were rheumatoid arthritis or other inflammatory arthropathies, history of knee arthroplasty, or technically unsuitable radiographs. Also the patients who had signs of acute infections in the previous three months were excluded. Other exclusion criteria were an evidence of secondary OA, such as trauma, gout, infection, or congenital and developmental disorders affecting the knee joints. Relevant clinical data, including urine samples, were collected at three time points – 1–2 days before TKR and 3 months, and 12 months after TKR (Fig. 7). A clinical status of the subjects was established by PROMs such as KOOS and SF-36 questionnaires and a visual analog scale for pain (VAS Pain), along with performance tests and knee radiographs.

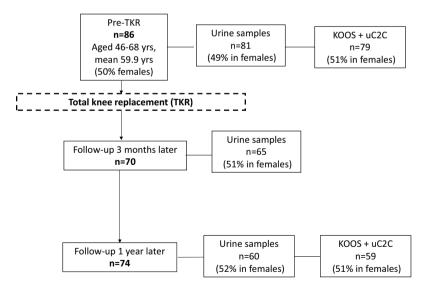


Figure 7. A flowchart that describes the total knee replacement (TKR) study groups at different time points in Paper III. Out of 86 patients at baseline, clinical and radiographic data of 70 patients was obtained 3 months after TKR, while the same data of 74 patients was collected after one year after TKR. Abbreviations: Pre-TKR – 1–2 days before total knee replacement; KOOS – Knee Injury and Osteoarthritis Outcome Score; n – numbers.

4.2. Urinary collagen type-II C-terminal cleavage neoepitope (uC2C) measurement

The second-morning void urine was collected. The collected urine samples were aliquoted and stored at -80 °C on the same day.

The level of uC2C was determined using the IBEX C2C human urine sandwich assay (IB-C2C-HUSATM) by IBEX Pharmaceuticals (IBEX Pharmaceuticals Inc., Montreal, Quebec, Canada) according to the manufacturer's instructions in duplicates (Poole et al. [2016] and https://www.ibex.ca/product-catalog/, accessed on 1 July 2020). In IB-C2C-HUSATM, two different antibodies are used to increase a specificity of the detection of Col2 fragments; capture antibodies are specific for the intrachain epitope with sequence GEPGDDGPS. and HRP-labeled tracer antibodies recognize neoepitope EGPPGPOG on Col2 fragments (Figure 8). The assay detects fragments longer than 20 amino acids including the most abundant 45-mer peptide containing the C2C necepitope. Intraassay and interassay variations of the method were $\leq 4.8\%$ and $\leq 6.7\%$, respectively. We corrected C2C concentrations with the concentration of creatinine in the same urine sample for better consideration of a urine dilution factor (A. O. Tamm et al. 2014) and the results were expressed in units such as ng/ mmol. The creatinine was measured by QuantiChromTM Creatinine Assay kit (DICT-500; BioAssay Systems, Hayward, USA) in Paper I and II, and by Cobas[®] Creatinine plus ver.2 (CREP2) kits (08057524190; Roche Diagnostics. Indianapolis, IN, USA) using a Roche Cobas c501 Analyzer in Paper III.

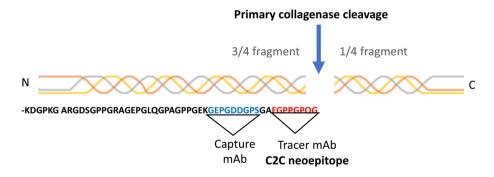


Figure 8. The epitopes of the Col2 fragments are detectable by antibodies in the IB-C2C-HUSA assayTM. Abbreviation: mAb – monoclonal antibodies.

4.3. Standardized radiographic investigation

Two grading systems — <u>the Nottingham system</u> (NSy) (Paper I-III), and the classic <u>KL system</u> (Paper III) were used for the assessment of kOA (Nagaosa *et. al* 2000; Kellgren & Lawrence, 1957). Two radiologists independently graded kOA severity by both systems. A final decision was made by a consensus between the two radiologists.

For NSy, a standardized anteroposterior radiographs of TF compartments and radiographs of PF compartments with the knees, flexed at 60° were used (Nagaosa *et. al* 2000; Kumm et. al 2012). The OA changes such as JSN and Oph in the TF and PF joints were graded on a four-point scale (grades 0–3). For bilateral cases, the knee with more severe OA served as the study knee. The highest grade of JSN or Oph was regarded as the grade of OA in the corresponding joint (TFOA or PFOA). The highest grade of OA in both compartments was defined as a radiographic global grade of the disease (gOA). A distribution of the subjects in the cross-sectional study was based on the radiographic evaluation as presented in Figure 9.

For the KL system, a five-grade classification of kOA severity (grades 0–4) was done using the same anteroposterior X-rays in Paper III. A severity of the formation of Oph, JSN, pseudocystic areas with sclerotic walls in the subchondral bone, and altered shape of bone ends was evaluated.

In Paper III, KL and NSy were used parallelly. A correspondence of a radiographic KL grade to NSy grade for the TKR cohort is presented in Table 2.

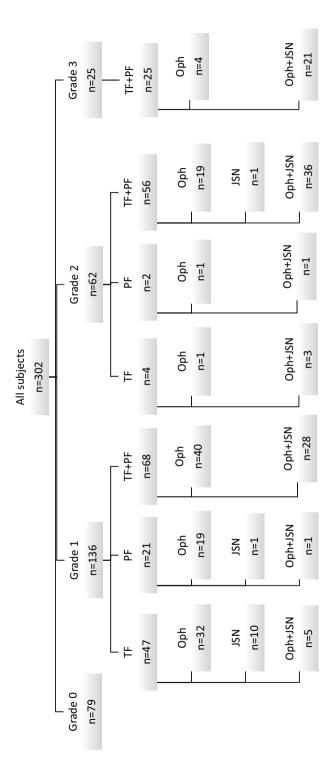


Figure 9. A distribution of the subjects of the cross-sectional study based on the grade, location, and type of radiographic changes. (Paper I). Abbreviations: Grade – radiographic global knee osteoarthritis (gOA) grade; TF – OA in a tibiofemoral joint; PF – OA in a patellofemoral joint; TF+PF - OA concurrently in TF and PF joints; JSN - joint space narrowing; Oph - osteophytes; Oph+JSN - osteophytes and joint space narrowing at the same joint; n – numbers.

Table 2. A distribution of the study cohort of Paper III by radiographic grades of kOA evaluated in 86 cases by two different systems: the Nottingham (NSy) system and the KL system.

Radiographic kOA grades/number of cases	gOA by NSy		
KL	Grade 2	Grade 3	
Grade 2	3	1	
Grade 3	11	28	
Grade 4	2	41	

Abbreviations: KL – A severity grade of knee OA assessed by Kellgren–Lawrence system; gOA – the highest grade of OA changes in the tibiofemoral and/or patellofemoral joints assessed by the Nottingham system (NSy).

4.3.1. Definitions of radiographic progression and distribution of the progression groups (Paper II)

kOA progression was evaluated by comparing radiographic changes in the subjects of the longitudinal study at time points – T0 and T3. Two main outcome groups were defined (Fig. 6): (1) the progressors' group, which was formed by the subjects with signs of radiographic kOA progression within 3 years; and (2) the non-progressors' group, which consisted of the subjects lacking the radiographic progression of kOA. The progressors' group was further divided into two subgroups according to the extent of the changes during 3 years: (a) gr(ade)-progressors, which included the subjects with radiographic worsening by at least one grade of gOA (≥1 grade); (b) the min(imal)-progressors, which contained the subjects with radiographic worsening (addition or increasing in the grade of Oph or JSN) within the same gOA grade. The subjects in gOA grade 0 at baseline but developing kOA grade 1 or more during the 3 years, were named the emerging kOA group. The subjects without radiographic findings of kOA during the whole study period (grade 0 of gOA) were defined as the without-kOA group. A long-term control group with a twelve-year follow-up period without the development of kOA formed a distinct group. A distribution of the subjects of the longitudinal study cohort (Paper II) by the progression status is demonstrated in Figure 10, and in addition, a correspondence to gOA grades in both sexes is presented in Table 3.

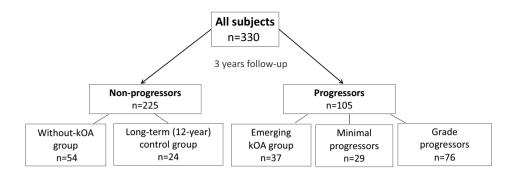


Figure 10. A distribution of the radiographic groups in Paper II. Abbreviation: kOA – knee osteoarthritis.

Table 3. A distribution of the study cohort by the progression status, gOA grades, and sex (Paper II) at T0.

	All grades	Grade 0	Grade 1	Grade 2	Grade 3
Subjects, <i>n</i> , (% all subjects)	330	91 (27.6)	169 (51.2)	55 (16.7)	15 (4.5)
Progressors, n , (%)	105	37 (41)	40 (24)	23 (42)	5 (33)
gr-progressors, n , (%)	76	37 (41)	24 (14)	15 (27)	-
min-progressors, n , (%)	29	_	16 (10)	8 (15)	5 (33)
Non-progressors, <i>n</i> , (%)	225	54 (59)	129 (76)	32 (58)	10 (67)
Males, n, (%)	131	41 (45)	61 (36)	21 (38)	8 (53)
Male progressors, <i>n</i>	37	15	12	7	3
Male gr-progressors, <i>n</i>	26	15	7	4	-
Male min-progressors, <i>n</i>	11	_	5	3	3
Male non-progressors, <i>n</i>	94	26	49	14	5
Females, n , (%)	199	50 (55)	108 (64)	34 (62)	7 (47)
Female progressors, <i>n</i>	68	22	28	16	2
Female gr-progressors, <i>n</i>	50	22	17	11	_
Female min-progressors, <i>n</i>	18	-	11	5	2
Female non-progressors, <i>n</i>	131	28	80	18	5
111 111 01	1 1 1	1 01	1	(1.0.4.) B	

Abbreviations: gOA grade – global grade of knee osteoarthritis (kOA); Progressors – the subjects with signs of radiographic kOA progression during 3 years; Non-progressors – subjects without radiographical changes of kOA during 3 years; gr-progressors – grade-progressors, subjects with kOA progression of gOA \geq 1 grade; min-progressors – minimal progressors, subjects with radiographic worsening of kOA within the same gOA grade; n – numbers.

4.4. Evaluation of articular cartilage lesions (Paper I)

Orthopedic surgeons performed a direct visual assessment and reported a status of articular cartilage of the knee (lesions' location, depth, and extent) by using a modified Outerbridge system (Ayral, 2005). The chondral area of interest was graded 0–IV (0=normal). The extent of the lesion (%) of the respective grades was evaluated on the two articular surfaces of the medial TF compartment, medial femoral condyle, and tibial plateau. The surgeons' findings (drawings on paper) were used to calculate *Société Francaise d'Arthroscopie (SFA)* scores for the medial TF compartments (Dougados *et al.* 1994; Ayral *et al.* 2005). The SFA score (continuous variable, scale 0–100) was obtained as follows: SFA score = extent (%) of grade I lesions x 0.14 + extent (%) of grade II lesions x 0.34 + extent (%) of grade III lesions x 0.65 + extent (%) of grade IV lesions x 1.00. A distribution of the subjects of the arthroscopic group with SFA score data by radiographic signs is shown in Table 4.

Table 4. The distribution of the subjects of the arthroscopic group with medial SFA score data (n=14) according to the presence of JSN and Oph.

Radiographic signs and grades by NSy	Presence of medial SFA score (n=14)	Values of medial SFA score
JSN grade 0 and Oph grade 0	4	0.1, 1.2, 1.4, 14
JSN grade 0 and Oph grade ≥2	3	9, 22, 44
JSN grade ≥2 and Oph grade ≥2	7	13, 15, 33, 38, 42, 46, 69

Abbreviations: JSN – joint space narrowing; Oph – osteophyte; NSy – radiographic grades by Nottingham system; SFA score – *Société Française d'Arthroscopie* score for cartilage visual evaluation.

4.5. Patient-relevant outcome questionnaires

4.5.1. Knee Injury and Osteoarthritis Outcome Score (KOOS)

The knee joint complaints of the recruited subjects' were evaluated using the Estonian version of the KOOS questionnaire (www.koos.nu). The five patient-relevant subscales were evaluated separately − symptoms (7 items); pain (9 items); ADL function (17 items); sport and recreation function (5 items), and quality of life (4 items). The scores express the health status of knees on 0–100% scale, wherein 0 represents extreme knee problems and 100 represents no knee problems. KOOS scores ≥85% were considered as 'healthy knees' (Roos et al. 1998).

The change in KOOS after one year of TKR was calculated by subtracting the preoperative score value from the postoperative one. To evaluate substantial clinical benefit (SCB) of KOOS, defined as a change by more than ± 20 units, we divided patients into SCB subgroups – worsening, no change, and improvement after TKR (Glassman *et al.* 2008; Ogura *et al.* 2020).

4.5.2. Visual analog scale for pain (VASpain) in joints of different skeletal areas

VASpain was self-completed by the study subjects. Fourteen joints (skeletal areas) were evaluated: right and left hip, right and left knee, right and left ankle, right and left shoulder, right and left elbow, right and left hand (wrist, metacarpal, and finger joints included), and upper (head, neck, and thorax) and lower (lumbar spine) back. The responders were asked to place a perpendicular line at a 100 mm scale on paper. The drawings were measured and transposed to a 0–10 pain score (0 = no pain; 10 = worst imaginable pain). Separate VAS scores were calculated for knee joints (mean knee VAS score) and for other joints (median summary VAS score; sumVAS). The number of noticeably affected joints was presented as sumJoint (VASpain > 5 was evaluated as noticeable involvement of a joint).

4.6. Statistical analysis

The sample size for power 80% was calculated using a sample size calculator (https://clincalc.com/stats/samplesize.aspx) for Mann–Whitney Test U (Wilco-xon Rank Sum Test) with a confidence interval set at 95%.

The other data were analyzed using R (Free Software Foundation, Boston, MA, USA; http://www.r-project.org) and GraphPad Prism 8 (GraphPad Software, San Diego, CA, USA). Descriptive variables (age, BMI, etc.) are presented as a number or mean with standard deviation (SD) and analyzed using parametric tests (e.g., chi-squared test, t-test, ANOVA). As the uC2C concentrations and KOOS subscale values were not distributed normally (checked by Shapiro–Wilk test), nonparametric tests (e.g., Kruskal–Wallis test, Mann–Whitney U test) were used for their evaluation. P-values < 0.05 were considered significant. Bonferroni correction was used for multiple comparisons.

In addition, the linear regression models (LM) were created to adjust the analysis of uC2C concentration for the confounding factors (age, sex, BMI) that can influence the associations. To determine the ability of OA features alone and in combinations to predict uC2C concentration. R-squared (R²) as a goodness-of-fit measure for linear regression models was used/applied (Paper I).

A forward method for multiple logistic regression (GLM) with the calculation of odds ratios (ORs) and 95% confidence intervals was used to assess the association between uC2C concentrations and gOA grades or OA progression or clinical improvement in kOA after TKR. GLM models were adjusted for age, sex, and BMI (Paper I-III). For the prediction of different gOA grades in Paper I, calculated OR reflected an increase in the odds of more severe gOA grade in case of 2-fold rise of uC2C. For prediction of kOA progression (Paper II), the models containing the subjects of all grades were adjusted additionally for gOA. In Paper II, the OR showed an increase in the odds of kOA progression in case of 2-fold rise of uC2C. The analysis was performed separately for the whole

group and the sexes. A discriminative ability of GLM models was assessed using the c statistic (area under the curve, AUC).

4.7. Ethics

Research Ethics Committee of the University of Tartu approved all the studies (protocol code 140/41, 22.08.2005; 156/8, 22.01.2007; 166/M-21 26.10.2009; 219/M-10 22.10.2012; and 265/T-22, 19.12.2016), which were conducted according to the precepts of the Declaration of Helsinki. All the subjects provided a written informed consent for the study participation.

5. RESULTS

5.1. Association of uC2C and kOA radiographic features (Paper I and III)

5.1.1. Prevalence of radiographic features according to clinical study groups

Two different groups were used in this set of the study: a subset from the Estonian Early Knee OA Study Cohort and The Arthroscopy Cohort (Paper I, n=302, Table 5) and TKR patients (Paper III, n=86, Table 6).

Table 5. Description of the study cohort of Paper I divided by radiographic grades of gOA.

	All grades	Grade 0	Grade 1	Grade 2	Grade 3	P-value
Subjects, n	302	79	136	62	25	<0.0001*
Mean Age, years ±SD	49.1 ±6.5	46.6 ±6.1	48.8 ±6.1	52.0 ±6.1	51.5 ±7.0	<0.0001*
BMI, kg/m ² ±SD	$\begin{array}{c} 28.4 \\ \pm \ 5.2 \end{array}$	$\begin{array}{c} 26.6 \\ \pm 4.9 \end{array}$	27.8 ±4.5	31.9 ±5.3	29.1 ±5.6	<0.0001*
Unilateral/bilateral gOA cases, <i>n</i>	51/172	-	48/88	3/59	-/25	
Females, %	61	58	60	66	60	0.8**

Abbreviations: gOA - global grade of kOA; n - numbers; SD - standard deviation; BMI - body mass index; * difference between grades by ANOVA; ** difference between grades by Chi-squared test.

Neither mean age $(48.4 \pm 5.8 \text{ years})$ nor mean BMI $(25.9 \pm 4.6 \text{ kg/m}^2)$ of the subjects in the long-term control group showed any significant difference from the subjects with gOA grade 0. Age and BMI were similar in females and males in the same gOA groups.

Isolated Oph was the most common radiographic finding (n=116) in the whole study group (Fig. 7). Among the cases with Ophs, JSN was more frequently seen in females than in males (52% vs. 34%; chi-squared test, p = 0.01). Mean age and BMI did not differ significantly between the sexes, although BMI was slightly higher in females (mean BMI: 33.1 vs. 31.5 kg/m2 for males; p = 0.07, t-test).

Table 6. Clinical characteristics of the total knee replacement (TKR) cohort by time-points (Paper III).

Clinical Characteristics/ Timepoints	Pre-TKR	3 Months Post-TKR	12 Months Post-TKR
Subjects, <i>n</i> (%)	86 (100)	70 (81)	74 (86)
Mean age, years \pm SD	59.9 ± 4.7	60.2 ± 4.7	60.9 ± 4.7
Mean BMI, $kg/m^2 \pm SD$	32.3 ± 4.2	32.3 ± 4.2	32.4 ± 4.2
Previous TKR of opposite knee, <i>n</i> (%)	20 (23)	16 (23)	15 (20)
Females, %	50	53	49
Obesity (BMI \geq 30), n (%)	59 (69)	49 (70)	50 (68)

Abbreviations: *n* – numbers; SD – standard deviation; BMI – body mass index; TKR – total knee replacement.

5.1.2. Association of uC2C level with gOA

Median uC2C level was the lowest in gOA grade 0 group and gradually elevated until grade 3 as assessed by NSy (Fig. 11A, Paper I). Similarly, a gradual increase in uC2C level was associated with the radiographic severity of kOA, when assessed by the KL system (Fig. 11B, Paper III).

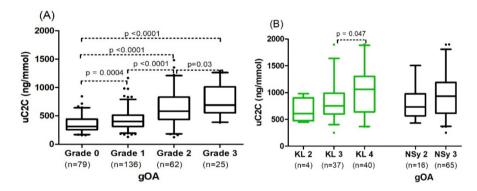


Figure 11. An association of uC2C with radiological grades of knee osteoarthritis (kOA): (A) Paper I. uC2C and gOA (n=302), (B) Paper III. uC2C and gOA by NSy or severity by KL system in the preoperative period (n=86). Box-whiskers plot with 5th-95th percentiles, the p-value determined by Mann–Whitney U-test. Abbreviations: KL – grades of radiographic knee osteoarthritis by the Kellgren–Lawrence scoring system; gOA – the highest grade of knee OA changes in two knee joint compartments (tibiofemoral and/or patellofemoral joints) by the Nottingham system (NSy), n – numbers.

As age, BMI and sex are confounders for uC2C analysis, we used GLM for further analysis. In GLM models adjusted for age, sex and BMI, the elevation of uC2C level was associated with a gradual increase in the risk of a more severe grade of kOA (adjusted OR per a grade change = 2.14–3.7; Fig. 12A, Paper I).

Similar to the unadjusted data, a significant increase in uC2C level was predicted gOA grade 1 (grade 1 vs. 0: adjusted OR = 2.14, AUC = 0.66, p = 0.01). However, the increase in uC2C level was better predicted the next gOA levels (grade 2 vs. 1: adjusted OR = 2.84, AUC = 0.79, p = 0.0007; grade 3 vs. 2: adjusted OR = 3.7, AUC = 0.74, p = 0.008). BMI had a significant prediction value only for grade 2 vs. 1 and grade 3 vs. 2, but not for grade 1 vs. 0. Overall, uC2C levels excellently predicted a presence of radiographic gOA (grade 3 vs. 0, AUC = 0.934; grade 2 vs. 0, AUC = 0.861, Fig. 12B).

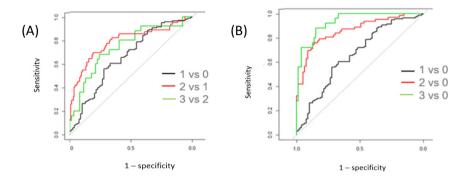


Figure 12. Receiver operating characteristic (ROC) curves of uC2C for predicting a presence of more severe gOA grade (Paper I): (A) Comparisons between consecutive gOA grades (0–3); (B) Comparisons between higher (1–3) grades of gOA and gOA grade 0. Abbreviations: gOA – the highest grade of knee OA changes in two knee joint compartments by the Nottingham system (NSy); AUC – area under the curve.

The uC2C level did not differ between the long-term (12 y) control group (gOA grade 0, without symptoms) and the subjects with knee symptoms but with radiographically normal knees (gOA grade 0, Paper I).

We detected no difference in uC2C level between unilateral and bilateral kOA cases (Paper I). There was no association between uC2C and the average pain scores of other joints (sumVAS pain) as well as with the status of previous TKR of the opposite knee (Paper III). Nevertheless, a tendency of higher uC2C levels in bilateral kOA as compared to unilateral cases could be demonstrated in females (Fig. 13, Paper III).

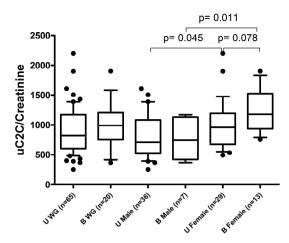


Figure 11. An association of preoperative uC2C levels with unilateral or bilateral kOA status. Boxplot with 10th–90th percentiles, Wilcoxon test for multiple comparisons was used to determine *p*–values. Abbreviations: U WG – unilateral kOA without previous TKR in the whole group; B WG – bilateral kOA with previous TKR in the whole group); U Male – unilateral kOA without previous TKR in males; B Male – bilateral kOA with previous TKR in males; U Female – unilateral kOA without previous TKR in females; B Female – bilateral kOA with previous TKR in females.

5.1.3. Association of uC2C level with TFOA and PFOA

The uC2C level was significantly higher in the combined TFOA and PFOA cases and isolated TFOA cases as compared to the cases without radiographic change (Fig. 14, Paper I). Moreover, we demonstrated a significantly higher uC2C in combined TFOA+PFOA cases as compared to isolated TFOA or PFOA.

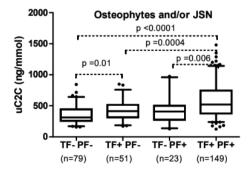


Figure 14. An association of uC2C with radiographic features of OA (osteophytes and/or JSN) in TF and/or PF joints. Box-whiskers plot with 5th-95th percentiles, the *p*-value was determined by Mann–Whitney U test. Abbreviations: TF+ – OA in a tibiofemoral joint; PF+ – OA in a patellofemoral joint; TF- no OA in a tibiofemoral joint; PF- – no OA concurrently in tibiofemoral and patellofemoral joints; JSN – joint space narrowing; n – numbers.

In linear regression analysis (Paper I), we demonstrated that gOA, TFOA, and PFOA equally predicted uC2C level (adjusted $R^2 = 0.33-0.36$). Also, a combined use of TFOA and PFOA in the same model did not increase the predictive power. Higher age and BMI, and male gender predicted higher uC2C levels.

5.1.4. Association of uC2C level with Ophs, JSN, and the SFA score

A presence of Oph in the TF joint or both, TF and PF joints, was associated with an increase in the uC2C level as compared to the cases without radiographic features of kOA (Fig. 15A, Paper I). The level of uC2C was higher in the patients with osteophytosis in both joint compartments. However, a presence of JSN did not increase the uC2C level within the same Oph grade (Fig. 15B, Paper I).

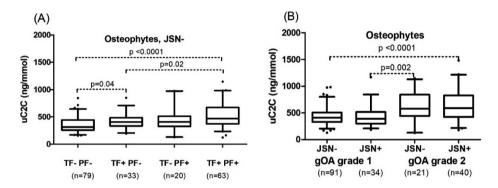


Figure 15. An association of uC2C with different combinations of radiographic features of knee OA: (A) osteophytes in TF and/or PF joints among subjects without joint space narrowing (JSN-); (B) osteophytes in gOA grade 1 and 2 in the cases with (JSN+) and without (JSN-) joint space narrowing.Box-whiskers plot with 5^{th} -95th percentiles, the *p*-value was determined by Mann–Whitney U-test. Abbreviations: TF+ – OA in a tibiofemoral joint; PF+ – OA in a patellofemoral joint, TF- no OA in a tibiofemoral joint; PF- no OA in patellofemoral joints; gOA – the highest grade of knee OA changes in two knee joint compartments by the Nottingham system; n – numbers.

Using a linear regression analysis, we demonstrated that Oph predict the uC2C concentrations better than JSN (Table 7); Oph was the most important predictor for uC2C in TF joint.

Table 7. Lm-models adjusted for age, sex, and body mass index, predicting an increase of uC2C by radiographic features and their combinations (Paper I).

Model (main parameter)	Adjusted R ²
TFOph	0.35
PFOph	0.33
TFOph+PFOph	0.35
TFJSN	0.27
PFJSN	0.27
TFJSN+PFJSN	0.29
TFOph+TFJSN+PFOph+PFJSN	0.36

Abbreviations: TF+ – OA in a tibiofemoral join; TFJSN, PFJSN – joint space narrowing (JSN) score in tibiofemoral (TF) or patellofemoral (PF) joint; TFOph, PFOph – osteophytes score in TF or PF joint.

We demonstrated that medial SFA scores ranged from 0.1 - 69 (median value of 18.6) in the study group (Paper I). Also, we found that macroscopic cartilage lesions were presented in four cases without radiographic change (Fig. 16). Generally, higher SFA scores are associated with the severity of gOA grade (p = 0.03, Kruskal-Wallis test). We demonstrated that the SFA score well predicted the uC2C level in regression models (adjusted $R^2 = 0.59$). The best prediction models for uC2C level (adjusted $R^2 = 0.75$) formed by the addition values of different OA signs: macroscopic cartilage lesion expressed by medial SFA, Ophs in the TF joint, and JSN in the PF joint. Both, the presence of Ophs and cartilage damage in TF were significantly associated with each other on statistical scale (p <0.05) and independently with the uC2C level in the model.

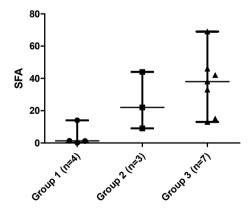


Figure 16. An association of *Société Francaise d'Arthroscopie* (SFA) score and radiographic features of knee osteoarthritis; Group 1: joint space narrowing (JSN) grade 0 and osteophytes (Oph) grade 0; Group 2: JSN grade 0 and Oph grade ≥2; Group 3: JSN grade ≥2 and Oph grade ≥2. Median and range are shown.

5.1.5. Sex-related differences in associations with uC2C and kOA features

The biomarker level did not differ between males and females in the same gOA grades 0–2 (Paper I). However, the uC2C level was significantly higher in females than in males during the preoperative period of advanced kOA (grade 3; p = 0.0039, Mann–Whitney U-test, Paper III). Moreover, a gradual increase in uC2C level was associated with the radiological grades in females (Fig. 17, Paper III; Fig. 20A, Paper II). In males, the uC2C level association was found only with the progressors of more severe radiological grades (Fig. 17, Paper III; Fig. 20B, Paper II).

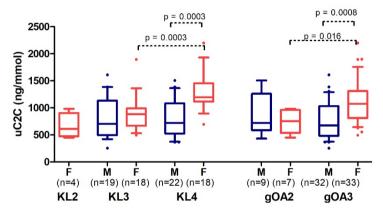


Figure 17. An association of uC2C levels with the radiological severity of knee osteoarthritis in the preoperative period in males (M) and females (F) (boxplot with 10th–90th percentiles, Mann–Whitney U-test). Abbreviations: KL – grades of radiographic knee osteoarthritis by the Kellgren–Lawrence scoring system; gOA – the highest grade of knee OA changes in two knee joint compartments (tibiofemoral and/or patellofemoral joints) by the Nottingham system (NSy); n – numbers.

Prominent sex-related differences were also demonstrated in the results of linear regression analysis combining different radiographic and clinical features. The prediction of uC2C levels by radiographic features, particularly OA features in TF joint, was better in females than in males (adjusted $R^2 = 0.35$ –0.42 vs. 0.09–0.27, Paper I). Moreover, the best uC2C prediction model worked better in females as compared to males. In females, it was comprised of a combination of Oph in the TF joint and JSN in both joint compartments (adjusted $R^2 = 0.43$). On the other hand, the prediction model in males was the combination of JSN in the TF joint and Ophs in both compartments (adjusted $R^2 = 0.28$).

5.2. uC2C level as a marker of progression of kOA (Paper II)

5.2.1. Clinical characteristics of the longitudinal study group at baseline

The subjects of the longitudinal study group were middle-aged and overweight (Table 8).

Table 8. Clinical characteristics of the subjects of the longitudinal study group at baseline in Paper II.

Groups	All gOA grades	gOA grade 0	Long-term control group
Subjects, n	330	91	24
Females, n (%)	199 (60)	50 (55)	10 (42)
	Mean age	e, years \pm SD	
Subjects	47.6 ± 6.5	46.1±6.5	49.1 ± 5.7
Males	46.6 ± 6.9	46.3 ± 6.4	48.9 ± 6.3
Females	48.2 ± 6.1	45.9 ± 6.6	49.4 ± 5.1
<i>p</i> -value*	0.04	0.8	0.8
	Body mass index	(BMI) , $kg/m^2 \pm SI$)
Subjects	28.0 ± 5.3	26.2±5.1	26.0 ± 4.6
Males	28.1 ± 4.7	27.5 ± 5.6	27.7 ± 5.0
Females	27.9 ± 5.6	25.2±4.4	23.6 ± 2.4
<i>p</i> -value*	0.7	0.03	0.01

Abbreviations: gOA - global grade of knee osteoarthritis; n - numbers; SD - standard deviation; BMI - body mass index; *difference between sexes by t-test.

The BMI of investigated subgroups (progressors/non-progressors) did not differ in males. In contrast, females presented higher BMI in progressors compared with non-progressors ($29.5 \pm 6.1 \text{ kg/m}^2 \text{ vs. } 27.1 \pm 5.2 \text{kg/m}^2$; p = 0.008, t-test). The characteristics of the control group and gOA grade 0 had no significant difference. The VAS pain score of other joints (knees excluded) did not differ between progressors and non-progressors (p = 0.3, t-test).

5.2.2. Association of uC2C at baseline level with the three-year follow-up of kOA progression

The level of uC2C was significantly higher in the progressors as compared to the non-progressors at baseline (T0) (16% difference in the median; p = 0.0008; Mann–Whitney U-test, Fig. 18). This association was found in both progression subgroups: gr-progressors (19% higher as compared to non-progressors; p = 0.003) and min-progressors (12% higher as compared to non-progressors;

p = 0.04). However, no significant difference in the uC2C levels was detected between the min-progressors and the gr-progressors.

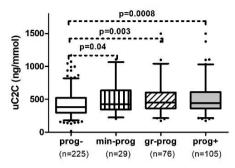


Figure 18. An association of uC2C with radiographic progression of knee osteoarthritis (kOA) in the whole study group. Box-whiskers plot with 5th–95th percentiles, the p-value determined by Mann–Whitney U test. Abbreviations: (prog-) − non-progressors, subjects without kOA radiographical changes during 3 years; (prog+) − progressors, subjects with the progression of kOA; (min-prog) − minimal progressors, subjects with radiographic worsening of kOA within the same gOA grade; (gr-prog) − grade progressors, the subjects with the progression of gOA ≥1 grade; n − numbers.

Using GLM models, we demonstrated that uC2C level successfully predicted kOA progression within three years in all the progressors (adjusted OR = 2.34 (1.48-3.68) for uC2C, p=0.0003, AUC = 0.67), and even better in gr-progressors subgroup (adjusted OR = 2.8 (1.66-4.72) for uC2C, p=0.0001, AUC = 0.73). Two confounding factors in the model contributed to the kOA progression risk – higher BMI slightly magnified a risk of kOA progression (adjusted OR = 1.07 (1.02-1.12); p=0.006), but higher gOA grade showed a mild protective effect (adjusted OR = 0.67 (0.47-0.96); p=0.03).

5.2.3. A prognostic value of uC2C for kOA progressors in distinct gOA

We presented that the level of uC2C of the progressors at gOA ≥ 1 was higher than the levels of the non-progressors at T0 (Fig. 19). We also found that the gr-progressors had a 25–33% higher median uC2C level as compared to the non-progressors (33% difference in grade 1; p = 0.001 and 25% difference in grade 2/3, p = 0.03; Mann–Whitney U test). However, no difference in the uC2C levels was found between the emerging kOA group and the non-progressors of kOA in grade 0. Interestingly, the uC2C levels of the emerging kOA subjects were rather similar to the non-progressors in gOA grade 1. Also, there was no significant difference between the uC2C values of the progressors in grade 1 and the non-progressors in grade 2/3 (p = 0.72, Mann–Whitney U test).

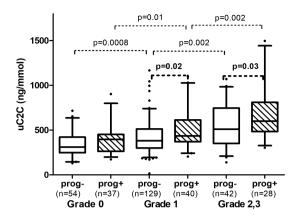


Figure 19. An association of uC2C with radiographic progression of knee osteoarthritis (kOA in all the subjects by global grade of kOA (gOA). Box-whiskers plot with 5th–95th percentiles, the p-value determined by Mann-Whitney U test. Abbreviations: (prog-) – non-progressors, the subjects without radiographical changes of kOA during 3 years; (prog+) – progressors, the subjects with kOA progression; n – numbers.

However, we demonstrated that the uC2C levels quite efficiently predicted the onset of kOA in GLM models given that the emerging kOA group was compared to the without-kOA-group, and the model was adjusted for age, gender, and BMI (adjusted OR = 2.58 (1.08-6.16) for uC2C, p = 0.03; AUC = 0.64). The best kOA prediction model was achieved when the emerging kOA was compared with the long-term control group (adjusted OR = 5.87 (1.71-20.22) for uC2C; p = 0.005; AUC = 0.79). In the same model, females demonstrated a significant risk for the developing of kOA (adjusted OR = 4.01 (1.12-14.39)) for uC2C; p = 0.03), while older age revealed some protective effect against emerging of kOA (adjusted OR = 0.86 (0.77-0.96)) per year of age; p = 0.009). Although a significant prediction power of the uC2C for the progression of the disease was demonstrated at baseline grade 1 (adjusted OR = 2.36 (1.19-4.67); p = 0.01; AUC = 0.67), we could not prove the same in the case of established kOA (gOA grade 2/3).

5.2.4. Sex-related differences between associations of uC2C and kOA progression

Albeit the uC2C level predicted the kOA progression in both sexes, several differences were observed.

In females, the levels of uC2C were 24–30% higher in the progressors as compared to the non-progressors (p = 0.0009 for the progressors and p = 0.001 for the gr-progressors, Mann–Whitney U test). Moreover, the uC2C was approximately 72% higher in the females with emerging kOA as compared to the long-term control group (p = 0.009, Mann–Whitney U test). The level of uC2C demonstrated a significantly higher efficacy to predict emerging kOA in the

GLM model as compared to the long-term control group in females when adjusted the confounders like age and BMI (OR = 23.0 (2.2–245) for uC2C; AUC = 0.91). In the advanced grades of the disease, a difference in the uC2C level between the progressors and the non-progressors was found decreased; this means, it was significant in gOA grade 1 and was found disappeared in gOA grade 2/3 (Fig.20A). This finding was also supported by GLM model analysis – the uC2C level had a significant prediction power of detecting the kOA progression in gOA grade 1 (OR = 2.67 (1.18–6.04) of uC2C; AUC = 0.68), but this ability was lost in gOA grade 2/3.

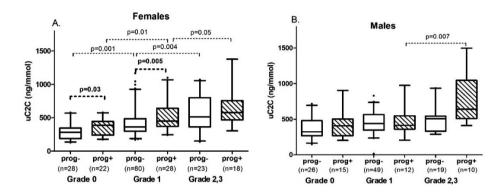


Figure 20. An association of uC2C with radiographic progression of knee osteoarthritis (kOA) in females (A) and males (B). Box-whiskers plot with 5th–95th percentiles, the p-value determined by Mann–Whitney U test. Abbreviations: (prog-) – non-progressors, the subjects without radiographical changes of kOA during 3 years; (prog+) – progressors, the subjects with the progression of kOA.

In contrast, uC2C levels in progressor and non-progressor males were not significantly different (Fig. 18B). Exception was the min-progressors, a sub-group of males with gOA grade 2/3, which demonstrated a significantly higher uC2C in comparison with the non-progressors of the same baseline gOA (833.0 (606.0–1019.5) vs. 507.0 (345.0–541.5) ng/mmol, p=0.02, Mann–Whitney U-test). We did not find any significant predictive value of uC2C in males by GLM analysis. Interestingly, mean values of uC2C in the non-progressor males were significantly higher in comparison with the values of non-progressor females (p=0.03, Mann–Whitney U-test); however, the uC2C levels did not differ in the progressors of both sexes.

5.3. uC2C and total knee replacement (Paper III)

5.3.1. The dynamics of uC2C levels after TKR

We demonstrated that although median level of uC2C was significantly elevated after three months of the TKR as compared to the pre-TKR level (p = 0.012, Mann–Whitney U test), it was found declined to the preoperative level after 12 months (Fig. 21).

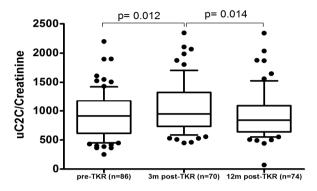


Figure 21. Dynamics of uC2C during the study period. Boxplot with 10th–90th percentiles, Mann–Whitney U test. Abbreviations: TKR - total knee replacement; pre-TKR - 1-2 days before TKR; 3m post-TKR - 3 months after TKR; 12m post-TKR - 12 months after TKR; n – numbers.

At the same time, a variety of individual changes existed – an excretion of uC2C could decrease, increase, or remain constant. The individual dynamics of uC2C over 12 months ranged from a decline of 87% to a rise of 120% in the uC2C levels (Fig. 22). Furthermore, a remarkable (~20%) increase in the postoperative uC2C levels was obvious in the cases with lower preoperative uC2C levels (16 out of the first 30 ranked cases). Conversely, several cases (12 out of the last 29 ranked cases) with a significant decrease in the post-TKR period were among those with higher preoperative uC2C levels.

Changes in uC2C values after 1 year follow-up

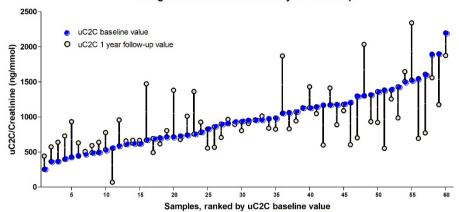


Figure 22. Individual dynamics of uC2C values (pre-TKR and 12 months after TKR) are ranked by preoperative (baseline) uC2C level. Abbreviation: TKR – total knee replacement.

Accounting a 20% change in the biomarker as a basis of grouping, we demonstrated that 28% of patients (17 out of 60) were "descenders", 32% were "ascenders" (19 out of 60), and the remaining 40% belonged to a "stable" group. A statistically significant difference between the uC2C values of "ascenders" and "descenders" was demonstrated in both pre-TKR and 12 months after TKR (p = 0.00022 and p = 0.021, respectively, Mann–Whitney U test; Fig. 21). However, the subgroups did not differ in the uC2C levels at the three-month postoperative timepoint.

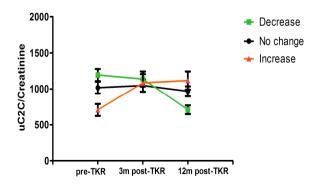


Figure 23. A line plot of uC2C dynamics in the whole group based on postoperative uC2C changes (decrease or increase means, > 20% decrease (green) or increase (red) in the 12-month post-TKR uC2C value as compared to the preoperative one; mean and standard error of the mean (SEM) shown). Abbreviations: pre-TKR – before total knee replacement; 3m post-TKR – 3 months after TKR; 12m post –TKR 12 months after TKR.

5.3.2. Sex-related differences in the dynamics of uC2C levels after TKR

In males, median uC2C was significantly increased after three months of TKR as compared to the preoperative value, and a decline, comparable to the preoperative level, was observed after 12 months, resembling the dynamic of the whole group. On the contrary, in females, the uC2C was lower for a year after surgery as compared to preoperative levels (Fig. 24).

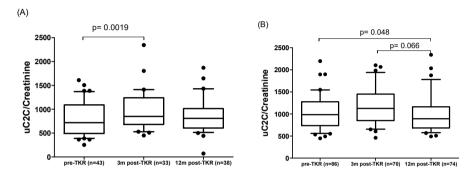


Figure 24. Dynamics of uC2C during the study period in (A) males, and (B) females (boxplot with 10th–90th percentiles, Mann–Whitney U-test). Abbreviations: pre-TKR – 1–2 days before total knee replacement; 3m post-TKR – 3 months after TKR; 12m post-TKR 12 months after TKR.

We found a statistically significant difference between "ascenders" and "descenders" males (p = 0.019 for pre-TKR and p = 0.035 for one-year post-TKR, Mann–Whitney U test; Fig. 25A), but not in females (p = 0.10 for pre-TKR and p = 0.053 for one-year post-TKR, Mann–Whitney U test; Fig. 25B).

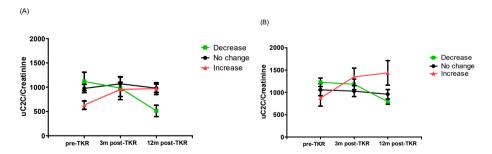


Figure 25. A line plot of uC2C dynamics in (A) males, and (B) females, grouped on the basis of postoperative uC2C changes (decrease or increase means, > 20% decrease (green) or increase (red) in the 12-month post-TKR uC2C/Crea value as compared to its preoperative value; mean and standard error of the mean (SEM) shown). Abbreviations: pre-TKR – before total knee replacement; 3m post-TKR – 3 months after TKR; 12m post-TKR 12 months after TKR.

5.4. uC2C level and patient's self-assessment outcomes (Paper III)

5.4.1. Associations between uC2C and pain

uC2C levels were weakly correlated with VAS pain of the knee joint in the whole group (Spearman's rho = 0.22, p = 0.045); however, there was not any significant association of uC2C with the sexes separately. A weak correlation between uC2C levels and KOOS pain was also observed in the whole group (Spearman's rho = -0.31, p = 0.006, for KOOS pain) and in the males (Spearman's rho = -0.33, p = 0.04), but it was missing in the females (Spearman's rho = -0.20, p = 0.21).

5.4.2. An association of uC2C levels with the improvement of KOOS subscales in the post-TKR

Significant improvements in all KOOS subscales were observed after one year of TKR in comparison with their pre-TKR scores (p < 0.0001, Mann–Whitney U test; Fig. 26). However, none of the scores of any subscale rose to the score of 85 (=healthy).

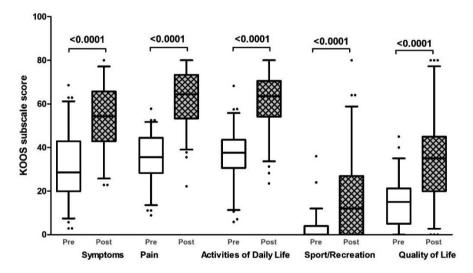


Figure 26. KOOS profiles before and after 12 months of TKR. Boxplot with 10th–90th percentiles. KOOS subscales: pain; symptoms; activities of daily living; sport and recreation; quality of life (Mann–Whitney U test *p*-values). Abbreviations: TKR – total knee replacement; KOOS – Knee Injury and Osteoarthritis Outcome Score; Pre – preoperative status; Post – postoperative (1 year after TKR) status.

Difficulties were more prominent in the activities such as squatting, kneeling, running, jumping, and twisting, as expressed by the function in the sport and recreation subscale.

A significant clinical worsening of KOOS was very rare (only one patient in this TKR study group). Approximately two-thirds of the patients reported significant clinical improvements in the pain and the function in daily living subscales (70% vs. 30% and 65% vs. 35%, respectively; Table 9). However, improvement in the KOOSsp/recr subscale was minor as compared to the nochange subgroup (36% vs. 64%).

Table 9. Substantial clinical benefit (SCB) in Knee Injury and Osteoarthritis Outcome Score (KOOS) subscales at 12-month follow-up after total knee replacement.

Clinically Important KOOS Change *	Substantial Worsening, n (%)	No Change, n (%)	Substantial Improvement, n (%)	Total **
KOOSsymp	1 (1%)	30 (42%)	40 (56%)	71
KOOSpain	0 (0%)	21 (30%)	49 (70%)	70
KOOSadl	0 (0%)	25 (35%)	46 (65%)	71
KOOSsp/recr	0 (0%)	46 (65%)	25 (35%)	71
KOOSqol	1 (1%)	32 (46%)	37 (56%)	70

Abbreviation: n – numbers; * Changes of more than ± 20 units were counted as SCB; **: number change due to missing data. Knee Injury and Osteoarthritis Outcome Score (KOOS) subscales: symptoms (KOOSsymp), pain (KOOSpain), activities of daily living (KOOSadl), sport and recreation (KOOSsp/recr), and knee-related quality of life (KOOSqol).

The preoperative uC2C level was significantly higher in the improvement subgroup of KOOSsymp as compared to the no-change subgroup of the same subscale (Table 9; p = 0.01, Mann–Whitney U test). There were no significant differences in uC2C concentrations in the other KOOS subscales.

Table 10. A comparison of preoperative uC2C between SCB subgroups of KOOS subscales.

KOOS Subscale	Sul	ostantial Improvement Group *	No Change Group		<i>p</i> -value (between
Subscale	n	uC2C ***, ng/mmol	n	uC2C ***, ng/mmol	Groups) **
KOOSsymp	38	1016.9(782.8–1204.9)	29	718.8(570.5–899.7)	0.01
KOOSpain	46	935.8(637.7–1178.9)	21	752.7(703.1–1315.5)	0.94
KOOSadl	44	974.0(626.3–1216.2)	24	731.9(668.7–978.0)	0.16
KOOSsp/recr	25	917.3(608.8–1203.5)	44	923.6(687.9–1178.7)	0.70
KOOSqol	36	956.3(626.3–1216.2)	30	839.6(685.4–1132.0)	0.55

Abbreviation: n – numbers; * Knee Injury and Osteoarthritis Outcome Score (KOOS) subscales changes of more than 20 units between the 12-month postoperative and preoperative periods were counted as SCB (substantial clinical benefit); ** Mann—Whitney U test; *** Median value (1st-3rd quantiles) of uC2C. KOOS subscales: symptoms (KOOSsymp), pain (KOOSpain), activities of daily living (KOOSadl), sport and recreation (KOOSsp/recr), and knee-related quality of life (KOOSqol).

In GLM models, the preoperative uC2C levels predicted a significant clinical improvement in KOOSsymp in the whole group (OR = 2.79; CI 95% 1.19–6.53, p = 0.02), albeit an adjustment of the confounders (age, BMI, and sex) removed the statistical significance of the model. However, in females, the power of uC2C to predict the improvement of KOOSsympt remained significant even after adjustment of cofounders (adjusted OR = 9.43; CI 95% 1.19–74.73, p = 0.03). The prediction value of uC2C was missing in males. Moreover, no associations were found between uC2C and other KOOS subscales in the whole group or separately in both sexes.

6. DISCUSSION

Despite the recent advantages in the field of OA research and joint biology, still many challenges persist in OA-related studies. This has slowed the development of OA treatment, and as a result, it has remained considerably behind the advances in other rheumatic diseases. Several aspects can be underlined in addressing the challenges – a limited biomarker toolbox and lack of appropriate surrogate endpoints in OA clinical trials, a lack of tools to identify a faster OA progression, and a poor correlation between structural progression and pain progression. New biomarkers may help to solve several among the listed problems and many aspects related to it are needed to be considered, including a proposed clinical value of new tests. Therefore, BIPEDS classification (Burden of disease, Investigational, Prognostic, Efficacy of intervention, Diagnostics, and Safety) of biomarkers (Bauer et al. 2006) or the BEST criteria (Diagnostic, Susceptibility/Risk, Prognostic, Monitoring, Pharmacodynamic/Response, Predictive and Safety Biomarkers) (FDA-NIH Biomarker Working Group, 2016) is worse to follow. There are a number of promising candidates, such as COMP, aggregan, and uCTX-II, although none is sufficiently discriminatory to distinguish the patients with different stages of OA (including emerging), to predict the individuals at a high risk of progression of the disease or to operate consistently so that it could act as a surrogate in clinical trials.

In the current thesis, we focused on exploring a potential role of uC2C, a new Col2 degradation biomarker.

6.1. uC2C: a risk and an early diagnostic biomarker of radiographic kOA

Early radiographic diagnosis of knee OA continues to be a challenge. Due to several diagnostic methods and approaches, it is quite difficult to compare different studies. To our knowledge, our studies were the first to show that increased uC2C values can predict the early development (emergence) of knee OA. Therefore, we believe that uC2C has a potential to be a good marker for a diagnosis of a minimal radiographic kOA (gOA grade 1 vs. 0, Paper I). Previous work in this aspect was lacking, partly due to a fact that the presence of JSN and Oph are often merged as a combined scale for OA severity in many studies. Moreover, KL grade 2 is more traditionally accepted as a definitive criterion for kOA, while OA grade 1 (early) is generally considered to indicate a 'doubtful' presence of Ophs underestimating its significance as reported in many publications and clinical practice (Hart & Spector, 2003). Recently, researchers specified that MRI-detected Ophs were associated with OA changes over time (Zhu et al. 2017; Katsuragi et al. 2015). Despite advances in MRI techniques in recent decades, radiography still remains as a primary imaging modality for the definition of inclusion and exclusion criteria for OA-related clinical trials

(Hayashi *et al.* 2018). Our data demonstrate that a use of the NSy system (Nagaosa *et al.* 2000) helps to overcome the limitations of the KL system in the evaluation of early stages of the disease. Therefore, our study underlines a need of careful evaluation of early stages of radiographic OA for biomarker studies.

Moreover, our study clearly showed that an appropriate well-defined control group is essential for the correct evaluation of OA biomarkers. This is particularly important because the incidence of the disease is high all over the world, and its early stages are often underdiagnosed. The question 'who is healthy' is still essential to correctly validate the results of OA studies. We demonstrated that the ability of uC2C to predict an onset of the disease was particularly prominent, when the long-term control group (twelve years without any sign of kOA) was taken into consideration.

It is important to note that because of recent methodological development, a use of two different well-characterized C2C-specific antibodies allowed to increase a specificity of IB-C2C-HUSATM for the measurement of C2C neoepitope fragments in urine. The first antibody effectively captures C2C fragments and the second, tracer antibody, provides a higher specificity by binding to the C2C neoepitope. There are several other *in vitro* ELISA methods available in the market for uC2C estimation that does not provide any information on the utilized antibodies, therefore, a comparison of uC2C levels in different publications is still quite tricky and the methods of estimation need to be compared. Moreover, standardization of the C2C concentrations with the concentration of creatinine in the same urine sample is important for a better consideration of urine dilution factor (A. O. Tamm *et al.* 2014).

The most investigated Col2 degradation marker, uCTX-II, was also observed as a biomarker of EKOA, however, the evidence in this regard is still inconclusive (Cibere et al. 2009; Saberi Hosnijeh et al. 2015; van Spil et al. 2015; P. Wang et al. 2019; Liu et al. 2020). Possible reasons for this could be, the use of KL classification (insensitive for early rkOA), control group selection criteria, or modest study groups (as discussed above). Moreover, comparative observations revealed that uC2C and uCTX-II assays have significant differences and the levels of uC2C correlated weakly with uCTX-II (Cibere et al. 2009; Kraus et al. 2017). Several other markers (serum COMP, sHA, and NTX-I) have been investigated in early-stage kOA, however, no definite conclusion was reached; furthermore, many of these studies have not been reproduced in other populations (Ren & Krawetz, 2018; Zhang, 2018). In addition, a combined usage of Coll2-1NO2, CS846, COMP, and urinary CTX-II provided an additional prediction power for early rkOA over the common demographic predictors such as age, BMI, and sex in the study using the Osteoarthritis Initiative (OAI) database (Liem et al. 2020a). The new approaches to study molecular alterations in EKOA, especially at the pre-kOA stage, are using epigenomics, transcriptomics, proteomics, and metabolomic and lipidomic platforms, a machine learning method to construct predictive models, and monitoring changes in the chondrocyte secretome (Steinberg et al. 2017; Lazzarini et al. 2017; Sanchez et al. 2017). Recently, a specific panel of serum

autoantibodies was detected at baseline in the subjects developing an emerging rkOA (Camacho-Encina *et al.* 2019).

In conclusion, our results demonstrated that uC2C could be a potential biomarker for early-stage kOA studies. Of course, further clinical validation of uC2C is needed. Availability of well-characterized biomarkers on the 'stage of molecular events' certainly opens up an opportunity for the development of new DMOAD and treating a disease in early stages, where metabolic perturbations are frequently considered as reversible (Chu *et al.* 2012).

6.2. uC2C: an early integrative biomarker of different kOA features in distinct joint compartments

As direct examination of cartilage is not suitable at early-stage, and a performance of multiple radiological or MRI examinations of the knee joint is time-consuming and/or expensive, we here present the data that support a use of uC2C as an early dynamic marker for the evaluation of cartilage status and associated processes in the context of kOA. Investigation of several radiological features together in a model clearly demonstrated that the mechanisms underlying uC2C excretion are complex and are associated with the status of Oph and JSN in both knee joint compartments, TF and PF.

Similar to the present study, the previous publications (Wancket *et al.* 2005; van der Kraan & van den Berg, 2007) showed that Oph formation is an important feature of early OA. Moreover, we found that the uC2C level was associated significantly not only with JSN but also with the status of Oph. Irrespective of the presence of JSN, the radiographic presence of marginal Ophs has a high sensitivity, specificity, and positive predictive value for the presence of MRI-detected cartilage defects in the TF joint and meniscal abnormalities (Boegard *et al.* 1998). Histomorphologically, Ophs appear as fibrocartilage with an admixture of cartilaginous and fibrous matrix components, such as Col2 and aggrecan on the one hand and Col1 on the other hand (Gelse *et al.* 2003). At the same time, osteophytic chondrocytes demonstrate an increased expression of tissue-remodelling enzymes (MMP-9, MMP-13, and hyaluronan synthase 1) (Gelse *et al.* 2012), possibly causing a generation of C2C during endochondral ossification in Oph.

It is well-known that radiographic JSN is an insensitive marker in the assessment of early-stage OA (Favero *et al.* 2015; Gly-Jones *et al.* 2015), and a direct examination of the cartilage status may eliminate a risk of underdiagnosis of early cartilage damage. Arthroscopic investigation of articular cartilage have shown that cartilage damage develops years before radiographic JSN (Kijowski *et al.* 2006). We demonstrated in our similarly aged cohort that the presence of cartilage lesions, indicated by the SFA score, might precede radiographic JSN. Furthermore, we also demonstrated that uC2C excretion is substantially associated with the presence of knee cartilage lesions, as determined by macroscopic examination. This finding is consistent with immunohistochemical

results indicating that areas of cartilage damage significantly increased levels of C2C (Wancket *et al.* 2005). The low values of medial SFA scores were characteristic of the subjects with early stages of kOA in our study.

In addition to cartilage thickness on radiographs, a position and degeneration of the meniscus account substantially for the explained variance in JSN (Hunter et al. 2006). Convincing evidence indicates that degenerative meniscal lesion is often suggestive of early-stage kOA (Englund et al. 2009). The menisci are structurally analogous to the surfaces of articular cartilage (Andrews et al. 2017) and are composed of Col1 as well as Col2 (Kambic & McDevitt, 2005). Therefore, we suggest that in addition to cartilage damage, meniscal degeneration is one of the sources of uC2C during OA development.

In recent years, kOA has been conceptualized as a multicompartmental disease (van der Esch *et al.* 2014). Although PFOA may occur in the absence of TFOA (McAlindon *et al.* 1992), a few studies focused only on the PF compartment. The Nottingham scoring system (Nagaosa *et al.* 2000) has a clear advantage here, as it enables a detailed and standardized radiographic evaluation of both knee compartments.

In our study, the presence of OA in an isolated compartment (TF or PF) was a characteristic of early stages of the disease. In advanced cases, a parallel involvement of both compartments was demonstrated. This aspect complicated the analysis of the association between uC2C and each joint compartment. However, we demonstrated a significant association between uC2C level and TFOA. On the other hand, we just found a tendency toward the association with PFOA, probably due to a small number of isolated PFOA cases. Another possible reason could be the distinct biochemical properties (water and proteoglycan content) of patellar cartilage as compared to the tibial and femoral cartilage (Hinman & Crossley, 2007). Nevertheless, the inclusion of the status of both compartments in the models increased the power of prediction of the uC2C level. Thus, while interpreting the uC2C results, especially in early-stage kOA and when the isolated presence is more common, an influence of the OA processes occurring in both compartments (as sources of uC2C) cannot be ignored. Moreover, not only Oph formation but minor cartilage lesions in different knee compartments may occur in EKOA. A combined use of different kOA features formed the basis of our best prediction model. Prediction of uC2C concentration was the best with the SFA score in combination with Ophs in the TF joint and JSN in PF joint.

Taken together, uC2C could serve as a parameter that integrates several important simultaneously on-going features of EKOA and may reflect an early disturbance of Col2 in any knee joint relevant structure.

6.3. uC2C: a biomarker to monitor a severity of disease

To the best of our knowledge, this study was also the first to show that uC2C production continuously increases with the aggravation of structural changes in kOA. We found that the uC2C level could be used to discriminate between the subjects with early and advanced gOA. Moreover, uC2C levels were higher in rapidly progressive cases of the TKR study cohort (Paper III) as compared to the levels of the same disease severity group of our cross-sectional study subset (Paper I). Thus, high levels of uC2C could help us to assess the Col2 degradation preoperatively as well. This phenomenon clearly contradicts to the old understanding that OA is the disease of 'wear and tear'.

In normal cartilage, matrix turnover is strictly regulated by a delicate balance between synthesis and degradation. In OA, this balance is disturbed, usually with enhanced Col2 degradation and synthesis (Sandell & Aigner, 2001). OA is characterized by an excessive damage of collagen fibrillar network, which appears to be primarily mediated by collagenases, especially, MMP-1 and MMP-13 (Poole *et al.* 2003). Thus, the uC2C level reflects the severity of OA changes and indirectly can reflect an activity of these MMPs (Li *et al.* 2007). In the meta-analysis it was revealed that the uCTX-II levels are positively correlated with the OA severity (Huang *et al.* 2017), supporting the finding of the same alteration in Col2 metabolism.

Considering the results of our studies, we can state that, uC2C is a marker of disease severity and we suggest that it could be useful for monitoring of disease activity. Hopefully, it will be reflected in some possible DMOAD treatment effects in the future. However, the fact needed to be evaluated in clinical studies that a decline in CTX-II did not associate with a slowing of radiographic progression in treatment studies (e.g. biphosphonate) as reported in the previous studies (Bingham III *et al.* 2006). This also underlines a need to expand our understanding about the molecular subtypes of the disease.

6.4. uC2C: a prognostic biomarker of kOA radiographic progression

Our observations revealed that the uC2C levels were positively associated with kOA radiographic progression assessed by the NSy (Nagaosa *et al.* 2000). This result is in line with the previous investigations on other populations, where the same uC2C detection method was used (A. Tamm *et al.* 2013; Poole *et al.* 2016; Kraus *et al.* 2017). Unmatching many previously published studies, the majority of the subjects from our study population were middle-aged and had mild kOA radiographic findings. Therefore, osteophytosis was the main radiographic feature at early grades as well as the main additional finding among the kOA progressors. The NSy allowed a better evaluation of osteophytes and was a clear advantage of our studies. One previous study revealed that the patients having KL grade I on radiographs are at a higher risk of structural progression than the

patients with KL grade 0 (Hart & Spector, 2003). Due to the heterogeneity of both OA-phenotypes and evaluated radiographic signs among the publications, a meta-analysis of the studied biochemical markers was not possible, as clearly mentioned in the systematic review (Hosnijeh *et al.* 2015). A lack of clear consensus on a definition of radiographic progression of OA or clinical endpoint creates a huge challenge for defining and validating the biomarkers.

In contrast with previous studies that focused on the radiographic changes only in the TF compartment, we also assessed the progression in the PF compartment. Therefore, we could evaluate the two different levels of radiographic changes – within-the-same grade and intergrade. We observed that higher uC2C values were associated with minimal radiographic progression of kOA within the same gOA grade. When the process expanded into the other joint compartments of the knee, as expected, uC2C predicted a better over-the-grade progression.

These results are more remarkable for the three-year follow-up period, as it is relatively shorter for evaluating ongoing kOA processes radiographically and the progression of kOA may be nonlinear with intermittent periods of stabilization (Kumm *et al.* 2012). Moreover, we revealed that the predictive value of uC2C was less pronounced in more advanced kOA (grades 2 and 3). Although this result may be explained because of a smaller size of the advanced kOA group, it may be a characteristic feature for uC2C, as a biomarker.

Several other biomarkers were associated with the progression of OA and are mentioned in the literature. Out of these the most promising were, cartilagederived Col2 markers and aggregan markers (Bay-Jensen et al. 2022). Recently, a predictive validity of 18 biomarkers was investigated in a large USA biomarker project (FNIH study) and several Col2 synthesis (e.g. PIIANP) or degradation markers (e.g. CTXII) were found associated with two-year radiographic progression of OA (Kraus et al. 2017). The PIIANP assay targets type IIA form (embryonic) of the splice variant of N-terminal propertide of Col2 and characterizes the Col2 syntesis (Rousseau et al. 2004). The low level of Col2 synthesis predicted the progression of JSN or a combination of pain and JSN progression (Kraus et al. 2017). In addition, OA progression is associated with elevated Col2 degradation markers, mainly CTXII. Eight catabolic biomarkers including uC2C, were shown as the best predictors of pain and radiographic progression over two years. The combination of the eight best biomarkers resulted in the best model for progression including uCTXII, sHA, and serum NTXI. However, the prediction power of the best model was relatively low (AUC up to 0.668), possibly due to the heterogeneity of the kOA study group.

In addition, OA-associated cofounders must be taken into account – an increase in BMI slightly magnified the risk of kOA progression. This finding seems to be consistent with other research studies, which reported that the individuals with higher BMIs had a higher rate of osteophyte progression (Zhu *et al.* 2018).

In summary, the uC2C values appear to be dependent on at least two factors – severity of gOA grade and the existence of radiographic kOA

progression. It is therefore likely that for estimating a risk of kOA progression, the baseline uC2C level in gOA grade must be considered; in the same stage of radiographic kOA, uC2C was found higher in progressors than in non-progressors.

However, further studies are warranted to clarify clinical implications of uC2C.

Genetic markers of OA have not been addressed in this work because they form a completely separate and very large topic, which is not associated directly with the thesis. However, an interesting topic in OA research is opened with small RNA molecules. MicroRNAs, small and stable noncoding RNA molecules, have emerged as powerful candidates for biomarkers in musculoskeletal disorders because of their impact on the expression of tissue proteins. (Ali *et al.* 2022). MicroRNA profiling, also called as 'liquid biopsies' identified that miR-320 family members are associated with fast-progressing radiographic kOA in the Osteoarthritis Initiative cohort.

6.5. uC2C: a biomarker to monitor pre-TKR and post-TKR response

To date, only a limited number of studies have investigated the behaviour of biomarkers after TKR – probably due to the understanding that the operation is the 'end-stage' of OA. Our study with the TKR cohort is the first to present the dynamics of uC2C levels in the subjects in pre- and post-TKR periods. Surprisingly, the uC2C level of the whole group was still high (not decreased) after three months of TKR but reverted to the baseline level after one year of the operation. Thus, the postoperative uC2C levels remained relatively high and did not drop to the level of the subjects without kOA or with EKOA. However, we found that the individual dynamics of uC2C levels were rather heterogeneous, this means, some patients had an increase, while others had a decrease.

To our knowledge, four similar studies evaluating the dynamics of cartilage biomarkers in serum (keratan sulfate, COMP, and Col2-derived fragments Coll2–1 and Coll2–1NO2) were performed in TKR patients (Sweet *et al.* 1988; Sharif *et al.* 2004; Deberg *et al.* 2008; Endres *et al.* 2020). A quite old study by Sweet *et al.* reported a short preliminary decrease in keratan sulphate levels one week after TKR, which subsequently increased up to preoperative levels after six months (Sweet *et al.* 1988). Sharif *et al.* demonstrated a substantial rise in serum COMP levels after TKR in all patients, persisting up to 12 months following the surgery (Sharif *et al.* 2004). Recently, Endres *et al.* showed a short reduction in COMP levels after a week of total hip replacement; later on, these levels returned to the preoperative baseline (Endres *et al.* 2020). These results were unexpected, as a decrease in COMP levels was expected, and the authors were forced to consider a possibility that COMP could also originate from the sources other than the operated joint alone. It was assumed that there might be an increased production or degradation of COMP in the contralateral

knee, or an increased release of COMP from other tissues – such as tendons, ligaments, and capsules – of the TKR joint. Endres *et al.* attributed the results to changes in physical activity – patients with recovery were much more mobile than pre-TKR. Similarly, a study of Coll2–1 and Coll2–1NO2 showed that the pathogenic metabolism of Col2 persisted after TKR (Deberg *et al.* 2008).

Summarizing the conclusions from these few studies on variations in biomarkers after total joint replacement, the level of several degradation biomarkers (e.g., uC2C, Coll2–1NO2, COMP) do not decrease after joint replacement. Thus, in general, there is scope for interpretation that TKR does not stop the breakdown of Col2 and possible synovitis (as reflected by COMP levels). It has been suggested that a sustained level of Col2 degradation markers after TKR can be attributed to a persistent Col2 degradation in different structures of a synovial joint, although a presence of OA lesions in other joints cannot be ruled out and could be considered as a secondary source. We presented a quite common involvement of both knees (in a quarter of cases); however, we found no associations between uC2C levels and previous history of TKR.

Nevertheless, OA has been proposed as a systemic disease (Visser *et al.* 2015), and its burden can be thought of in terms of severity and the number of joints involved (Henrotin *et al.* 2007). However, in a cross-sectional study (Paper I) we found that the uC2C level did not differ between unilateral and bilateral OA cases. Moreover, we did not prove that the uC2C level is significantly influenced by the involvement of other joints assessed by pain. Of course, a more detailed investigation – not just of pain – is needed to measure the total burden of OA (Addison *et al.* 2009). In addition, several authors have demonstrated that rather a large proportion of kOA patients may have a generalized joint disease (Stürmer *et al.* 1998; Kraus *et al.* 2010).

In conclusion, though we did not find a significant reduction in the uC2C level after 12 months of TKR, our study can be considered as the first attempt to investigate the behaviour of uC2C in monitoring the treatment effect of kOA.

6.6. uC2C: a predictive biomarker for clinical outcome of TKR

An improvement in quality of life and knee function is expected after TKR, however, a great heterogeneity is seen. Here, we demonstrated only a partial improvement in KOOS subscales after 1 year of TKR. This was particularly apparent in terms of restrictions in the use of legs for more demanding functions (expressed by KOOSsp/recr). Thus, a substantial proportion of these patients require post-surgery rehabilitation – especially exercise therapy – also years after TKR.

We demonstrated that higher preoperative uC2C level could predict the improvement in the KOOSsymp scale after surgery in women (OR > 9). There is no good explanation for why the reduction of pain did not associate with a drop in uC2C, but the pain and other symptoms of kOA probably present different

aspects of the disease. A possible explanation is that pain in OA could be multifactorial (Neogi, 2013), and may arise from multiple structural changes in the knee joint (Yusuf *et al.* 2011).

Interestingly, we presented that the preoperative uC2C values could predict its postoperative status; this means, if the preoperative value was low, Coll2 degradation had become more intense, and vice versa was true for high preoperative values. Likewise, in the patients with preoperative Coll2–1NO2 (a biomarker of oxidative damage) levels above the median showed a significant and progressive decrease postoperatively but, they tended to increase in the patients with preoperative Coll2–1NO2 values below the median (Deberg *et al.* 2008). If this phenomenon is confirmed by other studies, there may be an opportunity for an additional grouping of the patients undergoing TKR surgery. The possibility of a more precise categorization of patients became apparent here.

In total, this observation may support the hypothesis that the postoperative change in uC2C levels depends on its preoperative level.

6.7. Sex-related differences of uC2C level in the course of kOA

Although there was no general difference found in the uC2C excretion between the sexes in the cross-sectional study, several special sex-based features of uC2C in the incidence and course of kOA were noted.

At first, the mean value of uC2C in the male non-progressors was significantly higher than the female non-progressors, including the results in the long-term control group. This finding contrasts with a large FNIH/OARSI biomarker consortium study suggesting no effect of sex on reference levels of uC2C (Kraus *et al.* 2017). The reason for this contradiction is unclear, but it might be related to different subject characteristics, especially age, which was significantly lower in our study. Moreover, higher BMI in the progressors as compared to the non-progressors females was the characteristic of our longitudinal study.

Secondly, the best model for uC2C prediction emerged by a combination of Ophs and JSN, and especially in TFOA; it was almost 2-fold better in females than in males. Moreover, a combination of the radiographic signs like, Ophs and JSN were more common in females. In addition, we demonstrated that uC2C is a promising prognostic marker for emerging kOA in females but not in males; a gradual increase of uC2C level was associated with the radiological grades only in females, except the male progressors with more severe kOA grades.

Thirdly, we observed that the preoperative uC2C values could predict the postoperative trends more prominently in females than males. We found that low preoperative value was probably related to intense Col2 degradation in postoperative period, and vice versa was true for high preoperative values. Unlike in males, there was a significant decrease in the uC2C levels in females after one year of TKR.

Finally, the preoperative uC2C levels could predict an improvement in the KOOS symptoms score only in females. At the same time, the KOOS pain score was correlated with the uC2C value in men during the preoperative period.

The sex-related differences in the course of OA and probably in its pathogenesis, should be highlighted. Albeit sex-based differences in OA have been reported, they might be often overlooked by researchers (Boyan et al. 2013; Bihlet et al. 2019). Women are associated with a higher prevalence and severity of OA as compared to men, particularly following menopause (Srikanth et al. 2005). Gender discrepancies may be caused due to differences in hormones, bone strength, alignment, ligament laxity, and neuromuscular strength (Johnson & Hunter, 2014). A systematic review of risk factors for the onset of knee pain showed that one of the strongest factors associated with knee pain was female sex (Silverwood et al. 2015). A recent study in the Estonian Early Knee OA Study Cohort demonstrated a presence of significant sex-dependent differences in cytokine production, with predominant angiogenesis in females with grade 1 kOA, whereas activation of tissue remodelling was predominated in males (Kisand et al. 2018), indicating sex-related differences in the pathways of kOA pathogenesis. Recognition of the sex-based differences related to uC2C may aid the assessment of the efficacy of novel therapeutic agents that directly suppress MMP gene expression (Malemud, 2019) or angiogenesis (Hamilton et al. 2016). Nevertheless, further studies with larger sample sizes are needed to clarify the role of sex-related differences in uC2C throughout the course of OA.

According to these data, we can infer that the uC2C level behaves differently in males and females in kOA and could be a suitable kOA marker for females exclusively.

6.8. Limitations of the study

The study had several limitations. Main limitation was a small number of subjects in some sub-groups, which decreased a statistical power of inter-group comparison and models and could have reduced the chance of detecting a true effect of the results. It prevented drawing definitive conclusions about the utility of uC2C in some special aspects of kOA. Firstly, SFA scores and uC2C concentrations were determined simultaneously in a small number of crosssectional study subjects, therefore a statistical power of the models including SFA determination was limited (Paper I). Secondly, a prevalence of the crosssectional study subjects with isolated OA changes, especially JSN, was low (Paper I). Moreover, we had a limited number of cases with exclusive JSN progression; thus, we could indirectly interpret that the increased level of uC2C reflected the risk of subsequent cartilage damage (Paper II). Despite this, the association of a high level of uC2C with growing Oph is considered a prognostic of the progression of kOA. Thirdly, a low prevalence of the individuals with advanced kOA cases (gOA grade 2-3), especially in separate gender groups, complicated the conclusions at this stage of the disease. (Paper II, III). In order to confirm the results, future work should evaluate uC2C in several directions such as, using a larger population cohort with a higher prevalence of advanced kOA cases and recording the observations at intermediate timepoints with different imaging modalities. Additionally, an observation period more than three years would be required. Moreover, although the TKR study group was sufficient in number of subjects to allow a general assessment of uC2C behaviour in surgical patients, a larger cohort is needed for the evaluation of sex-related differences and additional relationships in the pathogenesis of kOA (Paper III). Also, one year may be too short period to assess far-reaching changes in OA biomarkers after TKR.

Next, we exclusively investigated uC2C as a single biomarker for an integration of several possible signs of kOA (Paper I-III) and did not measure the Col2 synthesis at the same time; therefore, we could not evaluate a balance between the synthesis and degradation of Col2. A combination of various biomarkers with uC2C would probably improve a prediction of the disease progression (Kraus *et al.* 2017). Investigation of a different set of biomarkers would help to explore certain phenotypes of OA (Mobasheri *et al.* 2019a; van Spil *et al.* 2019).

The following limitation should be considered regarding the adjustments. We adjusted the LM and GLM models for age, sex, and BMI as generally accepted OA risk factors, but not for the other possible confounders, e.g. menopausal status (Paper I-III). As uC2C was also observed in deep calcified cartilage (Wancket *et al.* 2005), it may be influenced by menopause like the markers of bone turnover.

Finally, although we used sumVAS for the assessment of pain in other joints (excluding the knee joint) (Paper II, III), the presence and progression of osteoarthritis at other sites were not excluded because the radiographs of other joints were not available (Paper I-III). It could cause confounding effects with regard to uC2C levels.

In summary, although the findings of our study demonstrate the potential roles of uC2C as a risk, diagnostic and prognostic marker, further validation and qualification are needed for its clinical use (*in vivo* diagnostics). In future studies, sex-specific differences in the pathogenesis of kOA should also be addressed.

7. CONCLUSIONS

• uC2C is an integrative marker for kOA; it is simultaneously associated with the main pathological processes of OA like, cartilage degradation and osteophytes (Ophs) formation development in the PF and TF compartments.

By regression analysis, we demonstrated that Oph describe uC2C level better than joint space narrowing (JSN). However, the best prediction was achieved by a combination of both radiographic features assessed separately in TF and PF joints, in the model. Replacement of radiographic TF JSN with macroscopically assessed cartilage lesions of the arthroscopy (SFA score) further improved the prediction power of the model for the uC2C level.

• uC2C is a good candidate for the development of an early diagnostic test for kOA.

We have shown that an increase in uC2C concentration already exists in the early stages of the disease (radiographic grade 1 by NSy), which is generally considered as a pre-radiological stage according to the KL evaluation system.

• uC2C is a potential kOA risk prediction marker in females.

Generalized linear models (GLM) analysis showed that a higher baseline value of uC2C is an excellent predictor of kOA initiation in women (gOA 0 becomes gOA 1) over the follow-up of 3 years. The best prediction value for the model (> 90%) was obtained by comparing the emerging kOA group with the long-term (12 years) control group.

• uC2C is higher in progressors than in non-progressors in the same radiographic severity stage kOA. For a proper clinical evaluation of uC2C, two aspects are important: 1) uC2C levels are positively associated with severity of kOA (uC2C is higher in higher radiographic grades); and 2) higher uC2C is associated with ongoing disease progression, particularly in females. The values of uC2C are highest in rapid progressors like pre-operative TKR cases.

We found that median uC2C values gradually elevate with the gOA grade, indicating a moderate diagnostic performance at each grade as compared to the previous grade. At each severity stage, a higher value of uC2C predicts the progression of kOA over the next 3 years. We found that uC2C is a sensitive marker of progression; this means, it predicts minimal radiographic changes within the same gOA grade like the addition of Oph or the worsening of JSN in any compartment of the knee joint.

• Following total knee replacement (TKR), the dynamics of uC2C are quite heterogeneous; excretion of uC2C may decrease, increase, or can remain unchanged.

We found that relatively higher baseline uC2C values were associated with a declining trend after TKR, and in contrast, the degradation of Col2 was accelerated after surgery in the subjects with low preoperative uC2C values. Thus, TKR could not stop or reduce the degradation of Col2 in the majority of cases. Baseline uC2C was also shown to predict postoperative improvement in KOOS symptom scores in females. We found no association between uC2C and pain score.

• uC2C appears as a better diagnostic and prognostic biomarker in females than in males.

We found that uC2C predicts the progression of kOA more accurately in females than in males; female uC2C values showed a positive correlation with radiographic severity grade. In males, this association was weak and less significant on the statistical scale. However, it should be noted, that the sample size of the male group was relatively smaller and might be a reason for low statistical power. Therefore, these results need to be confirmed on a larger sample size and different populations.

8. REFERENCES

- Abramson, S. B., & Attur, M. (2009). Developments in the scientific understanding of osteoarthritis. *Arthritis Research & Therapy*, 11(3), 227. https://doi.org/10.1186/ar2655
- Addison, S., Coleman, R. E., Feng, S., McDaniel, G., & Kraus, V. B. (2009). Whole-body bone scintigraphy provides a measure of the total-body burden of osteoarthritis for the purpose of systemic biomarker validation. *Arthritis and Rheumatism*, 60(11), 3366–3373. https://doi.org/10.1002/art.24856
- Aggarwal, R., Ringold, S., Khanna, D., Neogi, T., Johnson, S. R., Miller, A., Brunner,
 H. I., Ogawa, R., Felson, D., Ogdie, A., Aletaha, D., & Feldman, B. M. (2015).
 Distinctions Between Diagnostic and Classification Criteria? Arthritis Care & Research, 67(7), 891–897. https://doi.org/10.1002/acr.22583
- Akhtar, N., & Haqqi, T. M. (2011). Epigallocatechin-3-gallate suppresses the global interleukin-1beta-induced inflammatory response in human chondrocytes. *Arthritis Research & Therapy*, *13*(3), R93. https://doi.org/10.1186/ar3368
- Alexander, L. C., Jr, McHorse, G., Huebner, J. L., Bay-Jensen, A.-C., Karsdal, M. A., & Kraus, V. B. (2021). A matrix metalloproteinase-generated neoepitope of CRP can identify knee and multi-joint inflammation in osteoarthritis. *Arthritis Research & Therapy*, 23(1), 226. https://doi.org/10.1186/s13075-021-02610-y
- Alexandersen, P., Karsdal, M. A., Byrjalsen, I., & Christiansen, C. (2011). Strontium ranelate effect in postmenopausal women with different clinical levels of osteoarthritis. *Climacteric: The Journal of the International Menopause Society*, *14*(2), 236–243. https://doi.org/10.3109/13697137.2010.507887
- Ali, S. A., Espin-Garcia, O., Wong, A. K., Potla, P., Pastrello, C., McIntyre, M., Lively, S., Jurisica, I., Gandhi, R., & Kapoor, M. (2022). Circulating microRNAs differentiate fast-progressing from slow-progressing and non-progressing knee osteoarthritis in the Osteoarthritis Initiative cohort. *Therapeutic Advances in Musculoskeletal Disease*, 14, 1759720X221082917. https://doi.org/10.1177/1759720X221082917
- Allen, K. D., Thoma, L. M., & Golightly, Y. M. (2021). Epidemiology of osteoarthritis. *Osteoarthritis and Cartilage*, S1063458421008864. https://doi.org/10.1016/j.joca. 2021.04.020
- Altman, R., Asch, E., Bloch, D., Bole, G., Borenstein, D., Brandt, K., Christy, W., Cooke, T. D., Greenwald, R., Hochberg, M., Howell, D., Kaplan, D., Koopman, W., Longley, S., Mankin, H., McShane, D. J., Medsger, T., Meenan, R., Mikkelsen, W., ... Wolfe, F. (1986). Development of criteria for the classification and reporting of osteoarthritis: Classification of osteoarthritis of the knee. *Arthritis & Rheumatism*, 29(8), 1039–1049. https://doi.org/10.1002/art.1780290816
- Altman, R. D., & Gold, G. E. (2007). Atlas of individual radiographic features in osteo-arthritis, revised. *Osteoarthritis and Cartilage*, *15*, A1–A56. https://doi.org/10.1016/j.joca.2006.11.009
- Anderson, A. S., & Loeser, R. F. (2010). Why is Osteoarthritis an Age-Related Disease? *Best Practice & Research. Clinical Rheumatology*, 24(1), 15. https://doi.org/10.1016/j.berh.2009.08.006
- Andrews, S. H. J., Adesida, A. B., Abusara, Z., & Shrive, N. G. (2017). Current concepts on structure–function relationships in the menisci. *Connective Tissue Research*, 58(3–4), 271–281. https://doi.org/10.1080/03008207.2017.1303489

- Andriacchi, T. P., Mündermann, A., Smith, R. L., Alexander, E. J., Dyrby, C. O., & Koo, S. (2004). A framework for the in vivo pathomechanics of osteoarthritis at the knee. *Annals of Biomedical Engineering*, 32(3), 447–457. https://doi.org/10.1023/b:abme.0000017541.82498.37
- Ashraf, S., Wibberley, H., Mapp, P. I., Hill, R., Wilson, D., & Walsh, D. A. (2011). Increased vascular penetration and nerve growth in the meniscus: A potential source of pain in osteoarthritis. *Annals of the Rheumatic Diseases*, 70(3), 523–529. https://doi.org/10.1136/ard.2010.137844
- Aurich, M., Hofmann, G. O., & Rolauffs, B. (2017). Differences in type II collagen turnover of osteoarthritic human knee and ankle joints. *International Orthopaedics*, 41(5), 999–1005. https://doi.org/10.1007/s00264-017-3414-5
- Ayral, X., Pickering, E. H., Woodworth, T. G., Mackillop, N., & Dougados, M. (2005). Synovitis: A potential predictive factor of structural progression of medial tibiofemoral knee osteoarthritis results of a 1 year longitudinal arthroscopic study in 422 patients. *Osteoarthritis and Cartilage*, *13*(5), 361–367. https://doi.org/10.1016/j.joca.2005.01.005
- Bastick, A. N., Belo, J. N., Runhaar, J., & Bierma-Zeinstra, S. M. A. (2015). What Are the Prognostic Factors for Radiographic Progression of Knee Osteoarthritis? A Meta-analysis. *Clinical Orthopaedics and Related Research*, 473(9), 2969–2989. https://doi.org/10.1007/s11999-015-4349-z
- Bastick, A. N., Runhaar, J., Belo, J. N., & Bierma-Zeinstra, S. M. A. (2015). Prognostic factors for progression of clinical osteoarthritis of the knee: A systematic review of observational studies. *Arthritis Research & Therapy*, 17(1), 152. https://doi.org/10.1186/s13075-015-0670-x
- Bauer, D. C., Hunter, D. J., Abramson, S. B., Attur, M., Corr, M., Felson, D., Heinegård, D., Jordan, J. M., Kepler, T. B., Lane, N. E., Saxne, T., Tyree, B., & Kraus, V. B. (2006). *Classification of osteoarthritis biomarkers: A proposed approach*. *14*(8), 723–727. https://doi.org/10.1016/j.joca.2006.04.001
- Bay-Jensen, A. C., Mobasheri, A., Thudium, C. S., Kraus, V. B., & Karsdal, M. A. (2022). Blood and urine biomarkers in osteoarthritis an update on cartilage associated type II collagen and aggrecan markers. *Current Opinion in Rheumatology*, *34*(1), 54–60. https://doi.org/10.1097/BOR.00000000000000845
- Bay-Jensen, A. C., Reker, D., Kjelgaard-Petersen, C. F., Mobasheri, A., Karsdal, M. A., Ladel, C., Henrotin, Y., & Thudium, C. S. (2016). Osteoarthritis year in review 2015: Soluble biomarkers and the BIPED criteria. *Osteoarthritis and Cartilage*, 24(1), 9–20. https://doi.org/10.1016/j.joca.2015.10.014
- Bedson, J., & Croft, P. R. (2008). The discordance between clinical and radiographic knee osteoarthritis: A systematic search and summary of the literature. *BMC Musculoskeletal Disorders*, 9, 116. https://doi.org/10.1186/1471-2474-9-116
- Bellamy, N., Buchanan, W. W., Goldsmith, C. H., Campbell, J., & Stitt, L. W. (1988). Validation study of WOMAC: A health status instrument for measuring clinically important patient relevant outcomes to antirheumatic drug therapy in patients with osteoarthritis of the hip or knee. *The Journal of Rheumatology*, 15(12), 1833–1840.
- Belluzzi, E., El Hadi, H., Granzotto, M., Rossato, M., Ramonda, R., Macchi, V., De Caro, R., Vettor, R., & Favero, M. (2017). Systemic and Local Adipose Tissue in Knee Osteoarthritis. *Journal of Cellular Physiology*, *232*(8), 1971–1978. https://doi.org/10.1002/jcp.25716

- Benito, M., Veale, D., FitzGerald, O., van den Berg, W. B., & Bresnihan, B. (2005). Synovial tissue inflammation in early and late osteoarthritis. *Annals of the Rheumatic Diseases*, 64(9), 1263–1267. https://doi.org/10.1136/ard.2004.025270
- Berenbaum, F. (2013). Osteoarthritis as an inflammatory disease (osteoarthritis is not osteoarthrosis!). *Osteoarthritis and Cartilage*, 21(1), 16–21. https://doi.org/10.1016/j.joca.2012.11.012
- Berenbaum, F., Griffin, T. M., & Liu-Bryan, R. (2017). Metabolic Regulation of Inflammation in Osteoarthritis. *Arthritis & Rheumatology (Hoboken, N.J.)*, 69(1), 9–21. https://doi.org/10.1002/art.39842
- Berenbaum, F., Wallace, I. J., Lieberman, D. E., & Felson, D. T. (2018). Modern-day environmental factors in the pathogenesis of osteoarthritis. *Nature Reviews Rheumatology*, *14*(11), 674–681. https://doi.org/10.1038/s41584-018-0073-x
- Bettica, P., Cline, G., Hart, D. J., Meyer, J., & Spector, T. D. (2002). Evidence for increased bone resorption in patients with progressive knee osteoarthritis: Longitudinal results from the Chingford study. *Arthritis and Rheumatism*, 46(12), 3178–3184. https://doi.org/10.1002/art.10630
- Bhattacharyya, T., Gale, D., Dewire, P., Totterman, S., Gale, M. E., McLaughlin, S., Einhorn, T. A., & Felson, D. T. (2003). The Clinical Importance of Meniscal Tears Demonstrated by Magnetic Resonance Imaging in Osteoarthritis of the Knee*. *JBJS*, 85(1), 4–9. http://journals.lww.com/jbjsjournal/Fulltext/2003/01000/The_Clinical_Importance_of_Meniscal_Tears.2.aspx
- Bierma-Zeinstra, S. M., & van Middelkoop, M. (2017). In search of phenotypes. *Nature Reviews Rheumatology*, *13*(12), 705–706. https://doi.org/10.1038/nrrheum.2017.181
- Bihlet, A. R., Bjerre-Bastos, J. J., Andersen, J. R., Byrjalsen, I., Karsdal, M. A., & Bay-Jensen, A.-C. (2020). Clinical and biochemical factors associated with risk of total joint replacement and radiographic progression in osteoarthritis: Data from two phase III clinical trials. *Seminars in Arthritis and Rheumatism*, 50(6), 1374–1381. https://doi.org/10.1016/j.semarthrit.2020.03.002
- Bihlet, A. R., Byrjalsen, I., Bay-Jensen, A.-C., Andersen, J. R., Christiansen, C., Riis, B. J., & Karsdal, M. A. (2019). Associations between biomarkers of bone and cartilage turnover, gender, pain categories and radiographic severity in knee osteoarthritis. *Arthritis Research & Therapy*, 21. https://doi.org/10.1186/s13075-019-1987-7
- Billinghurst, R. C., Dahlberg, L., Ionescu, M., Reiner, A., Bourne, R., Rorabeck, C., Mitchell, P., Hambor, J., Diekmann, O., Tschesche, H., Chen, J., Van Wart, H., & Poole, A. R. (1997). Enhanced cleavage of type II collagen by collagenases in osteoarthritic articular cartilage. *Journal of Clinical Investigation*, 99(7), 1534–1545. https://doi.org/10.1172/JCI119316
- Bingham III, C. O., Buckland-Wright, J. C., Garnero, P., Cohen, S. B., Dougados, M., Adami, S., Clauw, D. J., Spector, T. D., Pelletier, J.-P., Raynauld, J.-P., Strand, V., Simon, L. S., Meyer, J. M., Cline, G. A., & Beary, J. F. (2006). Risedronate decreases biochemical markers of cartilage degradation but does not decrease symptoms or slow radiographic progression in patients with medial compartment osteoarthritis of the knee: Results of the two-year multinational knee osteoarthritis structural arthritis study. *Arthritis & Rheumatism*, 54(11), 3494–3507. https://doi.org/10.1002/art.22160
- Biomarkers Definitions Working Group. (2001). Biomarkers and surrogate endpoints: Preferred definitions and conceptual framework. *Clinical Pharmacology and Therapeutics*, 69(3), 89–95. https://doi.org/10.1067/mcp.2001.113989

- Bjerre-Bastos, J. J., Nielsen, H. B., Andersen, J. R., Karsdal, M., Bay-Jensen, A.-C., Boesen, M., Mackey, A. L., Byrjalsen, I., & Bihlet, A. R. (2021). Does moderate intensity impact exercise and non-impact exercise induce acute changes in collagen biochemical markers related to osteoarthritis? An exploratory randomized crossover trial. Osteoarthritis and Cartilage, 29(7), 986–994. https://doi.org/10.1016/j.joca.2021.02.569
- Blanco, F. J., Silva-Díaz, M., Quevedo Vila, V., Seoane-Mato, D., Pérez Ruiz, F., Juan-Mas, A., Pego-Reigosa, J. M., Narváez, J., Quilis, N., Cortés, R., Romero Pérez, A., Fábregas Canales, D., Font Gayá, T., Bordoy Ferrer, C., Sánchez-Piedra, C., Díaz-González, F., & Bustabad-Reyes, S. (2021). Prevalence of symptomatic osteo-arthritis in Spain: EPISER2016 study*. *Reumatología Clínica (English Edition)*, 17(8), 461–470. https://doi.org/10.1016/j.reumae.2020.01.005
- Boegard, T., Rudling, O., Petersson, I., & Jonsson, K. (1998). Correlation between radiographically diagnosed osteophytes and magnetic resonance detected cartilage defects in the tibiofemoral joint. *Annals of the Rheumatic Diseases*, *57*(7), 401–407. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1752666/
- Borzi, R. M., Mazzetti, I., Macor, S., Silvestri, T., Bassi, A., Cattini, L., & Facchini, A. (1999). Flow cytometric analysis of intracellular chemokines in chondrocytes in vivo: Constitutive expression and enhancement in osteoarthritis and rheumatoid arthritis. *FEBS Letters*, 455(3), 238–242. https://doi.org/10.1016/s0014-5793(99)00886-8
- Boyan, B. D., Tosi, L. L., Coutts, R. D., Enoka, R. M., Hart, D. A., Nicolella, D. P., Berkley, K. J., Sluka, K. A., Kwoh, C. K., O'Connor, M. I., Kohrt, W. M., & Resnick, E. (2013). Addressing the gaps: Sex differences in osteoarthritis of the knee. *Biology of Sex Differences*, 4(1), 4. https://doi.org/10.1186/2042-6410-4-4
- Brandt, K. D., Doherty, M., & Lohmander, L. S. (2003). *Osteoarthritis* (Second Edition). Oxford University Press.
- Burr, D. B., & Gallant, M. A. (2012). Bone remodelling in osteoarthritis. *Nature Reviews. Rheumatology*, 8(11), 665–673. https://doi.org/10.1038/nrrheum.2012.130
- Cahue, S., Sharma, L., Dunlop, D., Ionescu, M., Song, J., Lobanok, T., King, L., & Poole, A. R. (2007). The ratio of type II collagen breakdown to synthesis and its relationship with the progression of knee osteoarthritis. *Osteoarthritis and Cartilage*, 15(7), 819–823. https://doi.org/10.1016/j.joca.2007.01.016
- Camacho-Encina, M., Balboa-Barreiro, V., Rego-Perez, I., Picchi, F., VanDuin, J., Qiu, J., Fuentes, M., Oreiro, N., LaBaer, J., Ruiz-Romero, C., & Blanco, F. J. (2019). Discovery of an autoantibody signature for the early diagnosis of knee osteoarthritis: Data from the Osteoarthritis Initiative. *Annals of the Rheumatic Diseases*, 78(12), 1699–1705. https://doi.org/10.1136/annrheumdis-2019-215325
- Case, R., Thomas, E., Clarke, E., & Peat, G. (2015). Prodromal symptoms in knee osteoarthritis: A nested case–control study using data from the Osteoarthritis Initiative. *Osteoarthritis and Cartilage*, 23(7), 1083–1089. https://doi.org/10.1016/j.joca.2014.12.026
- Cheng, H., Hao, B., Sun, J., & Yin, M. (2020). C-Terminal Cross-Linked Telopeptides of Type II Collagen as Biomarker for Radiological Knee Osteoarthritis: A Meta-Analysis. *Cartilage*, 11(4), 512–520. https://doi.org/10.1177/1947603518798884
- Choi, J., Joseph, L., & Pilote, L. (2013). Obesity and C-reactive protein in various populations: A systematic review and meta-analysis. *Obesity Reviews: An Official Journal of the International Association for the Study of Obesity*, 14(3), 232–244. https://doi.org/10.1111/obr.12003

- Chu, C. R., Williams, A. A., Coyle, C. H., & Bowers, M. E. (2012). Early diagnosis to enable early treatment of pre-osteoarthritis. *Arthritis Research & Therapy*, 14(3), 212. https://doi.org/10.1186/ar3845
- Cibere, J., Zhang, H., Garnero, P., Poole, A. R., Lobanok, T., Saxne, T., Kraus, V. B., Way, A., Thorne, A., Wong, H., Singer, J., Kopec, J., Guermazi, A., Peterfy, C., Nicolaou, S., Munk, P. L., & Esdaile, J. M. (2009). Association of biomarkers with pre–radiographically defined and radiographically defined knee osteoarthritis in a population-based study. *Arthritis & Rheumatism*, 60(5), 1372–1380. https://doi.org/10.1002/art.24473
- Collins, J. E., Neogi, T., & Losina, E. (2021). Trajectories of Structural Disease Progression in Knee Osteoarthritis. *Arthritis Care & Research*, 73(9), 1354–1362. https://doi.org/10.1002/acr.24340
- Collins, K. H., Lenz, K. L., Pollitt, E. N., Ferguson, D., Hutson, I., Springer, L. E., Oestreich, A. K., Tang, R., Choi, Y.-R., Meyer, G. A., Teitelbaum, S. L., Pham, C. T. N., Harris, C. A., & Guilak, F. (2021). Adipose tissue is a critical regulator of osteoarthritis. *Proceedings of the National Academy of Sciences of the United States of America*, 118(1), e2021096118. https://doi.org/10.1073/pnas.2021096118
- Collins, K. H., Paul, H. A., Reimer, R. A., Seerattan, R. A., Hart, D. A., & Herzog, W. (2015). Relationship between inflammation, the gut microbiota, and metabolic osteoarthritis development: Studies in a rat model. *Osteoarthritis and Cartilage*, 23(11), 1989–1998. https://doi.org/10.1016/j.joca.2015.03.014
- Conaghan, P. G., Kloppenburg, M., Schett, G., Bijlsma, J. W. J., & EULAR osteoarthritis ad hoc committee. (2014). Osteoarthritis research priorities: A report from a EULAR ad hoc expert committee. *Annals of the Rheumatic Diseases*, 73(8), 1442–1445. https://doi.org/10.1136/annrheumdis-2013-204660
- Conde, J., Gomez, R., Bianco, G., Scotece, M., Lear, P., Dieguez, C., Gomez-Reino, J., Lago, F., & Gualillo, O. (2011). Expanding the adipokine network in cartilage: Identification and regulation of novel factors in human and murine chondrocytes. *Annals of the Rheumatic Diseases*, 70(3), 551–559. https://doi.org/10.1136/ard. 2010.132399
- Coryell, P. R., Diekman, B. O., & Loeser, R. F. (2021). Mechanisms and therapeutic implications of cellular senescence in osteoarthritis. *Nature Reviews. Rheumatology*, 17(1), 47–57. https://doi.org/10.1038/s41584-020-00533-7
- Crema, M. D., Roemer, F. W., Zhu, Y., Marra, M. D., Niu, J., Zhang, Y., Lynch, J. A., Javaid, M. K., Lewis, C. E., El-Khoury, G. Y., Felson, D. T., & Guermazi, A. (2010). Subchondral Cystlike Lesions Develop Longitudinally in Areas of Bone Marrow Edema–like Lesions in Patients with or at Risk for Knee Osteoarthritis: Detection with MR Imaging—The MOST Study1. *Radiology*, 256(3), 855–862. https://doi.org/10.1148/radiol.10091467
- Cui, A., Li, H., Wang, D., Zhong, J., Chen, Y., & Lu, H. (2020). Global, regional prevalence, incidence and risk factors of knee osteoarthritis in population-based studies. *EClinicalMedicine*, 29–30, 100587. https://doi.org/10.1016/j.eclinm.2020. 100587
- Daghestani, H. N., & Kraus, V. B. (2015). Inflammatory biomarkers in osteoarthritis. *Osteoarthritis and Cartilage*, 23(11), 1890–1896. https://doi.org/10.1016/j.joca. 2015.02.009
- Dam, E. B., Loog, M., Christiansen, C., Byrjalsen, I., Folkesson, J., Nielsen, M., Qazi, A. A., Pettersen, P. C., Garnero, P., & Karsdal, M. A. (2009). Identification of

- progressors in osteoarthritis by combining biochemical and MRI-based markers. *Arthritis Research & Therapy*, 11(4), R115. https://doi.org/10.1186/ar2774
- Davis, J. E., Liu, S.-H., Lapane, K., Harkey, M. S., Price, L. L., Lu, B., Lo, G. H., Eaton, C. B., Barbe, M. F., McAlindon, T. E., & Driban, J. B. (2018). Adults with incident accelerated knee osteoarthritis are more likely to receive a knee replacement: Data from the Osteoarthritis Initiative. *Clinical Rheumatology*, *37*(4), 1115–1118. https://doi.org/10.1007/s10067-018-4025-2
- Day, J. S., Ding, M., van der Linden, J. C., Hvid, I., Sumner, D. R., & Weinans, H. (2001). A decreased subchondral trabecular bone tissue elastic modulus is associated with pre-arthritic cartilage damage. *Journal of Orthopaedic Research: Official Publication of the Orthopaedic Research Society*, 19(5), 914–918. https://doi.org/10.1016/S0736-0266(01)00012-2
- Deberg, M. A., Labasse, A. H., Collette, J., Seidel, L., Reginster, J.-Y., & Henrotin, Y. E. (2005). One-year increase of Coll 2-1, a new marker of type II collagen degradation, in urine is highly predictive of radiological OA progression. *Osteoarthritis and Cartilage*, *13*(12), 1059–1065. https://doi.org/10.1016/j.joca.2005.06.014
- Deberg, M., Dubuc, J.-E., Labasse, A., Sanchez, C., Quettier, E., Bosseloir, A., Crielaard, J.-M., & Henrotin, Y. (2008). One-year follow-up of Coll2-1, Coll2-1NO2 and myeloperoxydase serum levels in osteoarthritis patients after hip or knee replacement. *Annals of the Rheumatic Diseases*, 67(2), 168–174. https://doi.org/10.1136/ard.2007.073452
- Deberg, M., Labasse, A., Christgau, S., Cloos, P., Bang Henriksen, D., Chapelle, J.-P., Zegels, B., Reginster, J.-Y., & Henrotin, Y. (2005). New serum biochemical markers (Coll 2-1 and Coll 2-1 NO2) for studying oxidative-related type II collagen network degradation in patients with osteoarthritis and rheumatoid arthritis. *Osteoarthritis and Cartilage*, 13(3), 258–265. https://doi.org/10.1016/j.joca.2004. 12.002
- Dell'Isola, A., Allan, R., Smith, S. L., Marreiros, S. S. P., & Steultjens, M. (2016). Identification of clinical phenotypes in knee osteoarthritis: A systematic review of the literature. *BMC Musculoskeletal Disorders*, 17(1), 1–12. https://doi.org/10.1186/ s12891-016-1286-2
- Deveza, L. A., & Loeser, R. F. (2018). Is osteoarthritis one disease or a collection of many? *Rheumatology (Oxford, England)*, *57*(Suppl 4), iv34–iv42. https://doi.org/10.1093/rheumatology/kex417
- Deyle, G. D., Allen, C. S., Allison, S. C., Gill, N. W., Hando, B. R., Petersen, E. J., Dusenberry, D. I., & Rhon, D. I. (2020). Physical Therapy versus Glucocorticoid Injection for Osteoarthritis of the Knee. *The New England Journal of Medicine*, 382(15), 1420–1429. https://doi.org/10.1056/NEJMoa1905877
- Dillon, C. F., Rasch, E. K., Gu, Q., & Hirsch, R. (2006). Prevalence of Knee Osteoarthritis in the United States: Arthritis Data from the Third National Health and Nutrition Examination Survey 1991–94. *The Journal of Rheumatology*, 33:11, 9.
- Donnelly, E., Chen, D. X., Boskey, A. L., Baker, S. P., & Meulen, M. C. H. van der. (2010). Contribution of mineral to bone structural behavior and tissue mechanical properties. *Calcified Tissue International*, 87(5), 450–460. https://doi.org/10.1007/ s00223-010-9404-x
- Dougados, M., Ayral, X., Listrat, V., Gueguen, A., Bahuaud, J., Beaufils, P., Beguin, J.
 A., Bonvarlet, J. P., Boyer, T., Coudane, H., Delaunay, C., Dorfmann, H., Dubos, J.
 P., Frank, A., Kempf, J. F., Locker, B., Prudhon, J. L., & Thiery, J. (1994). The SFA system for assessing articular cartilage lesions at arthroscopy of the knee.

- *Arthroscopy: The Journal of Arthroscopic & Related Surgery*, 10(1), 69–77. https://doi.org/10.1016/S0749-8063(05)80295-6
- Driban, J. B., Eaton, C. B., Lo, G. H., Ward, R. J., Lu, B., & McAlindon, T. E. (2014). Association of knee injuries with accelerated knee osteoarthritis progression: Data from the Osteoarthritis Initiative. *Arthritis Care & Research*, 66(11), 1673–1679. https://doi.org/10.1002/acr.22359
- Driban, J. B., Price, L. L., Lynch, J., Nevitt, M., Lo, G. H., Eaton, C. B., & McAlindon, T. E. (2016). Defining and evaluating a novel outcome measure representing end-stage knee osteoarthritis: Data from the Osteoarthritis Initiative. *Clinical Rheumatology*, 35(10), 2523–2530. https://doi.org/10.1007/s10067-016-3299-5
- Driban, J. B., Tassinari, A., Lo, G. H., Price, L. L., Schneider, E., Lynch, J. A., Eaton, C. B., & McAlindon, T. E. (2012). Bone Marrow Lesions are Associated with Altered Trabecular Morphometry. Osteoarthritis and Cartilage / OARS, Osteoarthritis Research Society, 20(12), 1519–1526. https://doi.org/10.1016/j.joca. 2012.08.013
- Duncan, R., Peat, G., Thomas, E., Hay, E., McCall, I., & Croft, P. (2007). Symptoms and radiographic osteoarthritis: Not as discordant as they are made out to be? *Annals of the Rheumatic Diseases*, 66(1), 86. https://doi.org/10.1136/ard.2006.052548
- Eastell, R., & Szulc, P. (2017). Use of bone turnover markers in postmenopausal osteoporosis. *The Lancet Diabetes & Endocrinology*, 5(11), 908–923. https://doi.org/10.1016/S2213-8587(17)30184-5
- Elsaid, K. A., & Chichester, C. O. (2006). Review: Collagen markers in early arthritic diseases. *Clinica Chimica Acta*, 365(1), 68–77. https://doi.org/10.1016/j.cca.2005.09.020
- Emery, C. A., Whittaker, J. L., Mahmoudian, A., Lohmander, L. S., Roos, E. M., Bennell, K. L., Toomey, C. M., Reimer, R. A., Thompson, D., Ronsky, J. L., Kuntze, G., Lloyd, D. G., Andriacchi, T., Englund, M., Kraus, V. B., Losina, E., Bierma-Zeinstra, S., Runhaar, J., Peat, G., ... Arden, N. K. (2019). Establishing outcome measures in early knee osteoarthritis. *Nature Reviews Rheumatology*, 15(7), 438–448. https://doi.org/10.1038/s41584-019-0237-3
- Endres, E., van Drongelen, S., Meurer, A., Zaucke, F., & Stief, F. (2020). Effect of total joint replacement in hip osteoarthritis on serum COMP and its correlation with mechanical-functional parameters of gait analysis. *Osteoarthritis and Cartilage Open*, 2(1), 100034. https://doi.org/10.1016/j.ocarto.2020.100034
- Ene, R., Sinescu, R. D., Ene, P., Cîrstoiu, M. M., & Cîrstoiu, F. C. (2015). Synovial inflammation in patients with different stages of knee osteoarthritis. *Romanian Journal of Morphology and Embryology = Revue Roumaine De Morphologie Et Embryologie*, 56(1), 169–173.
- Englund, M. (2010). The role of biomechanics in the initiation and progression of OA of the knee. *Best Practice & Research Clinical Rheumatology*, 24(1), 39–46. https://doi.org/10.1016/j.berh.2009.08.008
- Englund, M., Guermazi, A., & Lohmander, L. S. (2009). The Meniscus in Knee Osteoarthritis. *Rheumatic Disease Clinics of North America*, 35(3), 579–590. https://doi.org/10.1016/j.rdc.2009.08.004
- Englund, M., Roemer, F. W., Hayashi, D., Crema, M. D., & Guermazi, A. (2012). Meniscus pathology, osteoarthritis and the treatment controversy. *Nature Reviews Rheumatology*, 8(7), 412–419. https://doi.org/10.1038/nrrheum.2012.69
- Eymard, F., Pigenet, A., Citadelle, D., Tordjman, J., Foucher, L., Rose, C., Flouzat Lachaniette, C.-H., Rouault, C., Clément, K., Berenbaum, F., Chevalier, X., &

- Houard, X. (2017). Knee and hip intra-articular adipose tissues (IAATs) compared with autologous subcutaneous adipose tissue: A specific phenotype for a central player in osteoarthritis. *Annals of the Rheumatic Diseases*, 76(6), 1142–1148. https://doi.org/10.1136/annrheumdis-2016-210478
- Fan, Z., Bau, B., Yang, H., Soeder, S., & Aigner, T. (2005). Freshly isolated osteo-arthritic chondrocytes are catabolically more active than normal chondrocytes, but less responsive to catabolic stimulation with interleukin-1beta. *Arthritis and Rheumatism*, 52(1), 136–143. https://doi.org/10.1002/art.20725
- Favero, M., Ramonda, R., Goldring, M. B., Goldring, S. R., & Punzi, L. (2015). Early knee osteoarthritis: Figure 1. *RMD Open*, *1*(Suppl 1), e000062. https://doi.org/10. 1136/rmdopen-2015-000062
- FDA-NIH Biomarker Working Group. (2016). *BEST (Biomarkers, EndpointS, and other Tools) Resource*. Food and Drug Administration (US). http://www.ncbi.nlm.nih.gov/books/NBK326791/
- Felson, D., Niu, J., Sack, B., Aliabadi, P., McCullough, C., & Nevitt, M. C. (2013). Progression of osteoarthritis as a state of inertia. *Annals of the Rheumatic Diseases*, 72(6), 924–929. https://doi.org/10.1136/annrheumdis-2012-201575
- Felson, D. T., Naimark, A., Anderson, J., Kazis, L., Castelli, W., & Meenan, R. F. (1987). The prevalence of knee osteoarthritis in the elderly. The Framingham Osteoarthritis Study. *Arthritis & Rheumatism*, 30(8), 914–918. https://doi.org/10.1002/art. 1780300811
- Felson, D. T., Niu, J., Guermazi, A., Sack, B., & Aliabadi, P. (2011). Defining radiographic incidence and progression of knee osteoarthritis: Suggested modifications of the Kellgren and Lawrence scale. *Annals of the Rheumatic Diseases*, 70(11), 1884–1886. https://doi.org/10.1136/ard.2011.155119
- Felson, D. T., Zhang, Y., Hannan, M. T., Naimark, A., Weissman, B., Aliabadi, P., & Levy, D. (1997). Risk factors for incident radiographic knee osteoarthritis in the elderly. The Framingham study. *Arthritis & Rheumatism*, 40(4), 728–733. https://doi.org/10.1002/art.1780400420
- Flandry, F., & Hommel, G. (2011). Normal anatomy and biomechanics of the knee. *Sports Medicine and Arthroscopy Review*, 19(2), 82–92. https://doi.org/10.1097/JSA.0b013e318210c0aa
- Franceschi, C., & Bonafè, M. (2003). Centenarians as a model for healthy aging. *Biochemical Society Transactions*, 31(2), 457–461. https://doi.org/10.1042/bst0310457
- Franceschi, C., Capri, M., Monti, D., Giunta, S., Olivieri, F., Sevini, F., Panourgia, M. P., Invidia, L., Celani, L., Scurti, M., Cevenini, E., Castellani, G. C., & Salvioli, S. (2007). Inflammaging and anti-inflammaging: A systemic perspective on aging and longevity emerged from studies in humans. *Mechanisms of Ageing and Development*, 128(1), 92–105. https://doi.org/10.1016/j.mad.2006.11.016
- Frost, H. M. (2003). Perspective: Genetic and hormonal roles in bone disorders: insights of an updated bone physiology. *Journal of Musculoskeletal & Neuronal Interactions*, 3(2), 118–135.
- Funck-Brentano, T., & Cohen-Solal, M. (2011). Crosstalk between cartilage and bone: When bone cytokines matter. *Cytokine & Growth Factor Reviews*, 22(2), 91–97. https://doi.org/10.1016/j.cytogfr.2011.04.003
- Funck-Brentano, T., & Cohen-Solal, M. (2015). Subchondral bone and osteoarthritis. *Current Opinion in Rheumatology*, 27(4), 420–426. https://doi.org/10.1097/BOR. 000000000000181

- Gale, D. R., Chaisson, C. E., Totterman, S. M., Schwartz, R. K., Gale, M. E., & Felson, D. (1999). Meniscal subluxation: Association with osteoarthritis and joint space narrowing. *Osteoarthritis and Cartilage*, 7(6), 526–532. https://doi.org/10.1053/joca.1999.0256
- Gandhi, R., Takahashi, M., Rizek, R., Dessouki, O., & Mahomed, N. N. (2012). Obesity-related adipokines and shoulder osteoarthritis. *The Journal of Rheumatology*, *39*(10), 2046–2048. https://doi.org/10.3899/jrheum.111339
- Garnero, P., Sornay-Rendu, E., & Chapurlat, R. (2020). The cartilage degradation marker, urinary CTX-II, is associated with the risk of incident total joint replacement in postmenopausal women. A 18 year evaluation of the OFELY prospective cohort. *Osteoarthritis and Cartilage*, 28(4), 468–474. https://doi.org/10.1016/j.joca.2019.12.012
- Gelse, K., Ekici, A. B., Cipa, F., Swoboda, B., Carl, H. D., Olk, A., Hennig, F. F., & Klinger, P. (2012). Molecular differentiation between osteophytic and articular cartilage clues for a transient and permanent chondrocyte phenotype. *Osteoarthritis and Cartilage*, 20(2), 162–171. https://doi.org/10.1016/j.joca.2011.12.004
- Gelse, K., Söder, S., Eger, W., Diemtar, T., & Aigner, T. (2003). Osteophyte development—Molecular characterization of differentiation stages. *Osteoarthritis and Cartilage*, 11(2), 141–148. https://doi.org/10.1053/joca.2002.0873
- Gilbertson, E. M. (1975). Development of periarticular osteophytes in experimentally induced osteoarthritis in the dog. A study using microradiographic, microangiographic, and fluorescent bone-labelling techniques. *Annals of the Rheumatic Diseases*, 34(1), 12–25. https://doi.org/10.1136/ard.34.1.12
- Glassman, S. D., Copay, A. G., Berven, S. H., Polly, D. W., Subach, B. R., & Carreon, L. Y. (2008). Defining substantial clinical benefit following lumbar spine arthrodesis. *The Journal of Bone and Joint Surgery. American Volume*, *90*(9), 1839–1847. https://doi.org/10.2106/JBJS.G.01095
- Gly-Jones, S., Palmer, AJR, Agricola, R., Price, A., Vincent TL, Weinans H, & Carr AJ. (2015). Osteoarthritis. *The Lancet*, *386*(9991), 376–387. https://doi.org/10.1016/S0140-6736(14)60802-3
- Goldring, M. B., Birkhead, J., Sandell, L. J., Kimura, T., & Krane, S. M. (1988). Interleukin 1 suppresses expression of cartilage-specific types II and IX collagens and increases types I and III collagens in human chondrocytes. *Journal of Clinical Investigation*, 82(6), 2026–2037. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC442785/
- Goldring, M. B., & Goldring, S. R. (2007). Osteoarthritis. *Journal of Cellular Physiology*, 213(3), 626–634. https://doi.org/10.1002/jcp.21258
- Goldring, M. B., & Goldring, S. R. (2010). Articular cartilage and subchondral bone in the pathogenesis of osteoarthritis. *Annals of the New York Academy of Sciences*, 1192(1), 230–237. https://doi.org/10.1111/j.1749-6632.2009.05240.x
- Goldring, M. B., & Marcu, K. B. (2009). Cartilage homeostasis in health and rheumatic diseases. *Arthritis Research & Therapy*, 11(3), 224. https://doi.org/10.1186/ar2592
- Goldring, S. R. (2008). The Role of Bone in Osteoarthritis Pathogenesis. *Rheumatic Disease Clinics of North America*, 34(3), 561–571. https://doi.org/10.1016/j.rdc. 2008.07.001
- Goldring, S. R. (2012). Alterations in periarticular bone and cross talk between sub-chondral bone and articular cartilage in osteoarthritis. *Therapeutic Advances in Musculoskeletal Disease*, 4(4), 249–258. https://doi.org/10.1177/1759720X12437353

- Goldring, S. R., & Goldring, M. B. (2016). Changes in the osteochondral unit during osteoarthritis: Structure, function and cartilage-bone crosstalk. *Nature Reviews*. *Rheumatology*, 12(11), 632–644. https://doi.org/10.1038/nrrheum.2016.148
- Grainger, A. J., Rhodes, L. A., Keenan, A.-M., Emery, P., & Conaghan, P. G. (2007). Quantifying peri-meniscal synovitis and its relationship to meniscal pathology in osteoarthritis of the knee. *European Radiology*, *17*(1), 119–124. https://doi.org/10. 1007/s00330-006-0282-6
- Guermazi, A., Hayashi, D., Roemer, F., Felson, D. T., Wang, K., Lynch, J., Amin, S., Torner, J., Lewis, C. E., & Nevitt, M. C. (2015). Severe radiographic knee osteoarthritis does Kellgren and Lawrence grade 4 represent end stage disease? The MOST study. *Osteoarthritis and Cartilage*, 23(9), 1499–1505. https://doi.org/10.1016/j.joca.2015.04.018
- Guermazi, A., Niu, J., Hayashi, D., Roemer, F. W., Englund, M., Neogi, T., Aliabadi, P., McLennan, C. E., & Felson, D. T. (2012). Prevalence of abnormalities in knees detected by MRI in adults without knee osteoarthritis: Population based observational study (Framingham Osteoarthritis Study). *BMJ (Clinical Research Ed.)*, 345, e5339. https://doi.org/10.1136/bmj.e5339
- Hamada, D., Maynard, R., Schott, E., Drinkwater, C. J., Ketz, J. P., Kates, S. L., Jonason, J. H., Hilton, M. J., Zuscik, M. J., & Mooney, R. A. (2016). Suppressive Effects of Insulin on Tumor Necrosis Factor-Dependent Early Osteoarthritic Changes Associated With Obesity and Type 2 Diabetes Mellitus. *Arthritis & Rheumatology (Hoboken, N.J.)*, 68(6), 1392–1402. https://doi.org/10.1002/art.39561
- Hamilton, J. L., Nagao, M., Levine, B. R., Chen, D., Olsen, B. R., & Im, H.-J. (2016). Targeting VEGF and its Receptors for the Treatment of Osteoarthritis and Associated Pain. Journal of Bone and Mineral Research: The Official Journal of the American Society for Bone and Mineral Research, 31(5), 911–924. https://doi.org/ 10.1002/jbmr.2828
- Hao, H. Q., Zhang, J. F., He, Q. Q., & Wang, Z. (2019). Cartilage oligomeric matrix protein, C-terminal cross-linking telopeptide of type II collagen, and matrix metalloproteinase-3 as biomarkers for knee and hip osteoarthritis (OA) diagnosis: A systematic review and meta-analysis. *Osteoarthritis and Cartilage*, 27(5), 726–736. https://doi.org/10.1016/j.joca.2018.10.009
- Haraden, C. A., Huebner, J. L., Hsueh, M.-F., Li, Y.-J., & Kraus, V. B. (2019). Synovial fluid biomarkers associated with osteoarthritis severity reflect macrophage and neutrophil related inflammation. *Arthritis Research & Therapy*, 21(1), 146. https://doi.org/10.1186/s13075-019-1923-x
- Harasymowicz, N. S., Clement, N. D., Azfer, A., Burnett, R., Salter, D. M., & Simpson, A. H. W. R. (2017). Regional Differences Between Perisynovial and Infrapatellar Adipose Tissue Depots and Their Response to Class II and Class III Obesity in Patients With Osteoarthritis. Arthritis & Rheumatology (Hoboken, N.J.), 69(7), 1396–1406. https://doi.org/10.1002/art.40102
- Hart, D. J., & Spector, T. D. (2003). Kellgren & Lawrence grade 1 osteophytes in the knee Doubtful or definite? *Osteoarthritis and Cartilage*, 11(2), 149–150. https://doi.org/10.1053/joca.2002.0853
- Haseeb, A., & Haqqi, T. M. (2013). Immunopathogenesis of osteoarthritis. *Clinical Immunology*, *146*(3), 185–196. https://doi.org/10.1016/j.clim.2012.12.011
- Hasegawa, A., Otsuki, S., Pauli, C., Miyaki, S., Patil, S., Steklov, N., Kinoshita, M., Koziol, J., D'Lima, D. D., & Lotz, M. K. (2012). Anterior Cruciate Ligament

- Changes in Human Joint in Aging and Osteoarthritis. Arthritis and Rheumatism, 64(3), 696–704. https://doi.org/10.1002/art.33417
- Hawker, G. A. (2019). Osteoarthritis is a serious disease. *Clinical and Experimental Rheumatology*, 37 Suppl 120(5), 3–6.
- Hayashi, D., Roemer, F. W., & Guermazi, A. (2018). Imaging of osteoarthritis—Recent research developments and future perspective. *The British Journal of Radiology*, 91(1085). https://doi.org/10.1259/bjr.20170349
- He, Y., Jensen, K. E., Siebuhr, A. S., Karsdal, M. A., Larkin, J., & Bay-Jensen, A. C. (2021). Development of a highly sensitive chemiluminescence immunoassay for quantification of aggrecanase-generated ARGS aggrecan fragments in serum. Osteoarthritis and Cartilage Open, 3(2), 100162. https://doi.org/10.1016/j.ocarto. 2021.100162
- Henrotin, Y. (2021). Osteoarthritis in year 2021: Biochemical markers. *Osteoarthritis and Cartilage*, 0(0). https://doi.org/10.1016/j.joca.2021.11.001
- Henrotin, Y., Addison, S., Kraus, V., & Deberg, M. (2007). Type II collagen markers in osteoarthritis: What do they indicate?: *Current Opinion in Rheumatology*, 19(5), 444–450. https://doi.org/10.1097/BOR.0b013e32829fb3b5
- Hensor, E. M. A., Dube, B., Kingsbury, S. R., Tennant, A., & Conaghan, P. G. (2015). Toward a Clinical Definition of Early Osteoarthritis: Onset of Patient-Reported Knee Pain Begins on Stairs. Data From the Osteoarthritis Initiative. *Arthritis Care & Research*, 67(1), 40–47. https://doi.org/10.1002/acr.22418
- Hinman, R. S., & Crossley, K. M. (2007). Patellofemoral joint osteoarthritis: An important subgroup of knee osteoarthritis. *Rheumatology*, 46(7), 1057–1062. https://doi.org/10.1093/rheumatology/kem114
- Hoch, J. M., Mattacola, C. G., McKeon, J. M. M., Howard, J. S., & Lattermann, C. (2011). Serum cartilage oligomeric matrix protein (sCOMP) is elevated in patients with knee osteoarthritis: A systematic review and meta-analysis. *Osteoarthritis and Cartilage / OARS, Osteoarthritis Research Society*, 19(12), 1396–1404. https://doi.org/10.1016/j.joca.2011.09.005
- Houard, X., Goldring, M. B., & Berenbaum, F. (2013). Homeostatic Mechanisms in Articular Cartilage and Role of Inflammation in Osteoarthritis. *Current Rheumatology Reports*, 15(11), 375. https://doi.org/10.1007/s11926-013-0375-6
- Hu, W., Chen, Y., Dou, C., & Dong, S. (2021). Microenvironment in subchondral bone: Predominant regulator for the treatment of osteoarthritis. *Annals of the Rheumatic Diseases*, 80(4), 413–422. https://doi.org/10.1136/annrheumdis-2020-218089
- Huang, M., Zhao, J., Huang, Y., Dai, L., & Zhang, X. (2017). Meta-analysis of urinary C-terminal telopeptide of type II collagen as a biomarker in osteoarthritis diagnosis. *Journal of Orthopaedic Translation*, 13, 50–57. https://doi.org/10.1016/j. jot.2017.06.005
- Huebner, J. L., Bay-Jensen, A. C., Huffman, K. M., He, Y., Leeming, D. J., McDaniel, G. E., Karsdal, M. A., & Kraus, V. B. (2014). ALPHA-CTX is associated with subchondral bone turnover and predicts progression of joint space narrowing and osteophytes in osteoarthritis. *Arthritis & Rheumatology (Hoboken, N.J.)*, 66(9), 2440–2449. https://doi.org/10.1002/art.38739
- Hunter, D. J., & Bierma-Zeinstra, S. (2019). Osteoarthritis. *The Lancet*, *393*(10182), 1745–1759. https://doi.org/10.1016/S0140-6736(19)30417-9
- Hunter, D. J., March, L., & Chew, M. (2020). Osteoarthritis in 2020 and beyond: A Lancet Commission. *The Lancet*, 396(10264), 1711–1712. https://doi.org/10.1016/S0140-6736(20)32230-3

- Hunter, D. J., Zhang, Y. Q., Tu, X., LaValley, M., Niu, J. B., Amin, S., Guermazi, A., Genant, H., Gale, D., & Felson, D. T. (2006). Change in joint space width: Hyaline articular cartilage loss or alteration in meniscus? *Arthritis & Rheumatism*, 54(8), 2488–2495. https://doi.org/10.1002/art.22016
- Hutton, C. W. (1987). Generalised osteoarthritis: An evolutionary problem? *Lancet (London, England)*, 1(8548), 1463–1465. https://doi.org/10.1016/s0140-6736(87) 92209-4
- Ioan-Facsinay, A., & Kloppenburg, M. (2013). An emerging player in knee osteo-arthritis: The infrapatellar fat pad. *Arthritis Research & Therapy*, 15(6), 225. https://doi.org/10.1186/ar4422
- Jayasuriya, C. T., Hu, N., Li, J., Lemme, N., Terek, R., Ehrlich, M. G., & Chen, Q. (2018). Molecular characterization of mesenchymal stem cells in human osteo-arthritis cartilage reveals contribution to the OA phenotype. *Scientific Reports*, 8(1), 7044. https://doi.org/10.1038/s41598-018-25395-8
- Jin, X., Beguerie, J. R., Zhang, W., Blizzard, L., Otahal, P., Jones, G., & Ding, C. (2015). Circulating C reactive protein in osteoarthritis: A systematic review and meta-analysis. *Annals of the Rheumatic Diseases*, 74(4), 703–710. https://doi.org/10.1136/annrheumdis-2013-204494
- Jin, Z., Wang, D., Zhang, H., Liang, J., Feng, X., Zhao, J., & Sun, L. (2020). Incidence trend of five common musculoskeletal disorders from 1990 to 2017 at the global, regional and national level: Results from the global burden of disease study 2017. *Annals of the Rheumatic Diseases*, 79(8), 1014–1022. https://doi.org/10.1136/ annrheumdis-2020-217050
- Johnson, V. L., & Hunter, D. J. (2014). The epidemiology of osteoarthritis. *Best Practice & Research Clinical Rheumatology*, 28(1), 5–15. https://doi.org/10.1016/j.berh. 2014.01.004
- Jones, A. C., Ledingham, J., McAlindon, T., Regan, M., Hart, D., MacMillan, P. J., & Doherty, M. (1993). Radiographic assessment of patellofemoral osteoarthritis. *Annals of the Rheumatic Diseases*, 52(9), 655–658. https://doi.org/10.1136/ard. 52.9.655
- Jones, G., Ding, C., Scott, F., Glisson, M., & Cicuttini, F. (2004). Early radiographic osteoarthritis is associated with substantial changes in cartilage volume and tibial bone surface area in both males and females11Sources of support: National Health and Medical Research Council of Australia, Masonic Centenary Medical Research Foundation. *Osteoarthritis and Cartilage*, 12(2), 169–174. https://doi.org/10.1016/j.joca.2003.08.010
- Jordan, J. M., Helmick, C. G., Renner, J. B., Luta, G., Dragomir, A. D., Woodard, J., Fang, F., Schwartz, T. A., Abbate, L. M., Callahan, L. F., Kalsbeek, W. D., & Hochberg, M. C. (2007). Prevalence of Knee Symptoms and Radiographic and Symptomatic Knee Osteoarthritis in African Americans and Caucasians: The Johnston County Osteoarthritis Project. *The Journal of Rheumatology*, 34:1, 9.
- Kalaitzoglou, E., Lopes, E. B. P., Fu, Y., Herron, J. C., Flaming, J. M., Donovan, E. L., Hu, Y., Filiberti, A., Griffin, T. M., & Humphrey, M. B. (2019). TLR4 Promotes and DAP12 Limits Obesity-Induced Osteoarthritis in Aged Female Mice. *JBMR Plus*, 3(4), e10079. https://doi.org/10.1002/jbm4.10079
- Kambic, H. E., & McDevitt, C. A. (2005). Spatial organization of types I and II collagen in the canine meniscus. *Journal of Orthopaedic Research*, 23(1), 142–149. https://doi.org/10.1016/j.orthres.2004.06.016

- Kapoor, M., Martel-Pelletier, J., Lajeunesse, D., Pelletier, J.-P., & Fahmi, H. (2011). Role of proinflammatory cytokines in the pathophysiology of osteoarthritis. *Nature Reviews Rheumatology*, 7(1), 33–42. https://doi.org/10.1038/nrrheum.2010.196
- Karsdal, M. A., Byrjalsen, I., Alexandersen, P., Bihlet, A., Andersen, J. R., Riis, B. J., Bay-Jensen, A. C., & Christiansen, C. (2015). Treatment of symptomatic knee osteoarthritis with oral salmon calcitonin: Results from two phase 3 trials. Osteoarthritis and Cartilage, 23(4), 532–543. https://doi.org/10.1016/j.joca.2014. 12.019
- Karsdal, M. A., Christiansen, C., Ladel, C., Henriksen, K., Kraus, V. B., & Bay-Jensen, A. C. (2014). Osteoarthritis a case for personalized health care? *Osteoarthritis and Cartilage*, 22(1), 7–16. https://doi.org/10.1016/j.joca.2013.10.018
- Karsdal, M. A., Henriksen, K., & Bay-Jensen, A. C. (2019). Biochemical markers in osteoarthritis with lessons learned from osteoporosis. *Clin Exp Rheumatol*. https://www.clinexprheumatol.org/abstract.asp?a=14754
- Karsdal, M. A., Henriksen, K., Leeming, D. J., Woodworth, T., Vassiliadis, E., & Bay-Jensen, A.-C. (2010). Novel combinations of Post-Translational Modification (PTM) neo-epitopes provide tissue-specific biochemical markers—Are they the cause or the consequence of the disease? *Clinical Biochemistry*, 43(10–11), 793–804. https://doi.org/10.1016/j.clinbiochem.2010.03.015
- Katsuragawa, Y., Saitoh, K., Tanaka, N., Wake, M., Ikeda, Y., Furukawa, H., Tohma, S., Sawabe, M., Ishiyama, M., Yagishita, S., Suzuki, R., Mitomi, H., & Fukui, N. (2010). Changes of human menisci in osteoarthritic knee joints. *Osteoarthritis and Cartilage*, 18(9), 1133–1143. https://doi.org/10.1016/j.joca.2010.05.017
- Katsuragi, J., Sasho, T., Yamaguchi, S., Sato, Y., Watanabe, A., Akagi, R., Muramatsu, Y., Mukoyama, S., Akatsu, Y., Fukawa, T., Endo, J., Hoshi, H., Yamamoto, Y., Sasaki, T., & Takahashi, K. (2015). Hidden osteophyte formation on plain X-ray is the predictive factor for development of knee osteoarthritis after 48 months data from the Osteoarthritis Initiative. *Osteoarthritis and Cartilage*, 23(3), 383–390. https://doi.org/10.1016/j.joca.2014.11.026
- Katz, J. N., Arant, K. R., & Loeser, R. F. (2021). Diagnosis and Treatment of Hip and Knee Osteoarthritis: A Review. JAMA, 325(6), 568–578. https://doi.org/10.1001/ jama.2020.22171
- Kellgren, J. H., & Lawrence, J. S. (1957). Radiological Assessment of Osteo-Arthrosis. Annals of the Rheumatic Diseases, 16(4), 494–502. https://doi.org/10.1136/ard. 16.4.494
- Kerkhof, H. J. M., Meulenbelt, I., Akune, T., Arden, N. K., Aromaa, A., Bierma-Zeinstra, S. M. A., Carr, A., Cooper, C., Dai, J., Doherty, M., Doherty, S. A., Felson, D., Gonzalez, A., Gordon, A., Harilainen, A., Hart, D. J., Hauksson, V. B., Heliovaara, M., Hofman, A., ... van Meurs, J. B. J. (2011). Recommendations for standardization and phenotype definitions in genetic studies of osteoarthritis: The TREAT-OA consortium. *Osteoarthritis and Cartilage*, 19(3), 254–264. https://doi.org/10.1016/j.joca.2010.10.027
- Kerna, I., Kisand, K., Laitinen, P., Tamm, A. E., Kumm, J., Lintrop, M., & Tamm, A. O. (2012). Association of ADAM12-S protein with radiographic features of knee osteoarthritis and bone and cartilage markers. *Rheumatology International*, 32(2), 519–523. https://doi.org/10.1007/s00296-010-1717-6
- Kerna, I., Kisand, K., Laitinen, P., Tamm, A., & Tamm, A. (2010). 386 ASSOCIATION OF METALLOPEPTIDASE DOMAIN 12 (ADAM12) GENE POLYMORPHISMS AND ADAM12 PROTEIN WITH THE DEVELOPMENT OF

- KNEE OSTEOARTHRITIS. Osteoarthritis and Cartilage, 18, S170. https://doi.org/10.1016/S1063-4584(10)60413-X
- Kerna, I., Kisand, K., Tamm, A. E., Kumm, J., & Tamm, A. O. (2013). Two Single-Nucleotide Polymorphisms in ADAM12 Gene Are Associated with Early and Late Radiographic Knee Osteoarthritis in Estonian Population. *Arthritis*, 2013, 878126–878126. https://doi.org/10.1155/2013/878126
- Kerna, I., Kisand, K., Tamm, A. E., Lintrop, M., Veske, K., & Tamm, A. O. (2009). Missense single nucleotide polymorphism of the ADAM12 gene is associated with radiographic knee osteoarthritis in middle-aged Estonian cohort. *Osteoarthritis and Cartilage*, 17(8), 1093–1098. https://doi.org/10.1016/j.joca.2009.02.006
- Kijowski, R., Blankenbaker, D. G., Stanton, P. T., Fine, J. P., & De Smet, A. A. (2006). Radiographic findings of osteoarthritis versus arthroscopic findings of articular cartilage degeneration in the tibiofemoral joint. *Radiology*, *239*(3), 818–824. Scopus. https://doi.org/10.1148/radiol.2393050584
- Kisand, K., Tamm, A. E., Lintrop, M., & Tamm, A. O. (2018). New insights into the natural course of knee osteoarthritis: Early regulation of cytokines and growth factors, with emphasis on sex-dependent angiogenesis and tissue remodeling. A pilot study. *Osteoarthritis and Cartilage*, 26(8), 1045–1054. https://doi.org/10.1016/j.joca.2018.05.009
- Kobayashi, M., Squires, G. R., Mousa, A., Tanzer, M., Zukor, D. J., Antoniou, J., Feige, U., & Poole, A. R. (2005). Role of interleukin-1 and tumor necrosis factor alpha in matrix degradation of human osteoarthritic cartilage. *Arthritis and Rheumatism*, 52(1), 128–135. https://doi.org/10.1002/art.20776
- Kobayashi, S., Pappas, E., Fransen, M., Refshauge, K., & Simic, M. (2016). The prevalence of patellofemoral osteoarthritis: A systematic review and meta-analysis. *Osteoarthritis and Cartilage*, 24(10), 1697–1707. https://doi.org/10.1016/j.joca. 2016.05.011
- Korhonen, R. K., Julkunen, P., Wilson, W., & Herzog, W. (2008). Importance of collagen orientation and depth-dependent fixed charge densities of cartilage on mechanical behavior of chondrocytes. *Journal of Biomechanical Engineering*, 130(2), 021003. https://doi.org/10.1115/1.2898725
- Krasnokutsky, S., Belitskaya-Lévy, I., Bencardino, J., Samuels, J., Attur, M., Regatte, R., Rosenthal, P., Greenberg, J., Schweitzer, M., Abramson, S. B., & Rybak, L. (2011). Quantitative MRI Evidence of Synovial Proliferation is Associated with Radiographic Severity of Knee Osteoarthritis. *Arthritis and Rheumatism*, 63(10), 2983–2991. https://doi.org/10.1002/art.30471
- Kraus, V. B., Blanco, F. J., Englund, M., Karsdal, M. A., & Lohmander, L. S. (2015).
 Call for Standardized Definitions of Osteoarthritis and Risk Stratification for Clinical Trials and Clinical Use. Osteoarthritis and Cartilage / OARS, Osteoarthritis Research Society, 23(8), 1233–1241. https://doi.org/10.1016/j.joca.2015.03.036
- Kraus, V. B., Burnett, B., Coindreau, J., Cottrell, S., Eyre, D., Gendreau, M., Gardiner, J., Garnero, P., Hardin, J., Henrotin, Y., Heinegård, D., Ko, A., Lohmander, S., Matthews, G., Menetski, J., Moskowitz, R., Persiani, S., Poole, R., Rousseau, J. C., & Todman, M. (2011). Application of Biomarkers in the Development of Drugs Intended for the Treatment of Osteoarthritis. Osteoarthritis and Cartilage / OARS, Osteoarthritis Research Society, 19(5), 515–542. https://doi.org/10.1016/j.joca. 2010.08.019
- Kraus, V. B., Collins, J. E., Hargrove, D., Losina, E., Nevitt, M., Katz, J. N., Wang, S. X., Sandell, L. J., Hoffmann, S. C., & Hunter, D. J. (2017). Predictive validity of

- biochemical biomarkers in knee osteoarthritis: Data from the FNIH OA Biomarkers Consortium. *Annals of the Rheumatic Diseases*, 76(1), 186–195. https://doi.org/10.1136/annrheumdis-2016-209252
- Kraus, V. B., Hargrove, D. E., Hunter, D. J., Renner, J. B., & Jordan, J. M. (2017). Establishment of reference intervals for osteoarthritis-related soluble biomarkers: The FNIH/OARSI OA Biomarkers Consortium. *Annals of the Rheumatic Diseases*, 76(1), 179–185. https://doi.org/10.1136/annrheumdis-2016-209253
- Kraus, V. B., Jordan, J. M., Doherty, M., Wilson, A. G., Moskowitz, R., Hochberg, M., Loeser, R., Hooper, M., Renner, J. B., Crane, M. M., Hastie, P., Sundseth, S., & Atif, U. (2007). The Genetics of Generalized Osteoarthritis (GOGO) study: Study design and evaluation of osteoarthritis phenotypes. *Osteoarthritis and Cartilage*, 15(2), 120–127. https://doi.org/10.1016/j.joca.2006.10.002
- Kraus, V. B., & Karsdal, M. A. (2021). Osteoarthritis: Current Molecular Biomarkers and the Way Forward. *Calcified Tissue International*, 109(3), 329–338. https://doi.org/10.1007/s00223-020-00701-7
- Kraus, V. B., Kepler, T. B., Stabler, T., Renner, J., & Jordan, J. (2010). First qualification study of serum biomarkers as indicators of total body burden of osteoarthritis. *PloS One*, *5*(3), e9739. https://doi.org/10.1371/journal.pone.0009739
- Kumavat, R., Kumar, V., Malhotra, R., Pandit, H., Jones, E., Ponchel, F., & Biswas, S. (2021). Biomarkers of Joint Damage in Osteoarthritis: Current Status and Future Directions. *Mediators of Inflammation*, 2021, e5574582. https://doi.org/10.1155/2021/5574582
- Kumm, J., Ivaska, K., Rohtla, K., Vaananen, K., & Tamm, A. (2008). Urinary osteocalcin and other markers of bone metabolism: The effect of risedronate therapy. SCANDINAVIAN JOURNAL OF CLINICAL AND LABORATORY INVESTIGATION, 6, 459. http://ezproxy.utlib.ut.ee/login?url=http://search.ebscohost.com/login.aspx?direct=true&db=edsbl&AN=RN237695081&site=eds-live
- Kumm, J., Tamm, A., Lintrop, M., Sondergaard, B., & Tamm, A. (2009). Ultrasonographic findings and biomarkers of bone and cartilage in subjects with early knee OA. *Bone*, 44, S281–S282. https://doi.org/10.1016/j.bone.2009.03.502
- Kumm, J., Tamm, A., Lintrop, M., & Tamm, A. (2010). The predictive role of ultrasonographically detected synovitis in the radiographic progression of early knee osteoarthritis. *Bone*, 47, S76. https://doi.org/10.1016/j.bone.2010.04.148
- Kumm, J., Tamm, A., Lintrop, M., & Tamm, A. (2012). The prevalence and progression of radiographic knee osteoarthritis over 6 years in a population-based cohort of middle-aged subjects. *Rheumatology International*, 32(11), 3545–3550. https://doi.org/10.1007/s00296-011-2221-3
- Kumm, J., Tamm, A., Lintrop, M., & Tamm, A. (2013a). The prevalence and progression of radiographic knee osteoarthritis over 9 years in a population-based cohort of middle-aged subjects. *Osteoarthritis and Cartilage*, 21, S204. https://doi.org/10.1016/j.joca.2013.02.427
- Kumm, J., Tamm, A., Lintrop, M., & Tamm, A. (2013b). The value of cartilage biomarkers in progressive knee osteoarthritis: Cross-sectional and 6-year follow-up study in middle-aged subjects. *Rheumatology International*, *33*(4), 903–911. https://doi.org/10.1007/s00296-012-2463-8
- Kumm, J., Tamm, A., Lintrop, M., & Tamm, A. (2013c). Diagnostic and prognostic value of bone biomarkers in progressive knee osteoarthritis: A 6-year follow-up study in middle-aged subjects. *Osteoarthritis and Cartilage*, 21(6), 815–822. https://doi.org/10.1016/j.joca.2013.03.008

- Kumm, J., Tamm, A., Veske, K., Lintrop, M., & Tamm, A. (2006). Associations between cartilage oligomeric matrix protein and several articular tissues in early knee joint osteoarthritis. *Rheumatology*, 45(10), 1308–1309. https://doi.org/ 10.1093/rheumatology/kel271
- Lago, F., Dieguez, C., Gomezreino, J., & Gualillo, O. (2007). The emerging role of adipokines as mediators of inflammation and immune responses. *Cytokine & Growth Factor Reviews*, 18(3–4), 313–325. https://doi.org/10.1016/j.cytogfr.2007. 04.007
- Lanyon, P., O'Reilly, S., Jones, A., & Doherty, M. (1998). Radiographic assessment of symptomatic knee osteoarthritis in the community: Definitions and normal joint space. *Annals of the Rheumatic Diseases*, *57*(10), 595–601. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1752476/
- LaValley, M. P., McAlindon, T. E., Chaisson, C. E., Levy, D., & Felson, D. T. (2001). The validity of different definitions of radiographic worsening for longitudinal studies of knee osteoarthritis. *Journal of Clinical Epidemiology*, *54*(1), 30–39. https://doi.org/10.1016/S0895-4356(00)00273-0
- Lazzarini, N., Runhaar, J., Bay-Jensen, A. C., Thudium, C. S., Bierma-Zeinstra, S. M. A., Henrotin, Y., & Bacardit, J. (2017). A machine learning approach for the identification of new biomarkers for knee osteoarthritis development in overweight and obese women. *Osteoarthritis and Cartilage*, 25(12), 2014–2021. https://doi.org/10.1016/j.joca.2017.09.001
- Ledingham, J., Regan, M., Jones, A., & Doherty, M. (1993). Radiographic patterns and associations of osteoarthritis of the knee in patients referred to hospital. *Annals of the Rheumatic Diseases*, 52(7), 520–526. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1005091/
- Li, G., Yin, J., Gao, J., Cheng, T. S., Pavlos, N. J., Zhang, C., & Zheng, M. H. (2013). Subchondral bone in osteoarthritis: Insight into risk factors and microstructural changes. *Arthritis Research & Therapy*, 15(6), 223. https://doi.org/10.1186/ar4405
- Li, W. (Wendy), Nemirovskiy, O., Fountain, S., Rodney Mathews, W., & Szekely-Klepser, G. (2007). Clinical validation of an immunoaffinity LC–MS/MS assay for the quantification of a collagen type II neoepitope peptide: A biomarker of matrix metalloproteinase activity and osteoarthritis in human urine. *Analytical Biochemistry*, 369(1), 41–53. https://doi.org/10.1016/j.ab.2007.05.002
- Liem, Y., Judge, A., Kirwan, J., Ourradi, K., Li, Y., & Sharif, M. (2020a). Multivariable logistic and linear regression models for identification of clinically useful biomarkers for osteoarthritis. *Scientific Reports*, 10(1), 11328. https://doi.org/10.1038/ s41598-020-68077-0
- Liem, Y., Judge, A., Kirwan, J., Ourradi, K., Li, Y., & Sharif, M. (2020b). Multivariable logistic and linear regression models for identification of clinically useful biomarkers for osteoarthritis. *Scientific Reports*, *10*(1), 11328. https://doi.org/10.1038/s41598-020-68077-0
- Liu, C., Gao, G., Qin, X., Deng, C., & Shen, X. (2020). Correlation Analysis of C-terminal telopeptide of collagen type II and Interleukin-1β for Early Diagnosis of Knee Osteoarthritis. *Orthopaedic Surgery*, 12(1), 286–294. https://doi.org/10.1111/os.12586
- Loeser, R. F. (2006). Molecular Mechanisms of Cartilage Destruction: Mechanics, Inflammatory Mediators, and Aging Collide. Arthritis and Rheumatism, 54(5), 1357–1360. https://doi.org/10.1002/art.21813

- Loeser, R. F. (2009). Aging and Osteoarthritis: The Role of Chondrocyte Senescence and Aging Changes in the Cartilage Matrix. Osteoarthritis and Cartilage / OARS, Osteoarthritis Research Society, 17(8), 971–979. https://doi.org/10.1016/j.joca. 2009.03.002
- Loeser, R. F., Collins, J. A., & Diekman, B. O. (2016). Ageing and the pathogenesis of osteoarthritis. *Nature Reviews. Rheumatology*, *12*(7), 412–420. https://doi.org/10. 1038/nrrheum.2016.65
- Loeser, R. F., Goldring, S. R., Scanzello, C. R., & Goldring, M. B. (2012). Osteo-arthritis: A disease of the joint as an organ. *Arthritis & Rheumatism*, 64(6), 1697–1707. https://doi.org/10.1002/art.34453
- López-Otín, C., Blasco, M. A., Partridge, L., Serrano, M., & Kroemer, G. (2013). The Hallmarks of Aging. *Cell*, *153*(6), 1194–1217. https://doi.org/10.1016/j.cell.2013. 05.039
- Losina, E., Paltiel, A. D., Weinstein, A. M., Yelin, E., Hunter, D. J., Chen, S. P., Klara, K., Suter, L. G., Solomon, D. H., Burbine, S. A., Walensky, R. P., & Katz, J. N. (2015). Lifetime medical costs of knee osteoarthritis management in the United States: Impact of extending indications for total knee arthroplasty. *Arthritis Care & Research*, 67(2), 203–215. https://doi.org/10.1002/acr.22412
- Lotz, M., & Loeser, R. F. (2012). Effects of aging on articular cartilage homeostasis. *Bone*, 51(2), 241–248. https://doi.org/10.1016/j.bone.2012.03.023
- Luo, Y., He, Y., Reker, D., Gudmann, N. S., Henriksen, K., Simonsen, O., Ladel, C., Michaelis, M., Mobasheri, A., Karsdal, M., & Bay-Jensen, A.-C. (2018). A Novel High Sensitivity Type II Collagen Blood-Based Biomarker, PRO-C2, for Assessment of Cartilage Formation. *International Journal of Molecular Sciences*, 19(11), E3485. https://doi.org/10.3390/ijms19113485
- Luo, Y., Samuels, J., Krasnokutsky, S., Byrjalsen, I., Kraus, V. B., He, Y., Karsdal, M. A., Abramson, S. B., Attur, M., & Bay-Jensen, A. C. (2021). A low cartilage formation and repair endotype predicts radiographic progression of symptomatic knee osteoarthritis. *Journal of Orthopaedics and Traumatology*, 22(1), 10. https://doi.org/10.1186/s10195-021-00572-0
- Luyten, F. P., Bierma-Zeinstra, S., Dell'Accio, F., Kraus, V. B., Nakata, K., Sekiya, I., Arden, N. K., & Lohmander, L. S. (2018). Toward classification criteria for early osteoarthritis of the knee. *Seminars in Arthritis and Rheumatism*, 47(4), 457–463. https://doi.org/10.1016/j.semarthrit.2017.08.006
- Luyten, F. P., Denti, M., Filardo, G., Kon, E., & Engebretsen, L. (2012). Definition and classification of early osteoarthritis of the knee. *Knee Surgery, Sports Traumatology, Arthroscopy*, 20(3), 401–406. https://doi.org/10.1007/s00167-011-1743-2
- Lv, Z., & Shi, D. (2021). Molecule-based osteoarthritis diagnosis comes of age. *Annals of Translational Medicine*, *9*(14), 1112. https://doi.org/10.21037/atm-21-1745
- Lv, Z., Yang, Y. X., Li, J., Fei, Y., Guo, H., Sun, Z., Lu, J., Xu, X., Jiang, Q., Ikegawa, S., & Shi, D. (2021). Molecular Classification of Knee Osteoarthritis. Frontiers in Cell and Developmental Biology, 9, 725568. https://doi.org/10.3389/fcell.2021. 725568
- Mabey, T., & Honsawek, S. (2015). Cytokines as biochemical markers for knee osteo-arthritis. *World Journal of Orthopedics*, 6(1), 95–105. https://doi.org/10.5312/wjo.v6.i1.95
- Madry, H., Kon, E., Condello, V., Peretti, G. M., Steinwachs, M., Seil, R., Berruto, M., Engebretsen, L., Filardo, G., & Angele, P. (2016). Early osteoarthritis of the knee.

- Knee Surgery, Sports Traumatology, Arthroscopy, 24(6), 1753–1762. https://doi.org/10.1007/s00167-016-4068-3
- Mahmoudian, A., Lohmander, L. S., Mobasheri, A., Englund, M., & Luyten, F. P. (2021). Early-stage symptomatic osteoarthritis of the knee—Time for action. *Nature Reviews Rheumatology*, 17(10), 621–632. https://doi.org/10.1038/s41584-021-00673-4
- Malemud, C. J. (2019). Inhibition of MMPs and ADAM/ADAMTS. *Biochemical Pharmacology*, 165, 33–40. https://doi.org/10.1016/j.bcp.2019.02.033
- Marot, V., Murgier, J., Carrozzo, A., Reina, N., Monaco, E., Chiron, P., Berard, E., & Cavaignac, E. (2019). Determination of normal KOOS and WOMAC values in a healthy population. *Knee Surgery, Sports Traumatology, Arthroscopy*, 27(2), 541–548. https://doi.org/10.1007/s00167-018-5153-6
- Martel-Pelletier, J., Barr, A. J., Cicuttini, F. M., Conaghan, P. G., Cooper, C., Goldring, M. B., Goldring, S. R., Jones, G., Teichtahl, A. J., & Pelletier, J.-P. (2016). Osteoarthritis. *Nature Reviews Disease Primers*, 2(1), 1–18. https://doi.org/10.1038/nrdp.2016.72
- Martel-Pelletier, J., Boileau, C., Pelletier, J.-P., & Roughley, P. J. (2008). Cartilage in normal and osteoarthritis conditions. *Best Practice & Research Clinical Rheumatology*, 22(2), 351–384. https://doi.org/10.1016/j.berh.2008.02.001
- Mathiessen, A., & Conaghan, P. G. (2017). Synovitis in osteoarthritis: Current understanding with therapeutic implications. *Arthritis Research & Therapy*, 19(1), 18. https://doi.org/10.1186/s13075-017-1229-9
- Mazzuca, S. A., Poole, A. R., Brandt, K. D., Katz, B. P., Lane, K. A., & Lobanok, T. (2006). Associations between joint space narrowing and molecular markers of collagen and proteoglycan turnover in patients with knee osteoarthritis. *The Journal of Rheumatology*, 33(6), 1147–1151.
- McAlindon, T. E., Driban, J. B., Henrotin, Y., Hunter, D. J., Jiang, G.-L., Skou, S. T., Wang, S., & Schnitzer, T. (2015). OARSI Clinical Trials Recommendations: Design, conduct, and reporting of clinical trials for knee osteoarthritis. *Osteoarthritis and Cartilage*, 23(5), 747–760. https://doi.org/10.1016/j.joca.2015.03.005
- McAlindon, T. E., LaValley, M. P., Harvey, W. F., Price, L. L., Driban, J. B., Zhang, M., & Ward, R. J. (2017). Effect of Intra-articular Triamcinolone vs Saline on Knee Cartilage Volume and Pain in Patients With Knee Osteoarthritis: A Randomized Clinical Trial. *JAMA*, 317(19), 1967–1975. https://doi.org/10.1001/jama.2017.5283
- McAlindon, T. E., Snow, S., Cooper, C., & Dieppe, P. A. (1992). Radiographic patterns of osteoarthritis of the knee joint in the community: The importance of the patellofemoral joint. *Annals of the Rheumatic Diseases*, *51*(7), 844–849. Scopus. https://doi.org/10.1136/ard.51.7.844
- Metcalfe, A. J., Andersson, M. L., Goodfellow, R., & Thorstensson, C. A. (2012). Is knee osteoarthritis a symmetrical disease? Analysis of a 12 year prospective cohort study. BMC Musculoskeletal Disorders, 13(1), 153. https://doi.org/10.1186/1471-2474-13-153
- Meulenbelt, I., Kloppenburg, M., Kroon, H. M., Houwing-Duistermaat, J. J., Garnero, P., Graverand, M. H. L., DeGroot, J., & Slagboom, P. E. (2006). Urinary CTX-II levels are associated with radiographic subtypes of osteoarthritis in hip, knee, hand, and facet joints in subject with familial osteoarthritis at multiple sites: The GARP study. *Annals of the Rheumatic Diseases*, 65(3), 360–365. https://doi.org/10.1136/ard.2005.040642

- Migliore, A., & Massafra, U. (2014). Towards the identification of early stage osteoarthritis. *Clinical Cases in Mineral & Bone Metabolism*, 11(2), 114–116. http://search.ebscohost.com/login.aspx?direct=true&db=a9h&AN=99717583&site=ehost-live
- Mitchell, P. G., Magna, H. A., Reeves, L. M., Lopresti-Morrow, L. L., Yocum, S. A., Rosner, P. J., Geoghegan, K. F., & Hambor, J. E. (1996). Cloning, expression, and type II collagenolytic activity of matrix metalloproteinase-13 from human osteoarthritic cartilage. *Journal of Clinical Investigation*, 97(3), 761–768. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC507114/
- Mobasheri, A., Bay-Jensen, A.-C., Gualillo, O., Larkin, J., Levesque, M. C., & Henrotin, Y. (2017). Soluble biochemical markers of osteoarthritis: Are we close to using them in clinical practice? *Best Practice & Research Clinical Rheumatology*, 31(5), 705–720. https://doi.org/10.1016/j.berh.2018.07.002
- Mobasheri, A., Im, G., Katz, J. N., Loughlin, J., Kraus, V. B., Sandell, L. J., Berenbaum, F., Abramson, S., Lotz, M., Hochberg, M., Pelletier, J.-P., Madry, H., Block, J. A., Lohmander, L. S., & Altman, R. D. (2021). Osteoarthritis Research Society International (OARSI): Past, present and future. *Osteoarthritis and Cartilage Open*, *3*(2), 100146. https://doi.org/10.1016/j.ocarto.2021.100146
- Mobasheri, A., Lambert, C., & Henrotin, Y. (2019). Coll2-1 and Coll2-1NO2 as exemplars of collagen extracellular matrix turnover biomarkers to facilitate the treatment of osteoarthritis? *Expert Review of Molecular Diagnostics*, *19*(9), 803–812. https://doi.org/10.1080/14737159.2019.1646641
- Mobasheri, A., Matta, C., Zákány, R., & Musumeci, G. (2015). Chondrosenescence: Definition, hallmarks and potential role in the pathogenesis of osteoarthritis. *Maturitas*, 80(3), 237–244. https://doi.org/10.1016/j.maturitas.2014.12.003
- Mobasheri, A., Saarakkala, S., Finnilä, M., Karsdal, M. A., Bay-Jensen, A.-C., & Spil, W. E. van. (2019). *Recent advances in understanding the phenotypes of osteo-arthritis* (8:2091). F1000Research. https://doi.org/10.12688/f1000research.20575.1
- Mobasheri, A., van Spil, W. E., Budd, E., Uzieliene, I., Bernotiene, E., Bay-Jensen, A.-C., Larkin, J., Levesque, M. C., Gualillo, O., & Henrotin, Y. (2019). Molecular taxonomy of osteoarthritis for patient stratification, disease management and drug development: Biochemical markers associated with emerging clinical phenotypes and molecular endotypes. *Current Opinion in Rheumatology*, 31(1), 80–89. https://doi.org/10.1097/BOR.000000000000000567
- Muthuri, S. G., Hui, M., Doherty, M., & Zhang, W. (2011). What if we prevent obesity? Risk reduction in knee osteoarthritis estimated through a meta-analysis of observational studies. *Arthritis Care & Research*, 63(7), 982–990. https://doi.org/10. 1002/acr.20464
- Nagaosa, Y., Mateus, M., Hassan, B., Lanyon, P., & Doherty, M. (2000). Development of a logically devised line drawing atlas for grading of knee osteoarthritis. *Annals of the Rheumatic Diseases*, 59(8), 587–595. https://doi.org/10.1136/ard.59.8.587
- Nelson, A. E., Golightly, Y. M., Renner, J. B., Schwartz, T. A., Kraus, V. B., Helmick, C. G., & Jordan, J. M. (2013). Differences in Multi-joint Symptomatic Osteoarthritis Phenotypes by Race and Gender: The Johnston County Osteoarthritis Project. *Arthritis and Rheumatism*, 65(2), 373–377. https://doi.org/10.1002/art.37775
- Nelson, A. E., Smith, M. W., Golightly, Y. M., & Jordan, J. M. (2014). "Generalized Osteoarthritis": A Systematic Review. *Seminars in Arthritis and Rheumatism*, 43(6), 713–720. https://doi.org/10.1016/j.semarthrit.2013.12.007

- Nemirovskiy, O. V., Dufield, D. R., Sunyer, T., Aggarwal, P., Welsch, D. J., & Mathews, W. R. (2007). Discovery and development of a type II collagen neoepitope (TIINE) biomarker for matrix metalloproteinase activity: From in vitro to in vivo. *Analytical Biochemistry*, *361*(1), 93–101. https://doi.org/10.1016/j.ab. 2006.10.034
- Neogi, T. (2013). The epidemiology and impact of pain in osteoarthritis. *Osteoarthritis and Cartilage*, 21(9), 1145–1153. https://doi.org/10.1016/j.joca.2013.03.018
- Ogura, T., Ackermann, J., Barbieri Mestriner, A., Merkely, G., & Gomoll, A. H. (2020). Minimal Clinically Important Differences and Substantial Clinical Benefit in Patient-Reported Outcome Measures after Autologous Chondrocyte Implantation. *Cartilage*, 11(4), 412–422. https://doi.org/10.1177/1947603518799839
- O'Neill, T. W., McCabe, P. S., & McBeth, J. (2018). Update on the epidemiology, risk factors and disease outcomes of osteoarthritis. *Best Practice & Research Clinical Rheumatology*, 32(2), 312–326. https://doi.org/10.1016/j.berh.2018.10.007
- Oo, W. M., Little, C., Duong, V., & Hunter, D. J. (2021). The Development of Disease-Modifying Therapies for Osteoarthritis (DMOADs): The Evidence to Date. *Drug Design, Development and Therapy*, 15, 2921–2945. https://doi.org/10.2147/DDDT. S295224
- Outerbridge, R. E. (1961). The etiology of chondromalacia patellae. *The Journal of Bone and Joint Surgery. British Volume*, 43-B, 752–757. https://doi.org/10.1302/0301-620X.43B4.752
- Pauli, C., Grogan, S. P., Patil, S., Otsuki, S., Hasegawa, A., Koziol, J., Lotz, M. K., & D'Lima, D. D. (2011). Macroscopic and histopathologic analysis of human knee menisci in aging and osteoarthritis. *Osteoarthritis and Cartilage / OARS, Osteoarthritis Research Society*, 19(9), 1132–1141. https://doi.org/10.1016/j.joca.2011. 05.008
- Perruccio, A. V., Chandran, V., Power, J. D., Kapoor, M., Mahomed, N. N., & Gandhi, R. (2017). Systemic inflammation and painful joint burden in osteoarthritis: A matter of sex? *Osteoarthritis and Cartilage*, 25(1), 53–59. https://doi.org/10.1016/j.joca.2016.08.001
- Petersson, I. F., Boegård, T., Saxne, T., Silman, A. J., & Svensson, B. (1997). Radiographic osteoarthritis of the knee classified by the Ahlbäck and Kellgren & Lawrence systems for the tibiofemoral joint in people aged 35-54 years with chronic knee pain. *Annals of the Rheumatic Diseases*, 56(8), 493–496. https://doi.org/10.1136/ard.56.8.493
- Podlipská, J., Guermazi, A., Lehenkari, P., Niinimäki, J., Roemer, F. W., Arokoski, J. P., Kaukinen, P., Liukkonen, E., Lammentausta, E., Nieminen, M. T., Tervonen, O., Koski, J. M., & Saarakkala, S. (2016). Comparison of Diagnostic Performance of Semi-Quantitative Knee Ultrasound and Knee Radiography with MRI: Oulu Knee Osteoarthritis Study. *Scientific Reports*, 6, 22365. https://doi.org/10.1038/srep22365
- Poole, A. R., Ha, N., Bourdon, S., Sayre, E. C., Guermazi, A., & Cibere, J. (2016). Ability of a Urine Assay of Type II Collagen Cleavage by Collagenases to Detect Early Onset and Progression of Articular Cartilage Degeneration: Results from a Population-based Cohort Study. *The Journal of Rheumatology*, 43(10), 1864–1870. https://doi.org/10.3899/jrheum.150917
- Poole, A. R., Nelson, F., Dahlberg, L., Tchetina, E., Kobayashi, M., Yasuda, T., Laverty, S., Squires, G., Kojima, T., Wu, W., & Billinghurst, R. C. (2003). Proteolysis of the collagen fibril in osteoarthritis. *Biochemical Society Symposia*, 70, 115–123. https://doi.org/10.1042/bss0700115

- Pratta, M. A., Su, J. L., Leesnitzer, M. A., Struglics, A., Larsson, S., Lohmander, L. S., & Kumar, S. (2006). Development and characterization of a highly specific and sensitive sandwich ELISA for detection of aggrecanase-generated aggrecan fragments. *Osteoarthritis and Cartilage*, 14(7), 702–713. https://doi.org/10.1016/j.joca.2006.01.012
- Prieto-Alhambra, D., Judge, A., Javaid, M. K., Cooper, C., Diez-Perez, A., & Arden, N. K. (2014). Incidence and risk factors for clinically diagnosed knee, hip and hand osteoarthritis: Influences of age, gender and osteoarthritis affecting other joints. *Annals of the Rheumatic Diseases*, 73(9), 1659–1664. https://doi.org/10.1136/annrheumdis-2013-203355
- Primorac, D., Molnar, V., Rod, E., Jeleč, Ž., Čukelj, F., Matišić, V., Vrdoljak, T., Hudetz, D., Hajsok, H., & Borić, I. (2020). Knee Osteoarthritis: A Review of Pathogenesis and State-Of-The-Art Non-Operative Therapeutic Considerations. *Genes*, 11(8), 854. https://doi.org/10.3390/genes11080854
- Reijman, M., Hazes, J. M. W., Bierma-Zeinstra, S. M. A., Koes, B. W., Christgau, S., Christiansen, C., Uitterlinden, A. G., & Pols, H. a. P. (2004). A new marker for osteoarthritis: Cross-sectional and longitudinal approach. *Arthritis & Rheumatism*, 50(8), 2471–2478. https://doi.org/10.1002/art.20332
- Ren, G., & Krawetz, R. J. (2018). Biochemical Markers for the Early Identification of Osteoarthritis: Systematic Review and Meta-Analysis. *Molecular Diagnosis & Therapy*, 22(6), 671–682. https://doi.org/10.1007/s40291-018-0362-8
- Richter, M., Trzeciak, T., Owecki, M., Pucher, A., & Kaczmarczyk, J. (2015). The role of adipocytokines in the pathogenesis of knee joint osteoarthritis. *International Orthopaedics*, 39(6), 1211–1217. https://doi.org/10.1007/s00264-015-2707-9
- Riegger, J., Rehm, M., Büchele, G., Brenner, H., Günther, K.-P., Rothenbacher, D., & Brenner, R. E. (2020). Serum Cartilage Oligomeric Matrix Protein in Late-Stage Osteoarthritis: Association with Clinical Features, Renal Function, and Cardio-vascular Biomarkers. *Journal of Clinical Medicine*, *9*(1), 268. https://doi.org/10.3390/jcm9010268
- Robin Poole, A., Ionescu, M., Fitzcharles, M. A., & Clark Billinghursta, R. (2004). The assessment of cartilage degradation in vivo: Development of an immunoassay for the measurement in body fluids of type II collagen cleaved by collagenases. *Journal of Immunological Methods*, 294(1–2), 145–153. https://doi.org/10.1016/j.jim. 2004.09.005
- Roemer, F. W., Eckstein, F., Hayashi, D., & Guermazi, A. (2014). The role of imaging in osteoarthritis. *Best Practice & Research Clinical Rheumatology*, 28(1), 31–60. https://doi.org/10.1016/j.berh.2014.02.002
- Roemer, F. W., Kwoh, C. K., Hannon, M. J., Hunter, D. J., Eckstein, F., Fujii, T., Boudreau, R. M., & Guermazi, A. (2015). What comes first? Multitissue involvement leading to radiographic osteoarthritis: magnetic resonance imaging-based trajectory analysis over four years in the osteoarthritis initiative. *Arthritis & Rheumatology (Hoboken, N.J.)*, 67(8), 2085–2096. https://doi.org/10.1002/art.39176
- Rolfson, O., Wissig, S., van Maasakkers, L., Stowell, C., Ackerman, I., Ayers, D., Barber, T., Benzakour, T., Bozic, K., Budhiparama, N., Caillouette, J., Conaghan, P. G., Dahlberg, L., Dunn, J., Grady-Benson, J., Ibrahim, S. A., Lewis, S., Malchau, H., Manzary, M., ... Franklin, P. D. (2016). Defining an International Standard Set of Outcome Measures for Patients With Hip or Knee Osteoarthritis: Consensus of the International Consortium for Health Outcomes Measurement Hip and Knee

- Osteoarthritis Working Group. *Arthritis Care & Research*, 68(11), 1631–1639. https://doi.org/10.1002/acr.22868
- Roos, E. M., Roos, H. P., Lohmander, L. S., Ekdahl, C., & Beynnon, B. D. (1998). Knee Injury and Osteoarthritis Outcome Score (KOOS)—Development of a self-administered outcome measure. *The Journal Of Orthopaedic And Sports Physical Therapy*, 28(2), 88–96.
- Rousseau, J.-C., Chapurlat, R., & Garnero, P. (2021). Soluble biological markers in osteoarthritis. *Therapeutic Advances in Musculoskeletal Disease*, *13*, 1759720X211040300. https://doi.org/10.1177/1759720X211040300
- Rousseau, J.-C., Sandell, L. J., Delmas, P. D., & Garnero, P. (2004). Development and clinical application in arthritis of a new immunoassay for serum type IIA procollagen NH2 propeptide. *Methods in Molecular Medicine*, *101*, 25–37. https://doi.org/10.1385/1-59259-821-8:025
- Ryd, L., Brittberg, M., Eriksson, K., Jurvelin, J. S., Lindahl, A., Marlovits, S., Möller, P., Richardson, J. B., Steinwachs, M., & Zenobi-Wong, M. (2015). Pre-Osteoarthritis: Definition and Diagnosis of an Elusive Clinical Entity. *Cartilage*, 6(3), 156–165. https://doi.org/10.1177/1947603515586048
- Saberi Hosnijeh, F., Runhaar, J., van Meurs, J. B. J., & Bierma-Zeinstra, S. M. (2015). Biomarkers for osteoarthritis: Can they be used for risk assessment? A systematic review. *Maturitas*, 82(1), 36–49. https://doi.org/10.1016/j.maturitas.2015.04.004
- Saklatvala, J. (1986). Tumour necrosis factor alpha stimulates resorption and inhibits synthesis of proteoglycan in cartilage. *Nature*, 322(6079), 547–549. https://doi.org/10.1038/322547a0
- Sanchez, C., Bay-Jensen, A.-C., Pap, T., Dvir-Ginzberg, M., Quasnichka, H., Barrett-Jolley, R., Mobasheri, A., & Henrotin, Y. (2017). Chondrocyte secretome: A source of novel insights and exploratory biomarkers of osteoarthritis. *Osteoarthritis and Cartilage*, 25(8), 1199–1209. https://doi.org/10.1016/j.joca.2017.02.797
- Sandell, L. J., & Aigner, T. (2001). Articular cartilage and changes in Arthritis: Cell biology of osteoarthritis. Arthritis Research & Therapy, 3(2), 107. https://doi.org/10. 1186/ar148
- Scanzello, C. R. (2017). Role of low-grade inflammation in osteoarthritis. *Current Opinion in Rheumatology*, 29(1), 79–85. https://doi.org/10.1097/BOR.000000000000353
- Scanzello, C. R., & Goldring, S. R. (2012). The Role of Synovitis in Osteoarthritis pathogenesis. *Bone*, *51*(2), 249–257. https://doi.org/10.1016/j.bone.2012.02.012
- Scanzello, C. R., McKeon, B., Swaim, B. H., DiCarlo, E., Asomugha, E. U., Kanda, V., Nair, A., Lee, D. M., Richmond, J. C., Katz, J. N., Crow, M. K., & Goldring, S. R. (2011). Synovial inflammation in patients undergoing arthroscopic meniscectomy: Molecular characterization and relationship with symptoms. *Arthritis and Rheumatism*, 63(2), 391–400. https://doi.org/10.1002/art.30137
- Schiphof, D., de Klerk, B. M., Koes, B. W., & Bierma-Zeinstra, S. (2008). Good reliability, questionable validity of 25 different classification criteria of knee osteoarthritis: A systematic appraisal. *Journal of Clinical Epidemiology*, 61(12), 1205-1215.e2. https://doi.org/10.1016/j.jclinepi.2008.04.003
- Sekar, S., Shafie, S. R., Prasadam, I., Crawford, R., Panchal, S. K., Brown, L., & Xiao, Y. (2017). Saturated fatty acids induce development of both metabolic syndrome and osteoarthritis in rats. *Scientific Reports*, 7, 46457. https://doi.org/10.1038/srep46457

- Sellam, J., & Berenbaum, F. (2010). The role of synovitis in pathophysiology and clinical symptoms of osteoarthritis. *Nature Reviews Rheumatology*, *6*(11), 625–635. https://doi.org/10.1038/nrrheum.2010.159
- Sharif, M., Kirwan, J. R., Elson, C. J., Granell, R., & Clarke, S. (2004). Suggestion of nonlinear or phasic progression of knee osteoarthritis based on measurements of serum cartilage oligomeric matrix protein levels over five years. *Arthritis & Rheumatism*, 50(8), 2479–2488. https://doi.org/10.1002/art.20365
- Sharma, L. (2021). Osteoarthritis of the Knee. New England Journal of Medicine. https://doi.org/10.1056/NEJMcp1903768
- Siebuhr, A., Bay-Jensen, A., Jordan, J., Kjelgaard-Petersen, C., Christiansen, C., Abramson, S., Attur, M., Berenbaum, F., Kraus, V., & Karsdal, M. (2016). Inflammation (or synovitis)-driven osteoarthritis: An opportunity for personalizing prognosis and treatment? *Scandinavian Journal of Rheumatology*, 45(2), 87–98. https://doi.org/10.3109/03009742.2015.1060259
- Siebuhr, A. S., Bay-Jensen, A. C., Leeming, D. J., Plat, A., Byrjalsen, I., Christiansen, C., van de Heijde, D., & Karsdal, M. A. (2013). Serological identification of fast progressors of structural damage with rheumatoid arthritis. *Arthritis Research & Therapy*, 15(4), R86. https://doi.org/10.1186/ar4266
- Siebuhr, A. S., He, Y., Gudmann, N. S., Gram, A., Kjelgaard-Petersen, C. F., Qvist, P., Karsdal, M. A., & Bay-Jensen, A. C. (2014a). Biomarkers of cartilage and surrounding joint tissue. *Biomarkers in Medicine*, 8(5), 713–731. https://doi.org/10.2217/bmm.13.144
- Siebuhr, A. S., He, Y., Gudmann, N. S., Gram, A., Kjelgaard-Petersen, C. F., Qvist, P., Karsdal, M. A., & Bay-Jensen, A. C. (2014b). Biomarkers of cartilage and surrounding joint tissue. *Biomarkers in Medicine*, 8(5), 713–731. https://doi.org/10.2217/bmm.13.144
- Siebuhr, A. S., Petersen, K. K., Arendt-Nielsen, L., Egsgaard, L. L., Eskehave, T., Christiansen, C., Simonsen, O., Hoeck, H. C., Karsdal, M. A., & Bay-Jensen, A. C. (2014). Identification and characterisation of osteoarthritis patients with inflammation derived tissue turnover. *Osteoarthritis and Cartilage*, 22(1), 44–50. https://doi.org/10.1016/j.joca.2013.10.020
- Silverwood, V., Blagojevic-Bucknall, M., Jinks, C., Jordan, J. L., Protheroe, J., & Jordan, K. P. (2015). Current evidence on risk factors for knee osteoarthritis in older adults: A systematic review and meta-analysis. *Osteoarthritis and Cartilage*, 23(4), 507–515. https://doi.org/10.1016/j.joca.2014.11.019
- Skjøt-Arkil, H., Schett, G., Zhang, C., Larsen, D. V., Wang, Y., Zheng, Q., Larsen, M. R., Nawrocki, A., Bay-Jensen, A. C., Henriksen, K., Christiansen, C., Alexandersen, P., Leeming, D. J., & Karsdal, M. A. (2012). Investigation of two novel biochemical markers of inflammation, matrix metalloproteinase and cathepsin generated fragments of C-reactive protein, in patients with ankylosing spondylitis. *Clinical and Experimental Rheumatology*, 30(3), 371–379.
- Spector, T. D., Hart, D. J., Byrne, J., Harris, P. A., Dacre, J. E., & Doyle, D. V. (1993). Definition of osteoarthritis of the knee for epidemiological studies. *Annals of the Rheumatic Diseases*, 52(11), 790–794. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1005190/
- Spil, W. E. van, Drossaers-Bakker, K. W., & Lafeber, F. P. (2013). Associations of CTX-II with biochemical markers of bone turnover raise questions on its tissue origin: Data from CHECK, a cohort study of early osteoarthritis. *Annals of the*

- Rheumatic Diseases, 72(1), 29–36. https://doi.org/10.1136/annrheumdis-2011-201177
- Srikanth, V. K., Fryer, J. L., Zhai, G., Winzenberg, T. M., Hosmer, D., & Jones, G. (2005). A meta-analysis of sex differences prevalence, incidence and severity of osteoarthritis. *Osteoarthritis and Cartilage*, 13(9), 769–781. https://doi.org/10.1016/j.joca.2005.04.014
- Stanton, H., Rogerson, F. M., East, C. J., Golub, S. B., Lawlor, K. E., Meeker, C. T., Little, C. B., Last, K., Farmer, P. J., Campbell, I. K., Fourie, A. M., & Fosang, A. J. (2005). ADAMTS5 is the major aggrecanase in mouse cartilage in vivo and in vitro. *Nature*, 434(7033), 648–652. https://doi.org/10.1038/nature03417
- Steinberg, J., Ritchie, G. R. S., Roumeliotis, T. I., Jayasuriya, R. L., Clark, M. J., Brooks, R. A., Binch, A. L. A., Shah, K. M., Coyle, R., Pardo, M., Le Maitre, C. L., Ramos, Y. F. M., Nelissen, R. G. H. H., Meulenbelt, I., McCaskie, A. W., Choudhary, J. S., Wilkinson, J. M., & Zeggini, E. (2017). Integrative epigenomics, transcriptomics and proteomics of patient chondrocytes reveal genes and pathways involved in osteoarthritis. *Scientific Reports*, 7(1), 8935. https://doi.org/10.1038/s41598-017-09335-6
- Struglics, A., Larsson, S., Kumahashi, N., Frobell, R., & Lohmander, L. S. (2015). Changes in Cytokines and Aggrecan ARGS Neoepitope in Synovial Fluid and Serum and in C-Terminal Crosslinking Telopeptide of Type II Collagen and N-Terminal Crosslinking Telopeptide of Type I Collagen in Urine Over Five Years After Anterior Cruciate Ligament Rupture: An Exploratory Analysis in the Knee Anterior Cruciate Ligament, Nonsurgical Versus Surgical Treatment Trial. Arthritis & Rheumatology (Hoboken, N.J.), 67(7), 1816–1825. https://doi.org/10.1002/art.39146
- Stürmer, T., Sun, Y., Sauerland, S., Zeissig, I., Günther, K. P., Puhl, W., & Brenner, H. (1998). Serum cholesterol and osteoarthritis. The baseline examination of the Ulm Osteoarthritis Study. *The Journal of Rheumatology*, 25(9), 1827–1832.
- Sun, Y., Mauerhan, D. R., Kneisl, J. S., James Norton, H., Zinchenko, N., Ingram, J., Hanley, E. N., & Gruber, H. E. (2012). Histological Examination of Collagen and Proteoglycan Changes in Osteoarthritic Menisci. *The Open Rheumatology Journal*, 6, 24–32. https://doi.org/10.2174/1874312901206010024
- Sutton, S., Clutterbuck, A., Harris, P., Gent, T., Freeman, S., Foster, N., Barrett-Jolley, R., & Mobasheri, A. (2009). The contribution of the synovium, synovial derived inflammatory cytokines and neuropeptides to the pathogenesis of osteoarthritis. *The Veterinary Journal*, 179(1), 10–24. https://doi.org/10.1016/j.tvjl.2007.08.013
- Sweet, M. B. E., Coelho, A., Schnitzler, C. M., Schnitzer, T. J., Lenz, M. E., Jakim, I., Kuettner, K. E., & Thonar, E. J.-M. A. (1988). Serum keratan sulfate levels in osteoarthritis patients. *Arthritis & Rheumatism*, 31(5), 648–652. https://doi.org/10.1002/art.1780310510
- Taljanovic, M. S., Graham, A. R., Benjamin, J. B., Gmitro, A. F., Krupinski, E. A., Schwartz, S. A., Hunter, T. B., & Resnick, D. L. (2008). Bone marrow edema pattern in advanced hip osteoarthritis: Quantitative assessment with magnetic resonance imaging and correlation with clinical examination, radiographic findings, and histopathology. Skeletal Radiology, 37(5), 423–431. https://doi.org/10.1007/s00256-008-0446-3
- Tamm, A. E., Kumm, J., Lintrop, M., Sondergaard, B.-C., & Tamm, A. O. (2008a). 299 Knee injury and osteoarthritis outcome score (KOOS) and cartilage biomarkers in

- middle-aged women with early osteoarthritis (OA). Osteoarthritis and Cartilage, 16, S132–S133. https://doi.org/10.1016/S1063-4584(08)60343-X
- Tamm, A. E., Kumm, J., Lintrop, M., & Tamm, A. (2014). Knee function and KOOS index in subjects with different radiographic types of knee osteoarthritis based on an Estonian longitudinal study. *Osteoarthritis and Cartilage*, 22, S190. https://doi.org/10.1016/j.joca.2014.02.361
- Tamm, A. E., Lembra, K., Kumm, J., Kööbi, U., & Tamm, A. O. (2012). Lower limbs functional abilities in middle- aged estonian population with different radiographic grades of knee osteoartrhritis. *Osteoarthritis and Cartilage*, *20*, S165. https://doi.org/10.1016/j.joca.2012.02.247
- Tamm, A. E., Lintrop, M., Hansen, Y., Kumm, J., & Tamm, A. O. (2011). Associations between KOOS (knee injury and osteoarthritis outcome score) and bone–cartilage biomarkers. *Bone*, *48*, S261. https://doi.org/10.1016/j.bone.2011.03.641
- Tamm, A., Lintrop, M., Veske, K., Hansen, U., & Tamm, A. (2008b). Prevalence of patello- and tibiofemoral osteoarthritis in Elva, Southern Estonia. *The Journal of Rheumatology*, 35(3), 543–544. https://www.jrheum.org/content/35/3/543
- Tamm, A. O., Kumm, J., Tamm, A., Lintrop, M., Kukner, A., Saluse, T., Rips, L., Tein, T., Pintsaar, A., & Kukner, A. (2014). Cartilage collagen neoepitope c2c and clinical parameters in middle-aged patients with knee problems. Correlations of urinary output of C2C with cartilage lesions, koos values and functional abilities of lower limb. Osteoarthritis and Cartilage, 22, S70–S71. https://doi.org/10.1016/j.joca. 2014.02.144
- Tamm, A., Tamm, A., Kumm, J., Vija, M., & Lintrop, M. (2013). U-C2C in estonian early knee OA cohort: Progressive and non-progressive cases. *Bone Abstracts*, 1. https://doi.org/10.1530/boneabs.1.PP16
- Tarhan, S., & Unlu, Z. (2003). Magnetic resonance imaging and ultrasonographic evaluation of the patients with knee osteoarthritis: A comparative study. *Clinical Rheumatology*, 22(3), 181–188. https://doi.org/10.1007/s10067-002-0694-x
- Thorstensson, C. A., Andersson, M. L. E., Jönsson, H., Saxne, T., & Petersson, I. F. (2009). Natural course of knee osteoarthritis in middle-aged subjects with knee pain: 12-year follow-up using clinical and radiographic criteria. *Annals of the Rheumatic Diseases*, 68(12), 1890–1893. https://doi.org/10.1136/ard.2008.095158
- Troeberg, L., & Nagase, H. (2012). Proteases involved in cartilage matrix degradation in osteoarthritis. *Biochimica et Biophysica Acta*, 1824(1), 133–145. https://doi.org/10.1016/j.bbapap.2011.06.020
- Tu, C., He, J., Wu, B., Wang, W., & Li, Z. (2019). An extensive review regarding the adipokines in the pathogenesis and progression of osteoarthritis. *Cytokine*, 113, 1–12. https://doi.org/10.1016/j.cyto.2018.06.019
- Turkiewicz, A., Petersson, I. F., Bjork, J., Hawker, G., Dahlberg, L. E., Lohmander, L. S., & Englund, M. (2014). Current and future impact of osteoarthritis on health care: A population-based study with projections to year 2032. *Osteoarthritis and Cartilage*, 22(11), 1826–1832. https://doi.org/10.1016/j.joca.2014.07.015
- Uchino, M., Izumi, T., Tominaga, T., Wakita, R., Minehara, H., Sekiguchi, M., & Itoman, M. (2000). Growth factor expression in the osteophytes of the human femoral head in osteoarthritis. *Clinical Orthopaedics and Related Research*, *377*, 119–125. https://doi.org/10.1097/00003086-200008000-00017
- Valdes, A. M., Arden, N. K., Tamm, A., Kisand, K., Doherty, S., Pola, E., Cooper, C., Tamm, A., Muir, K. R., Kerna, I., Hart, D., O'Neil, F., Zhang, W., Spector, T. D., Maciewicz, R. A., & Doherty, M. (2010). A meta-analysis of interleukin-6 promoter

- polymorphisms on risk of hip and knee osteoarthritis. *Osteoarthritis and Cartilage*, 18(5), 699–704. https://doi.org/10.1016/j.joca.2009.12.012
- Valdes, A. M., Meulenbelt, I., Chassaing, E., Arden, N. K., Bierma-Zeinstra, S., Hart, D., Hofman, A., Karsdal, M., Kloppenburg, M., Kroon, H. M., Slagboom, E. P., Spector, T. D., Uitterlinden, A. G., van Meurs, J. B., & Bay-Jensen, A. C. (2014). Large scale meta-analysis of urinary C-terminal telopeptide, serum cartilage oligomeric protein and matrix metalloprotease degraded type II collagen and their role in prevalence, incidence and progression of osteoarthritis. Osteoarthritis and Cartilage, 22(5), 683–689. https://doi.org/10.1016/j.joca.2014.02.007
- Valdes, A. M., Spector, T. D., Tamm, A., Kisand, K., Doherty, S. A., Dennison, E. M., Mangino, M., Tamm, A., Kerna, I., Hart, D. J., Wheeler, M., Cooper, C., Lories, R. J., Arden, N. K., & Doherty, M. (2010). Genetic variation in the SMAD3 gene is associated with hip and knee osteoarthritis. *Arthritis & Rheumatism*, 62(8), 2347–2352. https://doi.org/10.1002/art.27530
- Valdes, A. M., Styrkarsdottir, U., Doherty, M., Morris, D. L., Mangino, M., Tamm, A., Doherty, S. A., Kisand, K., Kerna, I., Tamm, A., Wheeler, M., Maciewicz, R. A., Zhang, W., Muir, K. R., Dennison, E. M., Hart, D. J., Metrustry, S., Jonsdottir, I., Jonsson, G. F., ... Arden, N. K. (2011). Large Scale Replication Study of the Association between HLA Class II/BTNL2 Variants and Osteoarthritis of the Knee in European-Descent Populations. *PLoS ONE*, 6(8), e23371. https://doi.org/10.1371/journal.pone.0023371
- van der Esch, M., Knol, D. L., Schaffers, I. C., Reiding, D. J., van Schaardenburg, D., Knoop, J., Roorda, L. D., Lems, W. F., & Dekker, J. (2014). Osteoarthritis of the knee: Multicompartmental or compartmental disease? *Rheumatology*, *53*(3), 540–546. https://doi.org/10.1093/rheumatology/ket393
- van der Kraan, P. M., & van den Berg, W. B. (2007). Osteophytes: Relevance and biology. *Osteoarthritis and Cartilage*, *15*(3), 237–244. https://doi.org/10.1016/j.joca.2006.11.006
- van der Kraan, P. M., & van den Berg, W. B. (2008). Osteoarthritis in the context of ageing and evolution: Loss of chondrocyte differentiation block during ageing. *Ageing Research Reviews*, 7(2), 106–113. https://doi.org/10.1016/j.arr.2007.10.001
- van Saase, J. L., van Romunde, L. K., Cats, A., Vandenbroucke, J. P., & Valkenburg, H. A. (1989). Epidemiology of osteoarthritis: Zoetermeer survey. Comparison of radiological osteoarthritis in a Dutch population with that in 10 other populations. *Annals of the Rheumatic Diseases*, 48(4), 271–280. https://doi.org/10.1136/ard. 48.4.271
- van Spil, W. E., Bierma-Zeinstra, S. M. A., Deveza, L. A., Arden, N. K., Bay-Jensen, A.-C., Kraus, V. B., Carlesso, L., Christensen, R., Van Der Esch, M., Kent, P., Knoop, J., Ladel, C., Little, C. B., Loeser, R. F., Losina, E., Mills, K., Mobasheri, A., Nelson, A. E., Neogi, T., ... Hunter, D. J. (2020). A consensus-based framework for conducting and reporting osteoarthritis phenotype research. *Arthritis Research & Therapy*, *22*(1), 54. https://doi.org/10.1186/s13075-020-2143-0
- Van Spil, W. E., Kubassova, O., Boesen, M., Bay-Jensen, A.-C., & Mobasheri, A. (2019). Osteoarthritis phenotypes and novel therapeutic targets. *Biochemical Pharmacology*, *165*, 41–48. https://doi.org/10.1016/j.bcp.2019.02.037
- Van Spil, W. E., Welsing, P. M. J., Bierma-Zeinstra, S. M. A., Bijlsma, J. W. J., Roorda, L. D., Cats, H. A., & Lafeber, F. P. J. G. (2015). The ability of systemic biochemical markers to reflect presence, incidence, and progression of early-stage

- radiographic knee and hip osteoarthritis: Data from CHECK. Osteoarthritis and Cartilage, 23(8), 1388–1397. https://doi.org/10.1016/j.joca.2015.03.023
- Verma, P., & Dalal, K. (2013). Serum cartilage oligomeric matrix protein (COMP) in knee osteoarthritis: A novel diagnostic and prognostic biomarker. *Journal of Orthopaedic Research*, 31(7), 999–1006. https://doi.org/10.1002/jor.22324
- Veronese, N., Cereda, E., Maggi, S., Luchini, C., Solmi, M., Smith, T., Denkinger, M., Hurley, M., Thompson, T., Manzato, E., Sergi, G., & Stubbs, B. (2016). Osteoarthritis and mortality: A prospective cohort study and systematic review with metanalysis. Seminars in Arthritis and Rheumatism, 46(2), 160–167. https://doi.org/10.1016/j.semarthrit.2016.04.002
- Vincent, T. L. (2013). Targeting mechanotransduction pathways in osteoarthritis: A focus on the pericellular matrix. *Current Opinion in Pharmacology*, *13*(3), 449–454. https://doi.org/10.1016/j.coph.2013.01.010
- Vincent, T. L. (2019). Mechanoflammation in osteoarthritis pathogenesis. *Seminars in Arthritis and Rheumatism*, 49(3, Supplement), S36–S38. https://doi.org/10.1016/j.semarthrit.2019.09.018
- Visser, A. W., Ioan-Facsinay, A., de Mutsert, R., Widya, R. L., Loef, M., de Roos, A., le Cessie, S., den Heijer, M., Rosendaal, F. R., Kloppenburg, M., & NEO Study Group. (2014). Adiposity and hand osteoarthritis: The Netherlands Epidemiology of Obesity study. Arthritis Research & Therapy, 16(1), R19. https://doi.org/10.1186/ar4447
- Visser, A. W., Mutsert, R. de, Cessie, S. le, Heijer, M. den, Rosendaal, F. R., & Kloppenburg, M. (2015). The relative contribution of mechanical stress and systemic processes in different types of osteoarthritis: The NEO study. *Annals of the Rheumatic Diseases*, 74(10), 1842–1847. https://doi.org/10.1136/annrheumdis-2013-205012
- Vos, T., Lim, S. S., Abbafati, C., Abbas, K. M., Abbasi, M., Abbasifard, M., Abbasi-Kangevari, M., Abbastabar, H., Abd-Allah, F., Abdelalim, A., Abdollahi, M., Abdollahpour, I., Abolhassani, H., Aboyans, V., Abrams, E. M., Abreu, L. G., Abrigo, M. R. M., Abu-Raddad, L. J., Abushouk, A. I., ... Murray, C. J. L. (2020). Global burden of 369 diseases and injuries in 204 countries and territories, 1990–2019: A systematic analysis for the Global Burden of Disease Study 2019. *The Lancet*, 396(10258), 1204–1222. https://doi.org/10.1016/S0140-6736(20)30925-9
- Vuolteenaho, K., Koskinen, A., & Moilanen, E. (2014). Leptin—A link between obesity and osteoarthritis. Applications for prevention and treatment. *Basic & Clinical Pharmacology & Toxicology*, 114(1), 103–108. https://doi.org/10.1111/bcpt.12160
- Wagner, J. A. (2002). Overview of Biomarkers and Surrogate Endpoints in Drug Development. *Disease Markers*, 18(2), 41–46. https://doi.org/10.1155/2002/929274
- Wallace, I. J., Worthington, S., Felson, D. T., Jurmain, R. D., Wren, K. T., Maijanen, H., Woods, R. J., & Lieberman, D. E. (2017). Knee osteoarthritis has doubled in prevalence since the mid-20th century. *Proceedings of the National Academy of Sciences of the United States of America*, 114(35), 9332–9336. https://doi.org/10.1073/pnas.1703856114
- Walsh, D. A., Bonnet, C. S., Turner, E. L., Wilson, D., Situ, M., & McWilliams, D. F. (2007). Angiogenesis in the synovium and at the osteochondral junction in osteoarthritis. *Osteoarthritis and Cartilage*, *15*(7), 743–751. https://doi.org/10.1016/j.joca.2007.01.020
- Wancket, L. M., Baragi, V., Bove, S., Kilgore, K., Korytko, P. J., & Guzman, R. E. (2005). Anatomical Localization of Cartilage Degradation Markers in a Surgically

- Induced Rat Osteoarthritis Model. *Toxicologic Pathology*, 33(4), 484–489. https://doi.org/10.1080/01926230590965364
- Wang, P., Song, J., & Qian, D. (2019). CTX-II and YKL-40 in early diagnosis and treatment evaluation of osteoarthritis. *Experimental and Therapeutic Medicine*, 17(1), 423–431. https://doi.org/10.3892/etm.2018.6960
- Wang, X., Hunter, D., Xu, J., & Ding, C. (2015). Metabolic triggered inflammation in osteoarthritis. *Osteoarthritis and Cartilage*, 23(1), 22–30. https://doi.org/10.1016/j.joca.2014.10.002
- Wasilko, S. M., Tourville, T. W., DeSarno, M. J., Slauterbeck, J. R., Johnson, R. J., Struglics, A., & Beynnon, B. D. (2016). Relationship between synovial fluid biomarkers of articular cartilage metabolism and the patient's perspective of outcome depends on the severity of articular cartilage damage following ACL trauma. *Journal of Orthopaedic Research: Official Publication of the Orthopaedic Research Society*, 34(5), 820–827. https://doi.org/10.1002/jor.23084
- Wesseling, J., Dekker, J., van den Berg, W. B., Bierma-Zeinstra, S. M. A., Boers, M., Cats, H. A., Deckers, P., Gorter, K. J., Heuts, P. H. T. G., Hilberdink, W. K. H. A., Kloppenburg, M., Nelissen, R. G. H. H., Oosterveld, F. G. J., Oostveen, J. C. M., Roorda, L. D., Viergever, M. A., Wolde, S. ten, Lafeber, F. P. J. G., & Bijlsma, J. W. J. (2009). CHECK (Cohort Hip and Cohort Knee): Similarities and differences with the Osteoarthritis Initiative. *Annals of the Rheumatic Diseases*, 68(9), 1413–1419. https://doi.org/10.1136/ard.2008.096164
- Wojdasiewicz, P., Poniatowski, Ł. A., & Szukiewicz, D. (2014). The role of inflammatory and anti-inflammatory cytokines in the pathogenesis of osteoarthritis. *Mediators of Inflammation*, 2014, 561459. https://doi.org/10.1155/2014/561459
- Woolf, A. D. (2000). The Bone and Joint Decade 2000–2010. *Annals of the Rheumatic Diseases*, 59(2), 81–82. https://doi.org/10.1136/ard.59.2.81
- Xia, B., Chen, D., Zhang, J., Hu, S., Jin, H., & Tong, P. (2014). Osteoarthritis Pathogenesis: A Review of Molecular Mechanisms. *Calcified Tissue International*, 95(6), 495–505. https://doi.org/10.1007/s00223-014-9917-9
- Yang, Y., Li, P., Zhu, S., & Bi, R. (2020). Comparison of early-stage changes of osteoarthritis in cartilage and subchondral bone between two different rat models. *PeerJ*, 8, e8934. https://doi.org/10.7717/peerj.8934
- Yusuf, E., Kortekaas, M. C., Watt, I., Huizinga, T. W. J., & Kloppenburg, M. (2011). Do knee abnormalities visualised on MRI explain knee pain in knee osteoarthritis? A systematic review. *Annals of the Rheumatic Diseases*, 70(1), 60–67. https://doi.org/10.1136/ard.2010.131904
- Yusuf, E., Nelissen, R. G., Ioan-Facsinay, A., Stojanovic-Susulic, V., DeGroot, J., van Osch, G., Middeldorp, S., Huizinga, T. W. J., & Kloppenburg, M. (2010). Association between weight or body mass index and hand osteoarthritis: A systematic review. *Annals of the Rheumatic Diseases*, 69(4), 761–765. https://doi.org/10. 1136/ard.2008.106930
- Zhang, J. (2018). Meta-analysis of serum C-reactive protein and cartilage oligomeric matrix protein levels as biomarkers for clinical knee osteoarthritis. *BMC Musculoskeletal Disorders*, 19(1), 22. https://doi.org/10.1186/s12891-018-1932-y
- Zhang, M., Theleman, J. L., Lygrisse, K. A., & Wang, J. (2019). Epigenetic Mechanisms Underlying the Aging of Articular Cartilage and Osteoarthritis. *Gerontology*, 65(4), 387–396. https://doi.org/10.1159/000496688
- Zhang, W., Doherty, M., Peat, G., Bierma-Zeinstra, M. A., Arden, N. K., Bresnihan, B., Herrero-Beaumont, G., Kirschner, S., Leeb, B. F., Lohmander, L. S., Mazières, B.,

- Pavelka, K., Punzi, L., So, A. K., Tuncer, T., Watt, I., & Bijlsma, J. W. (2010). EULAR evidence-based recommendations for the diagnosis of knee osteoarthritis. *Annals of the Rheumatic Diseases*, 69(3), 483–489. https://doi.org/10.1136/ard.2009.113100
- Zhao, X., Shah, D., Gandhi, K., Wei, W., Dwibedi, N., Webster, L., & Sambamoorthi, U. (2019). Clinical, humanistic, and economic burden of osteoarthritis among noninstitutionalized adults in the United States. *Osteoarthritis and Cartilage*, 27(11), 1618–1626. https://doi.org/10.1016/j.joca.2019.07.002
- Zhu, Z., Ding, C., Han, W., Zheng, S., Winzenberg, T., Cicuttini, F., & Jones, G. (2018). MRI-detected osteophytes of the knee: Natural history and structural correlates of change. Arthritis Research & Therapy, 20(1), 237. https://doi.org/10.1186/s13075-018-1734-5
- Zhu, Z., Laslett, L. L., Han, W., Antony, B., Pan, F., Cicuttini, F., Jones, G., & Ding, C. (2017). Associations between MRI-detected early osteophytes and knee structure in older adults: A population-based cohort study. *Osteoarthritis and Cartilage*, *25*(12), 2055–2062. https://doi.org/10.1016/j.joca.2017.09.005

9. SUMMARY IN ESTONIAN

II tüüpi kollageeni neoepitoop C2C uriinis kui põlve osteoartriidi diagnoosimise ja kulu prognoosimise biomarker

Osteoartriit (OA) on tõsine haigus – kõige sagedasem lihasskeleti häire ja olulisim puude põhjustaja eakatel (Allen *et al.* 2021). OA haarab ligi 500 miljonit inimest maailmas (Vos *et al.* 2020), haigestumus suureneb umbes 0,3% aastas (Z. Jin *et al.* 2020). Põlveliiges on üks peamisi OA-st tabatud liigestest. Haigus võib kahjustada ühte või mõlemat põlve, kuid on kirjeldatud ka hulgiliigeste OA-d (Sharma, 2021). Põlve OA (pOA) võib haarata eraldi või mõlemat liigese funktsionaalset osa: tibiofemoraal- (TF) ja patellofemoraalliigest (PF) (Brandt *et al.* 2003).

Kuigi OA-le viitavad tunnused on avastatud juba küttide-korilaste skelettidel, siis haiguse levimuse kiire, kuni kahekordne kasv on toimunud alates 20. sajandi keskpaigast (Wallace *et al.* 2017; Berenbaum *et al.* 2018). OA tohutu sagenemise põhjuseks peetakse selliste teadaolevate riskifaktorite osakaalu tõusu nagu kõrge iga (rahvastiku vananemine), ülekaal ja rasvumine, vähene liikumine ning liigesevigastused.

Varasemalt on OA-d peetud liigese degeneratiivseks haiguseks, kõhre "kulumiseks" või hüpertroofiliseks artriidiks (Mobasheri *et al.* 2021). Kaasaja käsitluse alusel on OA patofüsioloogia kompleksne, haarates mitmeid liigesekudesid (kõhr, luu, sünoovium, menisk, ligament) molekulaarsetest muutustest kuni kudede struktuuri muutusteni (Lv & Shi, 2021). OA on etioloogiliselt heterogeenne hulgiteguriline haigus, mis kujuneb geneetiliste, mehaaniliste, metaboolsete ja põletikuliste faktorite kombineerumisel (Johnson & Hunter, 2014). Arvatakse, et OA tuleneb liigesekudede lammutus- ja paranemisprotsesside tasakaalustamatusest, mis viib liigese hävimiseni (Deveza & Loeser, 2018; Hunter & Bierma-Zeinstra, 2019).

2015. a esitas rahvusvaheline OA uurimise ühing OARSI OA definitsiooni, mille kohaselt "haigus avaldub algselt molekulaarse ümberkorraldusega (liigesekudede ainevahetuse häirumine), millele järgnevad anatoomilised ja/või füsioloogilised ümberkorraldused (kõhre lammutamine, luu remodelleerumine, osteofüütide moodustumine, liigese põletik ja liigesefunktsiooni häirumine), mis võib kulmineeruda haigustunnustena" (Kraus *et al.* 2015). See definitsioon juhib tähelepanu vajadusele diagnoosida OA-d juba presümptomaatilises, nn molekulaarses staadiumis. Arvatakse, et haigust modifitseerivate ravimite toime on kõige efektiivsem just haiguse varajastes staadiumites enne tõsiste struktuursete muutuste teket.

Peamised pOA-ga seotud radiograafilised tunnused on liigesepilu kitsenemine (LPK) ja luulised muutused, sh osteofüütide (OF) teke, kuid need väljenduvad alles haiguse hilisemas faasis ja on aeglase dünaamikaga (Altman & Gold, 2007). Molekulaarsed markerid võiksid olla tundlikumad ja dünaamilisemad kudede ainevahetuse markerid, peegeldades rohkem haiguse aktiivsust ja seeläbi ka haiguse progressiooni kiirust (Karsdal *et al.* 2010; Siebuhr *et al.*

2014a; Kraus & Karsdal, 2021). Kuna OA võib kulgeda varajases järgus kaua asümptomaatilisena, hoiatavad kudede ainevahetuse aktiivsust peegeldavad molekulaarsed markerid varakult koekahjustuse tekke eest ning seega võimaldavad OA varajasemat kliinilist diagnoosi (Kraus *et al.* 2011; Kraus & Karsdal, 2021). Samas on OA ebaühtlaselt progresseeruv haigus, mille puhul stabiilsemad perioodid vahelduvad kiiremate muutustega (Kumm *et al.* 2012; Kumm *et al.* 2013a). Seetõttu on molekulaarsed biomarkerid olulised ka haiguse kulu ja ravivastuse hindamisel (Kraus *et al.* 2011).

Kuna II tüüpi kollageen (Col2) on kõhre peamine struktuurne komponent, on OA mitteinvasiivseks ja objektiivseks hindamiseks välja töötatud üsna palju analüüse mitme erineva Col2 fragmendi kohta (uCTX-II, Coll2, Coll2-1, C1,2C jne) (Bay-Jensen *et al.* 2022). Ükski pOA molekulaarne marker ei ole veel piisavalt valideeritud kliiniliseks kasutamiseks (st ei ole saanud *in vitro* diagnostika märgist). Käesolevas uurimuses hindasime hiljuti tähelepanu pälvinud OA biomarkeri – II tüüpi kollageeni (Col2) lõhustumise neoepitoobi uriinis (uC2C) (Poole *et al.* 2016; Kraus *et al.* 2017) – kasutusvõimalusi pOA korral.

Uurimuse eesmärk ja hüpoteesid

Üldine eesmärk oli hinnata uC2C-d ja selgitada selle molekulaarse biomarkeri potentsiaali pOA diagnoosimisel ja kulu prognoosimisel haiguse erinevates staadiumites.

Töö hüpoteesid olid:

- 1. uC2C on radiograafilise pOA varajase staadiumi biomarker.
- 2. uC2C peegeldab samaaegselt OF-de ja LPK esinemist põlveliigese eri osade, TF-liigese ja PF-liigese haaratuse korral.
- 3. Sama radiograafilise pOA staadiumi korral on uC2C väärtus kõrgem isikutel, kellel haigus progresseerub.
- 4. Põlve asendusoperatsioon vähendab uC2C taset 12 kuud pärast operatsiooni. uC2C ja põlvespetsiifilise küsimustiku KOOS skooride muutus pärast operatsiooni sõltub nende operatsioonieelsest tasemest.
- 5. pOA ajal tekkivad uC2C muutused (biomarkeri dünaamika) on meestel ja naistel erinevad.

Töö spetsiifilised ülesanded:

- Määrata uC2C tase tervetel isikutel ja haigetel varajases (preradiograafilises) ja väljakujunenud pOA staadiumis ning analüüsida uC2C seoseid haiguse erinevate radiograafiliste tunnustega (OF-d ja LPK) TF- ja PF-liigestes.
- 2. Hindamaks biomarkeri võimet ennustada pOA teket, võrrelda uC2C algtasemeid kolmel uuritaval rühmal: 1) kontrollrühma isikud, kellel ei tekkinud 12 a jooksul pOA-d; 2) isikud, kellel ei teki 3 a jooksul pOA-d; 3) isikud, kellel tekib 3 a jooksul pOA.
- 3. Võrrelda uC2C baastaset pOA eri staadiumides olevatel progresseeruva haigusega ja mitteprogresseeruva haigusega isikutel.

- 4. Määrata operatsioonieelne uC2C väärtus ja biomarkeri dünaamika 12 kuu jooksul pärast põlve asendusoperatsiooni.
- 5. Võrrelda uC2C algväärtusi operatsioonijärgse KOOS-skoori muutusega, et hinnata biomarkeri võimet ennustada operatsiooni subjektiivset tulemit.
- 6. Võrrelda eelnevate ülesannete tulemusi meestel ja naistel võimalike sooliste erisuste tuvastamiseks.

Uuritavad isikud ja meetodid

Uurimuses kasutasime valimeid kolmest erinevast kohordist: Eesti varajase põlveliigese OA uuringu, artroskoopia ja põlveliigese täieliku asendamise kohordist. Kahe esimese kohordi uuritavad kuulusid nii läbilõikelise kui ka longitudinaalse uuringu valimitesse. Mõlema kohordi korral koguti igas uuringupunktis demograafilised ja kliinilised andmed, sh patsiendipõhised hinnangud: põlvespetsiifiline küsimustik KOOS (0–100% skaalal) ja valu subjektiivne hinnang (visuaalse analoogskaala (VAS) järgi, valuskoor 0–10). Lisaks põlvedele hinnati ka teisi liigeseid. Uuritavatele tehti mõlema põlve röntgenuuring, millel hinnati eraldi tibiofemoraalse (TF) ja patellofemoraalse (PF) osa OA muutusi (liigesvahemiku kitsenemist (LPK) ja osteofüüte (OF)) neljaastmelisel skaalal (astmed 0–3) Nottinghami süsteemi järgi (Nagaosa *et. al,* 2000). Mõlema põlve haaratuse korral võeti uuritavaks suurema OA raskusastmega põlv.

Põlve asendusoperatsiooni kohordi moodustasid pOA lõppstaadiumis olevad patsiendid (vanus <70 aastat), kellele tehti TÜ kliinikumi ortopeedia osakonnas esmane ühepoolne täielik põlveliigese asendusoperatsioon. Uuritavatelt koguti demograafilised ja kliinilised andmed ning uriiniproovid kolmes ajapunktis: 1–2 päeva enne operatsiooni, 3 kuud ja 12 kuud pärast operatsiooni. Kliinilise seisundi hindamiseks kasutati KOOS ja SF-36 küsimustikke ning valu visuaalset analoogskaalat, funktsionaalseid sooritusteste ja põlvede röntgenuuringuid. pOA preoperatiivseks radiograafiliseks hindamiseks kasutati kahte hindamissüsteemi: Nottinghami süsteemi ja klassikalist Kellgren-Lawrence'i (KL) süsteemi. KL-i korral tehti pOA raskusastme viieastmeline klassifikatsioon (astmed 0–4). Hinnati OF, LPK-d, subkondraalse luu sklerootiliste seintega pseudotsüstiliste piirkondade ja luuotste muutunud kuju raskusastmeid. KOOS-i muutusena määratleti muutumine rohkem kui ±20 ühiku võrra ühe aasta jooksul pärast põlveoperatsiooni.

uC2C määrasime 3 valimis järgmiselt:

- 1. Läbilõikeuuringusse kaasati 302 uuritavat, kellelt oli kogutud uriiniproov. Neist 14 uuritaval (5 mehel ja 9 naisel) hindasid ortopeedid artroskoopia ajal põlveliigese kõhre seisundit visuaalselt Outerbridge'i (modifitseeritud) süsteemi järgi. Kahjustuse pindala ja sügavuse põhjal arvutati Société Francophone d'Arthroscopie (SFA) skoor (skaala 0–100).
- 2. Longitudinaaluuringu valimisse kuulus 330 isikut, keda uuriti kahes ajapunktis: algpunktis ja järelkontrollis kolm aastat hiljem (keskmine

jälgimisperiood 38±5 kuud). uC2C määrati algpunktis kogutud uriinis. pOA progresseerumist hinnati algus- ja jälgimispunkti radiograafiliste leidude võrdlemise alusel. Progresseerujate rühma moodustasid 105 uuritavat, kellel esinesid põlveliigese radiograafilise OA progresseerumise tunnused (OF-d ja/või LPK) 3 a jooksul. See rühm jagunes omakorda kolmeks alarühmaks: (a) tekkiva pOA rühm, kuhu kuuluvatel isikutel algtasemel pOA radiograafilised tunnused puudusid, kuid need tekkisid 3 a jooksul; (b) minimaalsed progresseerujad (n=29), kellel radiograafilised tunnused süvenesid sama radiograafilise astme piires (OF-de või LPK lisandumine või suurenemine teises liigeseosas); (c) astme võrra progresseerujad (n=76), kellel radiograafiline aste süvenes vähemalt ühe astme võrra.

3. Operatsiooni prospektiivse uuringu valimisse kuulusid 86 uuritavat põlveliigese asendusoperatsiooni kohordist (n=105).

uC2C kontsentratsioon määrati hommikuses teises uriiniproovis *sandwich*-tüüpi ELISA-ga (IB-C2C-HUSATM, IBEX Pharmaceuticals Inc., Montreal, Quebec, Kanada). Uriini lahjendusteguri arvessevõtmiseks väljendasime tulemused C2C kontsentratsiooni ja kreatiniini suhtena (mõõdetud samast uriiniproovist).

Kliiniliste andmete (vanus, sugu, KMI jt) statistilisel töötlemisel kasutasime hii-ruudu testi, t-testi ja ANOVA meetodit. uC2C kontsentratsioonide seoseid analüüsisime mitteparameetriliste testidega (Kruskal-Wallise test, Mann-Whitney U-test). Segavate faktorite (vanus, sugu, KMI) arvesse võtmiseks kasutasime regressioonanalüüsi.

Uuring oli kooskõlastatud Tartu Ülikooli inimuuringute eetikakomiteega.

Tulemused ja järeldused

uC2C on integreeriv pOA marker ja on seotud haiguse mitme põhiprotsessiga: kõhre lammutamise ja osteofüütide moodustumisega emmaskummas põlveliigese osas – PF- ja TF-liigestes.

Demonstreerisime regressioonanalüüsiga, et OF-id kirjeldavad uC2C väärtust paremini kui LPK, kuid uC2C tase on kõige paremini prognoositud, kui mudelisse lisada mõlemad OA radiograafilised tunnused, hinnatuna põlveliigese mõlemas osas eraldi (TF- ja PF-liiges). Mudel ennustab uC2C taset veelgi paremini, kui selles asendada radiograafiline tunnus (TF-liigese LPK) artroskoopias hinnatava liigeskõhre makroskoopilise kahjustuse näitajaga (SFA skoor). Arvame, et mudelit võiks veelgi parandada meniski leid, kuid selle tõestuseks on vaja täiendavaid uuringuid.

uC2C on hea kandidaat pOA varajase diagnostilise testi väljaarendamiseks.

Näitasime, et uC2C kontsentratsiooni tõus uriinis esineb juba haiguse varajases staadiumis (radiograafiline aste 1), mida paljud peavad haiguse preradioloogiliseks staadiumiks.

• uC2C on naistel võimalik pOA riskimarker.

Analüüs üldistatud lineaarsete mudelitega (GLM) näitas, et C2C kõrgem algväärtus ennustab naistel väga hästi pOA teket (aste 0 muutub astmeks 1) järgneva 3 a jooksul. Parim mudeli ennustusväärtus (>90%) ilmnes tekkiva pOA grupi võrdlemisel pikaajalise (12 aastat pOA muutusteta) kontrollrühmaga.

uC2C hindamisel on olulised kaks aspekti: 1) uC2C tase on seotud pOA raskusastmega (uC2C on kõrgem suuremates astmetes); 2) kõrgem uC2C on seotud haiguse käimasoleva progressiooniga (eeskätt kehtib see naiste kohta). uC2C väärtused on kõige kõrgemad kiiresti progresseeruva haigusega isikutel enne TKR kirurgiat.

Leidsime, et uC2C mediaanväärtused kasvavad järk-järgult koos gOA astmega, näidates igas astmes mõõdukat diagnostilist võimekust eelmise astme suhtes. Iga astme väärtuste hulgas ennustab uC2C kõrgem väärtus pOA progressiooni järgneva 3 a jooksul. Lisaks tuvastasime, et uC2C on tundlik progressioonimarker: see ennustab minimaalseid radiograafilisi muutusi sama summaarse radiograafilise astme sees, milleks võib olla OF-de lisandumine või LPK süvenemine põlveliigese mistahes osas ilma astme muutuseta.

 Pärast põlveliigese asendamist on uC2C dünaamika üsna heterogeenne: uC2C eritumine uriiniga võib väheneda, suureneda või jääda muutumatuks.

Leidsime, et suhteliselt kõrgemad uC2C algväärtused seostuvad operatsioonijärgse langustrendiga ja vastupidi: Col2 lagunemine kiirenes peale operatsiooni madalate operatsioonieelsete uC2C väärtustega isikutel. Seega ei peata liigeseasendus paljudel juhtudel Col2 lagundamist organismis. Samuti selgus, et uC2C algväärtus ennustab operatsioonijärgset KOOS sümptomite skoori paranemist naistel, kuid me ei leidnud seost uC2C ja valu skoori vahel.

• uC2C näib olevat naistel võrreldes meestega parem diagnostiline ja prognostiline biomarker.

Me leidsime, et uC2C ennustab pOA progressiooni teket naistel täpsemini kui meestel ning naiste uC2C väärtused suurenevad koos haiguse raskusastme süvenemisega. Meestel olid nimetatud seosed biomarkeriga nõrgemad ja statistiliselt vähem olulised. Peab siiski mainima, et uuritud meesterühmade suurus oli suhteliselt väike, mistõttu tegemist võib olla statistilise analüüsi väikese võimsusega ja need tulemused tuleb üle kontrollida suuremal valimil.

10. ACKNOWLEDGEMENTS

The work was performed at the Department of Internal Medicine, University of Tartu. I would like to express my sincere gratitude to the people who have helped and supported me on my journey to this dissertation:

- Professor Kalle Kisand, my supervisor, for his continuous encouragement, motivation, inspiration, and unwavering support. I truly appreciate his incredible skills, excellent knowledge, and all the guidance and determination throughout the entire time. His mentorship has helped me to develop new skills and ways of working.
- Professor *emeritus* Agu Tamm, my supervisor, for his/her continuous support, thoughtful suggestions, and guidance, and for always pushing me to do better. His ideas and insights helped us to move in the right direction and to complete our studies successfully.
- Dr. Katre Maasalu and Dr. Raili Müller, reviewers of my manuscript, for their useful comments and suggestions that substantially improved the quality of this manuscript.
- Dr. Ann Tamm, my co-author, for the dedication, knowledge, and invaluable collaboration in examing the study subjects and preparing the study material.
- Professor Aare Märtson, my co-author and the head of the Department of Traumatology and Orthopedics, Tartu University Hospital, for sharing his knowledge and expertise, and organizing a patient survey in his department.
- Dr. Jaanika Kumm, Dr. Mare Lintrop, and Dr. Kristel Järv, for evaluation of knee radiographs and valuable comments.
- The surgeons of the Department of Traumatology and Orthopedics, Tartu University Hospital, who evaluated the status of knee cartilage during the arthroscopy or TKR Dr. Eiki Strauss, Dr. Anna Helena Kase, Dr. Jaan Laos, Dr. Mart Parv, Dr. Sigrid Paul, Dr. Egon Puuorg, Dr. Alo Rull, and Dr. Viktor Šapovalov, Dr. Andres Kukner, Dr. Leho Rips, Dr. Toomas Tein, Dr. Aivar Pintsaar and Dr. Toomas Saluse.
- Physiotherapist Pärt Prommik for the evaluation of functional tests of knees.
- Professor emeritus Ene-Margit Tiit for practical advice in the field of statistical analysis
- Mrs. Anne Krips for always lending a hand, when I need assistance, and for management of subjects' data and materials.
- Ms. Maret Vija and Ingrid Liimand for their technical assistance with the subjects' data and materials.
- The staff of the Department of Immunology, University of Tartu for their confidence in providing me the opportunity to use the lab devices for our experiments.
- All my colleagues and friends from the Central Laboratory of East Tallinn Central Hospital. I appreciate them for covering for my work when I was out of the laboratory. I am so thankful to have such caring and supportive

- colleagues. Thanks to Marianne Noormets for performing creatinine estimation in part of the study samples.
- I am deeply grateful to my family, particularly my husband and sons for their understanding, caring, and supporting me during all my troublesome and busy days. Thanks for always being with me. And I am thankful to my parents for always motivating me to be my best.

PUBLICATIONS

CURRICULUM VITAE

Name: Liisa Kuhi
Date of Birth: June 1, 1971
Citizenship: Estonian

Address: East Tallinn Central Hospital, Ravi 18, 10138 Tallinn, Estonia

Phone: + 372 620 7241 **E-mail:** liisa.kuhi@itk.ee

Employment

2014	East Tallinn Central Hospital, Central laboratory, Head of
	laboratory
2008–2014	East Tallinn Central Hospital, Central laboratory, Quality

manager
2002–2008 East Tallinn Central Hospital, Central laboratory, Laboratory

physician

Education

2017– University of Tartu, Faculty of Medicine, PhD studies	5
1997–2002 University of Tartu, Faculty of Medicine, Residency t	raining in
laboratory medicine	
1995–1997 University of Tartu, Faculty of Medicine, Medical pos	stgraduate
training (internship)	
1989–1995 University of Tartu, Faculty of Medicine	
1978–1989 Tartu Secondary School N 5	

Scientific work and professional organizations: Research fields

Clinical Chemistry, Laboratory Medicine, Clinical Immunology

Membership

2020	Osteoarthritis Research Society International
2010	Estonian Society for Immunology and Allergology
1998	Estonian Society of Laboratory Medicine

Publications

- 1. Kuhi, L; Tamm, AE; Kumm, J; Järv, K; Märtson A; Tamm, AO; Kisand, K (2022). Associations of Urinary Collagen II Neoepitope C2C with Total Knee Replacement Outcomes: Is OA a Systemic Disease in Rapidly Progressive Cases? Applied Sciences, 12, 1. DOI: 10.3390/app12010164.
- Kuhi Liisa; Tamm, Ann E.; Tamm, Agu O.; Kisand, Kalle (2021). Risk Assessment of the Progression of Early Knee Osteoarthritis by Collagen Neoepitope C2C: A Longitudinal Study of an Estonian Middle-

- Aged Cohort. Diagnostics, 11 (7), ARTN 1236. DOI: 10.3390/diagnostics11071236.
- 3. Nagy, E.; Infantino, M.; Bizzaro, N.; Andreeva, H.; Bontkes, H.J.; Bossuyt, X.; Fabien, N.; Fischer, K.; Heijnen, I.A.F.M.; Herold, M.; Kozmar, A.; Kuhi, L.; López-Hoyos, M.; Pullerits, R.; Sousa, M.J.R.; Tsirogianni, A.; Damoiseaux, J. (2021). The impact of the COVID-19 pandemic on autoimmune diagnostics in Europe: A lesson to be learned. Autoimmunity Reviews, 102985. DOI: 10.1016/j.autrev.2021.102985.
- 4. Kuhi, Liisa; Tamm, Ann E.; Tamm, Agu O.; Kisand, Kalle (2020). Cartilage collagen neoepitope C2C in urine as an integrative diagnostic marker for early knee osteoarthritis. Osteoarthritis and Cartilage Open, 100096. DOI: 10.1016/j.ocarto.2020.100096.
- 5. Van Hoovels, L.; Broeders, S.; Chan, E.K.L.; Andrade, L.; de Melo Cruvinel, W.; Damoiseaux, J.; Viander, M.; Herold, M.; Coucke, W.; Heijnen, I.; Bogdanos, D.; Calvo-Alen, J.; Eriksson, C.; Kozmar, A.; Kuhi, L.; Bonroy, C.; Lauwerys, B.; Schouwers, S.; Lutteri, L.; Vercammen, M. ... Bossuyt, X. (2020). Current laboratory and clinical practices in reporting and interpreting anti-nuclear antibody indirect immunofluorescence (ANA IIF) patterns: results of an international survey. Autoimmunity Highlights, 11 (1), ARTN 17. DOI: 10.1186/s13317-020-00139-9.
- 6. Kuhi, L.; Kisand, K.; Kumm, J.; Lintrop, M.; Tamm, A.; Kukner, A.; Pintsaar, A.; Rips, L.; Saluse, T.; Tein, T.; Vija, M.; Tamm, A. (2020). Urinary cartilage collagen neoepitope c2c levels in preradiographic and radiographic knee OA. Osteoarthritis and Cartilage, 28, 63–64.
- 7. Veski, M.; Kuhi, L. (2019). Müosiidispetsiifiliste ja müosiidiga seotud antikehade kliiniline olulisus. *Eesti Arst.*, 98 (Lisa 3): Tartu Ülikooli arstiteaduskonna aastapäeva teaduskonverents 2019, Tartu, Estonia, October 10-11, 2019. OÜ Celsius Healthcare, 55.
- 8. Kuhi, L.; Ellervee, E.; Tammaru, M. (2016). The role of client-initiated laboratory testing in discovering hidden diseases at East-Tallinn Central Hospital. *Eesti Arst, 95 (Supplement1): XIII Baltic Congress in Laboratory Medicine; Tartu, Estonia; May 12-14 2016.* OÜ Celsius Healthcare, 52.
- 9. Kuhi, L.; Kutt, M.; Suigom, M.; Ress, K. (2014). Lymphocyte subgroups in patients with common variable immunodeficiency. *Journal of Clinical Immunology, 34: 100th J Project Meeting; Antalya, Turkey; Mar 12-14, 2014.*, 726.
- 10. Kuhi, L.; Kütt, M. (2014). The request of anti-dsDNA test should depend on result of anti-nuclear antibodies. *Clinical Chemistry and Laboratory Medicine (CCLM)*, 52: IFCC WorldLab Istanbul 2014; Istanbul, Turkey; 22-26 June 2014., \$289. (Special Suppl).
- 11. Kuhi, L. (2013). The diagnostic performance of anti-nucleosomes, anti-dsDNA-NcX ELISA and anti-dsDNA detected by immunoblot method in diagnosing of systemic lupus erythematosus. 37: EuroMedLab 2013; Milano; 19-23 May 2013., SS.

12. Kuhi, L.; Kütt, M. (2011). The diagnostic significance of disease specific antibodies to nuclear antigens in primary biliary cirrhosis. *Clinical Chemistry and Laboratory Medicine*, 49. Saksamaa: Walter de Gruyter & Co, S208. (1).

ELULOOKIRJELDUS

Nimi: Liisa Kuhi Sünniaeg: 1. juuni 1971

Kodakondsus: Eesti

Ida-Tallinna Keskhaigla, Ravi 18, 10138 Tallinn, Eesti Aadress:

Telefon: + 372 620 7241 E-post: liisa.kuhi@itk.ee

Teenistuskäik

2014	AS Ida-Tallinna Keskhaigla, kesklabori juhataja
2008-2014	AS Ida-Tallinna Keskhaigla, kesklabori kvaliteedijuht
2002-2008	AS Ida-Tallinna Keskhaigla, laboriarst

Haridus

2017	Tartu Ülikool, meditsiiniteaduste valdkond, doktorantuur
1997-2002	Tartu Ülikool, residentuur laborimeditsiini alal
1995–1997	Tartu Ülikool, üldinternatuur
1989–1995	Tartu Ülikool, arstiteaduskond, ravi eriala
1978-1989	Tartu 5. Keskkool

Teadus- ja erialane tegevus:

Uurimisvaldkonnad

Kliiniline keemia, laborimeditsiin, kliiniline immuonoloogia

Liikmelisus

2020	Rahvusvaheline Osteoartriidi Teadusuuringute Ühing
2010	Eesti Immunoloogide ja Allergoloogide Selts
1998-	Festi Labormeditsiini Ühing

DISSERTATIONES MEDICINAE UNIVERSITATIS TARTUENSIS

- 1. **Heidi-Ingrid Maaroos**. The natural course of gastric ulcer in connection with chronic gastritis and *Helicobacter pylori*. Tartu, 1991.
- 2. **Mihkel Zilmer**. Na-pump in normal and tumorous brain tissues: Structural, functional and tumorigenesis aspects. Tartu, 1991.
- 3. **Eero Vasar**. Role of cholecystokinin receptors in the regulation of behaviour and in the action of haloperidol and diazepam. Tartu, 1992.
- 4. **Tiina Talvik**. Hypoxic-ischaemic brain damage in neonates (clinical, biochemical and brain computed tomographical investigation). Tartu, 1992.
- 5. **Ants Peetsalu**. Vagotomy in duodenal ulcer disease: A study of gastric acidity, serum pepsinogen I, gastric mucosal histology and *Helicobacter pylori*. Tartu, 1992.
- 6. **Marika Mikelsaar**. Evaluation of the gastrointestinal microbial ecosystem in health and disease. Tartu, 1992.
- 7. **Hele Everaus**. Immuno-hormonal interactions in chronic lymphocytic leukaemia and multiple myeloma. Tartu, 1993.
- 8. **Ruth Mikelsaar**. Etiological factors of diseases in genetically consulted children and newborn screening: dissertation for the commencement of the degree of doctor of medical sciences. Tartu, 1993.
- 9. **Agu Tamm**. On metabolic action of intestinal microflora: clinical aspects. Tartu, 1993.
- 10. **Katrin Gross**. Multiple sclerosis in South-Estonia (epidemiological and computed tomographical investigations). Tartu, 1993.
- 11. **Oivi Uibo**. Childhood coeliac disease in Estonia: occurrence, screening, diagnosis and clinical characterization. Tartu, 1994.
- 12. **Viiu Tuulik**. The functional disorders of central nervous system of chemistry workers. Tartu, 1994.
- 13. **Margus Viigimaa**. Primary haemostasis, antiaggregative and anticoagulant treatment of acute myocardial infarction. Tartu, 1994.
- 14. **Rein Kolk**. Atrial versus ventricular pacing in patients with sick sinus syndrome. Tartu, 1994.
- 15. **Toomas Podar**. Incidence of childhood onset type 1 diabetes mellitus in Estonia. Tartu, 1994.
- 16. **Kiira Subi**. The laboratory surveillance of the acute respiratory viral infections in Estonia. Tartu. 1995.
- 17. **Irja Lutsar**. Infections of the central nervous system in children (epidemiologic, diagnostic and therapeutic aspects, long term outcome). Tartu, 1995.
- 18. **Aavo Lang**. The role of dopamine, 5-hydroxytryptamine, sigma and NMDA receptors in the action of antipsychotic drugs. Tartu, 1995.
- 19. **Andrus Arak**. Factors influencing the survival of patients after radical surgery for gastric cancer. Tartu, 1996.

- 20. **Tõnis Karki**. Quantitative composition of the human lactoflora and method for its examination. Tartu, 1996.
- 21. **Reet Mändar**. Vaginal microflora during pregnancy and its transmission to newborn. Tartu, 1996.
- 22. **Triin Remmel**. Primary biliary cirrhosis in Estonia: epidemiology, clinical characterization and prognostication of the course of the disease. Tartu, 1996.
- 23. **Toomas Kivastik**. Mechanisms of drug addiction: focus on positive reinforcing properties of morphine. Tartu, 1996.
- 24. **Paavo Pokk**. Stress due to sleep deprivation: focus on GABA_A receptor-chloride ionophore complex. Tartu, 1996.
- 25. **Kristina Allikmets**. Renin system activity in essential hypertension. Associations with atherothrombogenic cardiovascular risk factors and with the efficacy of calcium antagonist treatment. Tartu, 1996.
- 26. **Triin Parik**. Oxidative stress in essential hypertension: Associations with metabolic disturbances and the effects of calcium antagonist treatment. Tartu, 1996.
- 27. **Svetlana Päi**. Factors promoting heterogeneity of the course of rheumatoid arthritis. Tartu, 1997.
- 28. **Maarike Sallo**. Studies on habitual physical activity and aerobic fitness in 4 to 10 years old children. Tartu, 1997.
- 29. **Paul Naaber**. *Clostridium difficile* infection and intestinal microbial ecology. Tartu, 1997.
- 30. **Rein Pähkla**. Studies in pinoline pharmacology. Tartu, 1997.
- 31. Andrus Juhan Voitk. Outpatient laparoscopic cholecystectomy. Tartu, 1997.
- 32. **Joel Starkopf**. Oxidative stress and ischaemia-reperfusion of the heart. Tartu, 1997.
- 33. **Janika Kõrv**. Incidence, case-fatality and outcome of stroke. Tartu, 1998.
- 34. **Ülla Linnamägi**. Changes in local cerebral blood flow and lipid peroxidation following lead exposure in experiment. Tartu, 1998.
- 35. **Ave Minajeva**. Sarcoplasmic reticulum function: comparison of atrial and ventricular myocardium. Tartu, 1998.
- 36. **Oleg Milenin**. Reconstruction of cervical part of esophagus by revascularised ileal autografts in dogs. A new complex multistage method. Tartu, 1998
- 37. **Sergei Pakriev**. Prevalence of depression, harmful use of alcohol and alcohol dependence among rural population in Udmurtia. Tartu, 1998.
- 38. **Allen Kaasik**. Thyroid hormone control over β-adrenergic signalling system in rat atria. Tartu, 1998.
- 39. **Vallo Matto**. Pharmacological studies on anxiogenic and antiaggressive properties of antidepressants. Tartu, 1998.
- 40. **Maire Vasar**. Allergic diseases and bronchial hyperreactivity in Estonian children in relation to environmental influences. Tartu, 1998.
- 41. **Kaja Julge**. Humoral immune responses to allergens in early childhood. Tartu, 1998.

- 42. **Heli Grünberg**. The cardiovascular risk of Estonian schoolchildren. A cross-sectional study of 9-, 12- and 15-year-old children. Tartu, 1998.
- 43. **Epp Sepp.** Formation of intestinal microbial ecosystem in children. Tartu, 1998.
- 44. **Mai Ots**. Characteristics of the progression of human and experimental glomerulopathies. Tartu, 1998.
- 45. **Tiina Ristimäe**. Heart rate variability in patients with coronary artery disease. Tartu, 1998.
- 46. **Leho Kõiv**. Reaction of the sympatho-adrenal and hypothalamo-pituitary-adrenocortical system in the acute stage of head injury. Tartu, 1998.
- 47. **Bela Adojaan**. Immune and genetic factors of childhood onset IDDM in Estonia. An epidemiological study. Tartu, 1999.
- 48. **Jakov Shlik**. Psychophysiological effects of cholecystokinin in humans. Tartu, 1999.
- 49. **Kai Kisand**. Autoantibodies against dehydrogenases of α-ketoacids. Tartu, 1999.
- 50. **Toomas Marandi**. Drug treatment of depression in Estonia. Tartu, 1999.
- 51. Ants Kask. Behavioural studies on neuropeptide Y. Tartu, 1999.
- 52. **Ello-Rahel Karelson**. Modulation of adenylate cyclase activity in the rat hippocampus by neuropeptide galanin and its chimeric analogs. Tartu, 1999.
- 53. **Tanel Laisaar**. Treatment of pleural empyema special reference to intrapleural therapy with streptokinase and surgical treatment modalities. Tartu, 1999.
- 54. **Eve Pihl**. Cardiovascular risk factors in middle-aged former athletes. Tartu, 1999.
- 55. **Katrin Õunap**. Phenylketonuria in Estonia: incidence, newborn screening, diagnosis, clinical characterization and genotype/phenotype correlation. Tartu, 1999.
- 56. **Siiri Kõljalg**. *Acinetobacter* an important nosocomial pathogen. Tartu, 1999.
- 57. **Helle Karro**. Reproductive health and pregnancy outcome in Estonia: association with different factors. Tartu, 1999.
- 58. **Heili Varendi**. Behavioral effects observed in human newborns during exposure to naturally occurring odors. Tartu, 1999.
- 59. **Anneli Beilmann**. Epidemiology of epilepsy in children and adolescents in Estonia. Prevalence, incidence, and clinical characteristics. Tartu, 1999.
- 60. **Vallo Volke**. Pharmacological and biochemical studies on nitric oxide in the regulation of behaviour. Tartu, 1999.
- 61. **Pilvi Ilves**. Hypoxic-ischaemic encephalopathy in asphyxiated term infants. A prospective clinical, biochemical, ultrasonographical study. Tartu, 1999.
- 62. **Anti Kalda**. Oxygen-glucose deprivation-induced neuronal death and its pharmacological prevention in cerebellar granule cells. Tartu, 1999.
- 63. **Eve-Irene Lepist**. Oral peptide prodrugs studies on stability and absorption. Tartu, 2000.

- 64. **Jana Kivastik**. Lung function in Estonian schoolchildren: relationship with anthropometric indices and respiratory symptomas, reference values for dynamic spirometry. Tartu, 2000.
- 65. **Karin Kull**. Inflammatory bowel disease: an immunogenetic study. Tartu, 2000.
- 66. **Kaire Innos**. Epidemiological resources in Estonia: data sources, their quality and feasibility of cohort studies. Tartu, 2000.
- 67. **Tamara Vorobjova**. Immune response to *Helicobacter pylori* and its association with dynamics of chronic gastritis and epithelial cell turnover in antrum and corpus. Tartu, 2001.
- 68. **Ruth Kalda**. Structure and outcome of family practice quality in the changing health care system of Estonia. Tartu, 2001.
- 69. **Annika Krüüner**. *Mycobacterium tuberculosis* spread and drug resistance in Estonia. Tartu, 2001.
- 70. **Marlit Veldi**. Obstructive Sleep Apnoea: Computerized Endopharyngeal Myotonometry of the Soft Palate and Lingual Musculature. Tartu, 2001.
- 71. **Anneli Uusküla**. Epidemiology of sexually transmitted diseases in Estonia in 1990–2000. Tartu, 2001.
- 72. **Ade Kallas**. Characterization of antibodies to coagulation factor VIII. Tartu, 2002.
- 73. **Heidi Annuk**. Selection of medicinal plants and intestinal lactobacilli as antimicrobil components for functional foods. Tartu, 2002.
- 74. **Aet Lukmann**. Early rehabilitation of patients with ischaemic heart disease after surgical revascularization of the myocardium: assessment of health-related quality of life, cardiopulmonary reserve and oxidative stress. A clinical study. Tartu, 2002.
- 75. **Maigi Eisen**. Pathogenesis of Contact Dermatitis: participation of Oxidative Stress. A clinical biochemical study. Tartu, 2002.
- 76. **Piret Hussar**. Histology of the post-traumatic bone repair in rats. Elaboration and use of a new standardized experimental model bicortical perforation of tibia compared to internal fracture and resection osteotomy. Tartu, 2002.
- 77. **Tõnu Rätsep**. Aneurysmal subarachnoid haemorrhage: Noninvasive monitoring of cerebral haemodynamics. Tartu, 2002.
- 78. **Marju Herodes**. Quality of life of people with epilepsy in Estonia. Tartu, 2003.
- 79. **Katre Maasalu**. Changes in bone quality due to age and genetic disorders and their clinical expressions in Estonia. Tartu, 2003.
- 80. **Toomas Sillakivi**. Perforated peptic ulcer in Estonia: epidemiology, risk factors and relations with *Helicobacter pylori*. Tartu, 2003.
- 81. **Leena Puksa**. Late responses in motor nerve conduction studies. F and A waves in normal subjects and patients with neuropathies. Tartu, 2003.
- 82. **Krista Lõivukene**. *Helicobacter pylori* in gastric microbial ecology and its antimicrobial susceptibility pattern. Tartu, 2003.

- 83. **Helgi Kolk**. Dyspepsia and *Helicobacter pylori* infection: the diagnostic value of symptoms, treatment and follow-up of patients referred for upper gastrointestinal endoscopy by family physicians. Tartu, 2003.
- 84. **Helena Soomer**. Validation of identification and age estimation methods in forensic odontology. Tartu, 2003.
- 85. **Kersti Oselin**. Studies on the human MDR1, MRP1, and MRP2 ABC transporters: functional relevance of the genetic polymorphisms in the *MDR1* and *MRP1* gene. Tartu, 2003.
- 86. **Jaan Soplepmann**. Peptic ulcer haemorrhage in Estonia: epidemiology, prognostic factors, treatment and outcome. Tartu, 2003.
- 87. **Margot Peetsalu**. Long-term follow-up after vagotomy in duodenal ulcer disease: recurrent ulcer, changes in the function, morphology and *Helico-bacter pylori* colonisation of the gastric mucosa. Tartu, 2003.
- 88. **Kersti Klaamas**. Humoral immune response to *Helicobacter pylori* a study of host-dependent and microbial factors. Tartu, 2003.
- 89. **Pille Taba**. Epidemiology of Parkinson's disease in Tartu, Estonia. Prevalence, incidence, clinical characteristics, and pharmacoepidemiology. Tartu, 2003.
- 90. **Alar Veraksitš**. Characterization of behavioural and biochemical phenotype of cholecystokinin-2 receptor deficient mice: changes in the function of the dopamine and endopioidergic system. Tartu, 2003.
- 91. **Ingrid Kalev**. CC-chemokine receptor 5 (CCR5) gene polymorphism in Estonians and in patients with Type I and Type II diabetes mellitus. Tartu, 2003.
- 92. **Lumme Kadaja**. Molecular approach to the regulation of mitochondrial function in oxidative muscle cells. Tartu, 2003.
- 93. **Aive Liigant**. Epidemiology of primary central nervous system tumours in Estonia from 1986 to 1996. Clinical characteristics, incidence, survival and prognostic factors. Tartu, 2004.
- 94. **Andres, Kulla**. Molecular characteristics of mesenchymal stroma in human astrocytic gliomas. Tartu, 2004.
- 95. **Mari Järvelaid**. Health damaging risk behaviours in adolescence. Tartu, 2004.
- 96. **Ülle Pechter**. Progression prevention strategies in chronic renal failure and hypertension. An experimental and clinical study. Tartu, 2004.
- 97. **Gunnar Tasa**. Polymorphic glutathione S-transferases biology and role in modifying genetic susceptibility to senile cataract and primary open angle glaucoma. Tartu, 2004.
- 98. **Tuuli Käämbre**. Intracellular energetic unit: structural and functional aspects. Tartu, 2004.
- 99. **Vitali Vassiljev**. Influence of nitric oxide syntase inhibitors on the effects of ethanol after acute and chronic ethanol administration and withdrawal. Tartu, 2004.

- 100. **Aune Rehema**. Assessment of nonhaem ferrous iron and glutathione redox ratio as markers of pathogeneticity of oxidative stress in different clinical groups. Tartu, 2004.
- 101. **Evelin Seppet**. Interaction of mitochondria and ATPases in oxidative muscle cells in normal and pathological conditions. Tartu, 2004.
- 102. **Eduard Maron**. Serotonin function in panic disorder: from clinical experiments to brain imaging and genetics. Tartu, 2004.
- 103. **Marje Oona**. *Helicobacter pylori* infection in children: epidemiological and therapeutic aspects. Tartu, 2004.
- 104. **Kersti Kokk**. Regulation of active and passive molecular transport in the testis. Tartu, 2005.
- 105. **Vladimir Järv**. Cross-sectional imaging for pretreatment evaluation and follow-up of pelvic malignant tumours. Tartu, 2005.
- 106. **Andre Õun**. Epidemiology of adult epilepsy in Tartu, Estonia. Incidence, prevalence and medical treatment. Tartu, 2005.
- 107. **Piibe Muda**. Homocysteine and hypertension: associations between homocysteine and essential hypertension in treated and untreated hypertensive patients with and without coronary artery disease. Tartu, 2005.
- 108. **Külli Kingo**. The interleukin-10 family cytokines gene polymorphisms in plaque psoriasis. Tartu, 2005.
- 109. **Mati Merila**. Anatomy and clinical relevance of the glenohumeral joint capsule and ligaments. Tartu, 2005.
- 110. **Epp Songisepp**. Evaluation of technological and functional properties of the new probiotic *Lactobacillus fermentum* ME-3. Tartu, 2005.
- 111. **Tiia Ainla**. Acute myocardial infarction in Estonia: clinical characteristics, management and outcome. Tartu, 2005.
- 112. **Andres Sell**. Determining the minimum local anaesthetic requirements for hip replacement surgery under spinal anaesthesia a study employing a spinal catheter. Tartu, 2005.
- 113. **Tiia Tamme**. Epidemiology of odontogenic tumours in Estonia. Pathogenesis and clinical behaviour of ameloblastoma. Tartu, 2005.
- 114. **Triine Annus**. Allergy in Estonian schoolchildren: time trends and characteristics. Tartu, 2005.
- 115. **Tiia Voor**. Microorganisms in infancy and development of allergy: comparison of Estonian and Swedish children. Tartu, 2005.
- 116. **Priit Kasenõmm**. Indicators for tonsillectomy in adults with recurrent tonsillitis clinical, microbiological and pathomorphological investigations. Tartu, 2005.
- 117. **Eva Zusinaite**. Hepatitis C virus: genotype identification and interactions between viral proteases. Tartu, 2005.
- 118. **Piret Kõll**. Oral lactoflora in chronic periodontitis and periodontal health. Tartu, 2006.
- 119. **Tiina Stelmach**. Epidemiology of cerebral palsy and unfavourable neuro-developmental outcome in child population of Tartu city and county, Estonia Prevalence, clinical features and risk factors. Tartu, 2006.

- 120. **Katrin Pudersell**. Tropane alkaloid production and riboflavine excretion in the field and tissue cultures of henbane (*Hyoscyamus niger* L.). Tartu, 2006
- 121. **Külli Jaako**. Studies on the role of neurogenesis in brain plasticity. Tartu, 2006.
- 122. **Aare Märtson**. Lower limb lengthening: experimental studies of bone regeneration and long-term clinical results. Tartu, 2006.
- 123. Heli Tähepõld. Patient consultation in family medicine. Tartu, 2006.
- 124. **Stanislav Liskmann**. Peri-implant disease: pathogenesis, diagnosis and treatment in view of both inflammation and oxidative stress profiling. Tartu, 2006.
- 125. **Ruth Rudissaar**. Neuropharmacology of atypical antipsychotics and an animal model of psychosis. Tartu, 2006.
- 126. **Helena Andreson**. Diversity of *Helicobacter pylori* genotypes in Estonian patients with chronic inflammatory gastric diseases. Tartu, 2006.
- 127. **Katrin Pruus**. Mechanism of action of antidepressants: aspects of sero-toninergic system and its interaction with glutamate. Tartu, 2006.
- 128. **Priit Põder**. Clinical and experimental investigation: relationship of ischaemia/reperfusion injury with oxidative stress in abdominal aortic aneurysm repair and in extracranial brain artery endarterectomy and possibilities of protection against ischaemia using a glutathione analogue in a rat model of global brain ischaemia. Tartu, 2006.
- 129. **Marika Tammaru**. Patient-reported outcome measurement in rheumatoid arthritis. Tartu, 2006.
- 130. **Tiia Reimand**. Down syndrome in Estonia. Tartu, 2006.
- 131. **Diva Eensoo**. Risk-taking in traffic and Markers of Risk-Taking Behaviour in Schoolchildren and Car Drivers. Tartu, 2007.
- 132. **Riina Vibo**. The third stroke registry in Tartu, Estonia from 2001 to 2003: incidence, case-fatality, risk factors and long-term outcome. Tartu, 2007.
- 133. **Chris Pruunsild**. Juvenile idiopathic arthritis in children in Estonia. Tartu, 2007.
- 134. **Eve Õiglane-Šlik**. Angelman and Prader-Willi syndromes in Estonia. Tartu, 2007.
- 135. **Kadri Haller**. Antibodies to follicle stimulating hormone. Significance in female infertility. Tartu, 2007.
- 136. **Pille Ööpik**. Management of depression in family medicine. Tartu, 2007.
- 137. **Jaak Kals**. Endothelial function and arterial stiffness in patients with atherosclerosis and in healthy subjects. Tartu, 2007.
- 138. **Priit Kampus**. Impact of inflammation, oxidative stress and age on arterial stiffness and carotid artery intima-media thickness. Tartu, 2007.
- 139. Margus Punab. Male fertility and its risk factors in Estonia. Tartu, 2007.
- 140. **Alar Toom**. Heterotopic ossification after total hip arthroplasty: clinical and pathogenetic investigation. Tartu, 2007.

- 141. **Lea Pehme**. Epidemiology of tuberculosis in Estonia 1991–2003 with special regard to extrapulmonary tuberculosis and delay in diagnosis of pulmonary tuberculosis. Tartu, 2007.
- 142. **Juri Karjagin**. The pharmacokinetics of metronidazole and meropenem in septic shock. Tartu, 2007.
- 143. **Inga Talvik**. Inflicted traumatic brain injury shaken baby syndrome in Estonia epidemiology and outcome. Tartu, 2007.
- 144. **Tarvo Rajasalu**. Autoimmune diabetes: an immunological study of type 1 diabetes in humans and in a model of experimental diabetes (in RIP-B7.1 mice). Tartu, 2007.
- 145. **Inga Karu**. Ischaemia-reperfusion injury of the heart during coronary surgery: a clinical study investigating the effect of hyperoxia. Tartu, 2007.
- 146. **Peeter Padrik**. Renal cell carcinoma: Changes in natural history and treatment of metastatic disease. Tartu, 2007.
- 147. **Neve Vendt**. Iron deficiency and iron deficiency anaemia in infants aged 9 to 12 months in Estonia. Tartu, 2008.
- 148. **Lenne-Triin Heidmets**. The effects of neurotoxins on brain plasticity: focus on neural Cell Adhesion Molecule. Tartu, 2008.
- 149. **Paul Korrovits**. Asymptomatic inflammatory prostatitis: prevalence, etiological factors, diagnostic tools. Tartu, 2008.
- 150. **Annika Reintam**. Gastrointestinal failure in intensive care patients. Tartu, 2008.
- 151. **Kristiina Roots**. Cationic regulation of Na-pump in the normal, Alzheimer's and CCK₂ receptor-deficient brain. Tartu, 2008.
- 152. **Helen Puusepp**. The genetic causes of mental retardation in Estonia: fragile X syndrome and creatine transporter defect. Tartu, 2009.
- 153. **Kristiina Rull**. Human chorionic gonadotropin beta genes and recurrent miscarriage: expression and variation study. Tartu, 2009.
- 154. **Margus Eimre**. Organization of energy transfer and feedback regulation in oxidative muscle cells. Tartu, 2009.
- 155. **Maire Link**. Transcription factors FoxP3 and AIRE: autoantibody associations. Tartu, 2009.
- 156. **Kai Haldre**. Sexual health and behaviour of young women in Estonia. Tartu, 2009.
- 157. **Kaur Liivak**. Classical form of congenital adrenal hyperplasia due to 21-hydroxylase deficiency in Estonia: incidence, genotype and phenotype with special attention to short-term growth and 24-hour blood pressure. Tartu, 2009.
- 158. **Kersti Ehrlich**. Antioxidative glutathione analogues (UPF peptides) molecular design, structure-activity relationships and testing the protective properties. Tartu, 2009.
- 159. Anneli Rätsep. Type 2 diabetes care in family medicine. Tartu, 2009.
- 160. **Silver Türk**. Etiopathogenetic aspects of chronic prostatitis: role of mycoplasmas, coryneform bacteria and oxidative stress. Tartu, 2009.

- 161. **Kaire Heilman**. Risk markers for cardiovascular disease and low bone mineral density in children with type 1 diabetes. Tartu, 2009.
- 162. **Kristi Rüütel**. HIV-epidemic in Estonia: injecting drug use and quality of life of people living with HIV. Tartu, 2009.
- 163. **Triin Eller**. Immune markers in major depression and in antidepressive treatment. Tartu, 2009.
- 164. **Siim Suutre**. The role of TGF-β isoforms and osteoprogenitor cells in the pathogenesis of heterotopic ossification. An experimental and clinical study of hip arthroplasty. Tartu, 2010.
- 165. **Kai Kliiman**. Highly drug-resistant tuberculosis in Estonia: Risk factors and predictors of poor treatment outcome. Tartu, 2010.
- 166. **Inga Villa**. Cardiovascular health-related nutrition, physical activity and fitness in Estonia. Tartu, 2010.
- 167. **Tõnis Org**. Molecular function of the first PHD finger domain of Autoimmune Regulator protein. Tartu, 2010.
- 168. **Tuuli Metsvaht**. Optimal antibacterial therapy of neonates at risk of early onset sepsis. Tartu, 2010.
- 169. **Jaanus Kahu**. Kidney transplantation: Studies on donor risk factors and mycophenolate mofetil. Tartu, 2010.
- 170. **Koit Reimand**. Autoimmunity in reproductive failure: A study on associated autoantibodies and autoantigens. Tartu, 2010.
- 171. **Mart Kull**. Impact of vitamin D and hypolactasia on bone mineral density: a population based study in Estonia. Tartu, 2010.
- 172. **Rael Laugesaar**. Stroke in children epidemiology and risk factors. Tartu, 2010.
- 173. **Mark Braschinsky**. Epidemiology and quality of life issues of hereditary spastic paraplegia in Estonia and implemention of genetic analysis in everyday neurologic practice. Tartu, 2010.
- 174. **Kadri Suija**. Major depression in family medicine: associated factors, recurrence and possible intervention. Tartu, 2010.
- 175. **Jarno Habicht**. Health care utilisation in Estonia: socioeconomic determinants and financial burden of out-of-pocket payments. Tartu, 2010.
- 176. **Kristi Abram**. The prevalence and risk factors of rosacea. Subjective disease perception of rosacea patients. Tartu, 2010.
- 177. **Malle Kuum**. Mitochondrial and endoplasmic reticulum cation fluxes: Novel roles in cellular physiology. Tartu, 2010.
- 178. **Rita Teek**. The genetic causes of early onset hearing loss in Estonian children. Tartu, 2010.
- 179. **Daisy Volmer**. The development of community pharmacy services in Estonia public and professional perceptions 1993–2006. Tartu, 2010.
- 180. **Jelena Lissitsina**. Cytogenetic causes in male infertility. Tartu, 2011.
- 181. **Delia Lepik**. Comparison of gunshot injuries caused from Tokarev, Makarov and Glock 19 pistols at different firing distances. Tartu, 2011.
- 182. **Ene-Renate Pähkla**. Factors related to the efficiency of treatment of advanced periodontitis. Tartu, 2011.

- 183. **Maarja Krass**. L-Arginine pathways and antidepressant action. Tartu, 2011.
- 184. **Taavi Lai**. Population health measures to support evidence-based health policy in Estonia. Tartu, 2011.
- 185. **Tiit Salum**. Similarity and difference of temperature-dependence of the brain sodium pump in normal, different neuropathological, and aberrant conditions and its possible reasons. Tartu, 2011.
- 186. **Tõnu Vooder**. Molecular differences and similarities between histological subtypes of non-small cell lung cancer. Tartu, 2011.
- 187. **Jelena Štšepetova**. The characterisation of intestinal lactic acid bacteria using bacteriological, biochemical and molecular approaches. Tartu, 2011.
- 188. **Radko Avi**. Natural polymorphisms and transmitted drug resistance in Estonian HIV-1 CRF06 cpx and its recombinant viruses. Tartu, 2011, 116 p.
- 189. **Edward Laane**. Multiparameter flow cytometry in haematological malignancies. Tartu, 2011, 152 p.
- 190. **Triin Jagomägi**. A study of the genetic etiology of nonsyndromic cleft lip and palate. Tartu, 2011, 158 p.
- 191. **Ivo Laidmäe**. Fibrin glue of fish (*Salmo salar*) origin: immunological study and development of new pharmaceutical preparation. Tartu, 2012, 150 p.
- 192. **Ülle Parm**. Early mucosal colonisation and its role in prediction of invasive infection in neonates at risk of early onset sepsis. Tartu, 2012, 168 p.
- 193. **Kaupo Teesalu**. Autoantibodies against desmin and transglutaminase 2 in celiac disease: diagnostic and functional significance. Tartu, 2012, 142 p.
- 194. **Maksim Zagura**. Biochemical, functional and structural profiling of arterial damage in atherosclerosis. Tartu, 2012, 162 p.
- 195. **Vivian Kont**. Autoimmune regulator: characterization of thymic gene regulation and promoter methylation. Tartu, 2012, 134 p.
- 196. **Pirje Hütt**. Functional properties, persistence, safety and efficacy of potential probiotic lactobacilli. Tartu, 2012, 246 p.
- 197. **Innar Tõru**. Serotonergic modulation of CCK-4- induced panic. Tartu, 2012, 132 p.
- 198. **Sigrid Vorobjov**. Drug use, related risk behaviour and harm reduction interventions utilization among injecting drug users in Estonia: implications for drug policy. Tartu, 2012, 120 p.
- 199. **Martin Serg**. Therapeutic aspects of central haemodynamics, arterial stiffness and oxidative stress in hypertension. Tartu, 2012, 156 p.
- 200. **Jaanika Kumm**. Molecular markers of articular tissues in early knee osteoarthritis: a population-based longitudinal study in middle-aged subjects. Tartu, 2012, 159 p.
- 201. **Kertu Rünkorg**. Functional changes of dopamine, endopioid and endocannabinoid systems in CCK2 receptor deficient mice. Tartu, 2012, 125 p.
- 202. **Mai Blöndal**. Changes in the baseline characteristics, management and outcomes of acute myocardial infarction in Estonia. Tartu, 2012, 127 p.

- 203. **Jana Lass**. Epidemiological and clinical aspects of medicines use in children in Estonia. Tartu, 2012, 170 p.
- 204. **Kai Truusalu**. Probiotic lactobacilli in experimental persistent *Salmonella* infection. Tartu, 2013, 139 p.
- 205. **Oksana Jagur**. Temporomandibular joint diagnostic imaging in relation to pain and bone characteristics. Long-term results of arthroscopic treatment. Tartu, 2013, 126 p.
- 206. **Katrin Sikk**. Manganese-ephedrone intoxication pathogenesis of neurological damage and clinical symptomatology. Tartu, 2013, 125 p.
- 207. **Kai Blöndal**. Tuberculosis in Estonia with special emphasis on drugresistant tuberculosis: Notification rate, disease recurrence and mortality. Tartu, 2013, 151 p.
- 208. **Marju Puurand**. Oxidative phosphorylation in different diseases of gastric mucosa. Tartu, 2013, 123 p.
- 209. **Aili Tagoma**. Immune activation in female infertility: Significance of autoantibodies and inflammatory mediators. Tartu, 2013, 135 p.
- 210. **Liis Sabre**. Epidemiology of traumatic spinal cord injury in Estonia. Brain activation in the acute phase of traumatic spinal cord injury. Tartu, 2013, 135 p.
- 211. **Merit Lamp**. Genetic susceptibility factors in endometriosis. Tartu, 2013, 125 p.
- 212. **Erik Salum**. Beneficial effects of vitamin D and angiotensin II receptor blocker on arterial damage. Tartu, 2013, 167 p.
- 213. **Maire Karelson**. Vitiligo: clinical aspects, quality of life and the role of melanocortin system in pathogenesis. Tartu, 2013, 153 p.
- 214. **Kuldar Kaljurand**. Prevalence of exfoliation syndrome in Estonia and its clinical significance. Tartu, 2013, 113 p.
- 215. **Raido Paasma**. Clinical study of methanol poisoning: handling large outbreaks, treatment with antidotes, and long-term outcomes. Tartu, 2013, 96 p.
- 216. **Anne Kleinberg**. Major depression in Estonia: prevalence, associated factors, and use of health services. Tartu, 2013, 129 p.
- 217. **Triin Eglit**. Obesity, impaired glucose regulation, metabolic syndrome and their associations with high-molecular-weight adiponectin levels. Tartu, 2014, 115 p.
- 218. **Kristo Ausmees**. Reproductive function in middle-aged males: Associations with prostate, lifestyle and couple infertility status. Tartu, 2014, 125 p.
- 219. **Kristi Huik**. The influence of host genetic factors on the susceptibility to HIV and HCV infections among intravenous drug users. Tartu, 2014, 144 p.
- 220. **Liina Tserel**. Epigenetic profiles of monocytes, monocyte-derived macrophages and dendritic cells. Tartu, 2014, 143 p.
- 221. **Irina Kerna**. The contribution of *ADAM12* and *CILP* genes to the development of knee osteoarthritis. Tartu, 2014, 152 p.

- 222. **Ingrid Liiv**. Autoimmune regulator protein interaction with DNA-dependent protein kinase and its role in apoptosis. Tartu, 2014, 143 p.
- 223. **Liivi Maddison**. Tissue perfusion and metabolism during intra-abdominal hypertension. Tartu, 2014, 103 p.
- 224. **Krista Ress**. Childhood coeliac disease in Estonia, prevalence in atopic dermatitis and immunological characterisation of coexistence. Tartu, 2014, 124 p.
- 225. **Kai Muru**. Prenatal screening strategies, long-term outcome of children with marked changes in maternal screening tests and the most common syndromic heart anomalies in Estonia. Tartu, 2014, 189 p.
- 226. **Kaja Rahu**. Morbidity and mortality among Baltic Chernobyl cleanup workers: a register-based cohort study. Tartu, 2014, 155 p.
- 227. **Klari Noormets**. The development of diabetes mellitus, fertility and energy metabolism disturbances in a Wfs1-deficient mouse model of Wolfram syndrome. Tartu, 2014, 132 p.
- 228. **Liis Toome**. Very low gestational age infants in Estonia. Tartu, 2014, 183 p.
- 229. **Ceith Nikkolo**. Impact of different mesh parameters on chronic pain and foreign body feeling after open inguinal hernia repair. Tartu, 2014, 132 p.
- 230. **Vadim Brjalin**. Chronic hepatitis C: predictors of treatment response in Estonian patients. Tartu, 2014, 122 p.
- 231. **Vahur Metsna**. Anterior knee pain in patients following total knee arthroplasty: the prevalence, correlation with patellar cartilage impairment and aspects of patellofemoral congruence. Tartu, 2014, 130 p.
- 232. **Marju Kase**. Glioblastoma multiforme: possibilities to improve treatment efficacy. Tartu, 2015, 137 p.
- 233. **Riina Runnel**. Oral health among elementary school children and the effects of polyol candies on the prevention of dental caries. Tartu, 2015, 112 p.
- 234. **Made Laanpere**. Factors influencing women's sexual health and reproductive choices in Estonia. Tartu, 2015, 176 p.
- 235. **Andres Lust**. Water mediated solid state transformations of a polymorphic drug effect on pharmaceutical product performance. Tartu, 2015, 134 p.
- 236. **Anna Klugman**. Functionality related characterization of pretreated wood lignin, cellulose and polyvinylpyrrolidone for pharmaceutical applications. Tartu, 2015, 156 p.
- 237. **Triin Laisk-Podar**. Genetic variation as a modulator of susceptibility to female infertility and a source for potential biomarkers. Tartu, 2015, 155 p.
- 238. **Mailis Tõnisson**. Clinical picture and biochemical changes in blood in children with acute alcohol intoxication. Tartu, 2015, 100 p.
- 239. **Kadri Tamme**. High volume haemodiafiltration in treatment of severe sepsis impact on pharmacokinetics of antibiotics and inflammatory response. Tartu, 2015, 133 p.

- 240. **Kai Part**. Sexual health of young people in Estonia in a social context: the role of school-based sexuality education and youth-friendly counseling services. Tartu, 2015, 203 p.
- 241. **Urve Paaver**. New perspectives for the amorphization and physical stabilization of poorly water-soluble drugs and understanding their dissolution behavior. Tartu, 2015, 139 p.
- 242. **Aleksandr Peet**. Intrauterine and postnatal growth in children with HLA-conferred susceptibility to type 1 diabetes. Tartu. 2015, 146 p.
- 243. **Piret Mitt**. Healthcare-associated infections in Estonia epidemiology and surveillance of bloodstream and surgical site infections. Tartu, 2015, 145 p.
- 244. **Merli Saare**. Molecular Profiling of Endometriotic Lesions and Endometria of Endometriosis Patients. Tartu, 2016, 129 p.
- 245. **Kaja-Triin Laisaar**. People living with HIV in Estonia: Engagement in medical care and methods of increasing adherence to antiretroviral therapy and safe sexual behavior. Tartu, 2016, 132 p.
- 246. **Eero Merilind**. Primary health care performance: impact of payment and practice-based characteristics. Tartu, 2016, 120 p.
- 247. **Jaanika Kärner**. Cytokine-specific autoantibodies in AIRE deficiency. Tartu, 2016, 182 p.
- 248. **Kaido Paapstel**. Metabolomic profile of arterial stiffness and early biomarkers of renal damage in atherosclerosis. Tartu, 2016, 173 p.
- 249. **Liidia Kiisk**. Long-term nutritional study: anthropometrical and clinicolaboratory assessments in renal replacement therapy patients after intensive nutritional counselling. Tartu, 2016, 207 p.
- 250. **Georgi Nellis**. The use of excipients in medicines administered to neonates in Europe. Tartu, 2017, 159 p.
- 251. **Aleksei Rakitin**. Metabolic effects of acute and chronic treatment with valproic acid in people with epilepsy. Tartu, 2017, 125 p.
- 252. **Eveli Kallas**. The influence of immunological markers to susceptibility to HIV, HBV, and HCV infections among persons who inject drugs. Tartu, 2017, 138 p.
- 253. **Tiina Freimann**. Musculoskeletal pain among nurses: prevalence, risk factors, and intervention. Tartu, 2017, 125 p.
- 254. **Evelyn Aaviksoo**. Sickness absence in Estonia: determinants and influence of the sick-pay cut reform. Tartu, 2017, 121 p.
- 255. **Kalev Nõupuu**. Autosomal-recessive Stargardt disease: phenotypic heterogeneity and genotype-phenotype associations. Tartu, 2017, 131 p.
- 256. Ho Duy Binh. Osteogenesis imperfecta in Vietnam. Tartu, 2017, 125 p.
- 257. **Uku Haljasorg**. Transcriptional mechanisms in thymic central tolerance. Tartu, 2017, 147 p.
- 258. **Živile Riispere**. IgA Nephropathy study according to the Oxford Classification: IgA Nephropathy clinical-morphological correlations, disease progression and the effect of renoprotective therapy. Tartu, 2017, 129 p.

- 259. **Hiie Soeorg**. Coagulase-negative staphylococci in gut of preterm neonates and in breast milk of their mothers. Tartu, 2017, 216 p.
- 260. **Anne-Mari Anton Willmore**. Silver nanoparticles for cancer research. Tartu, 2017, 132 p.
- 261. **Ott Laius**. Utilization of osteoporosis medicines, medication adherence and the trend in osteoporosis related hip fractures in Estonia. Tartu, 2017, 134 p.
- 262. **Alar Aab**. Insights into molecular mechanisms of asthma and atopic dermatitis. Tartu, 2017, 164 p.
- 263. **Sander Pajusalu**. Genome-wide diagnostics of Mendelian disorders: from chromosomal microarrays to next-generation sequencing. Tartu, 2017, 146 p.
- 264. **Mikk Jürisson**. Health and economic impact of hip fracture in Estonia. Tartu, 2017, 164 p.
- 265. **Kaspar Tootsi**. Cardiovascular and metabolomic profiling of osteoarthritis. Tartu, 2017, 150 p.
- 266. **Mario Saare**. The influence of AIRE on gene expression studies of transcriptional regulatory mechanisms in cell culture systems. Tartu, 2017, 172 p.
- 267. **Piia Jõgi**. Epidemiological and clinical characteristics of pertussis in Estonia. Tartu, 2018, 168 p.
- 268. **Elle Põldoja**. Structure and blood supply of the superior part of the shoulder joint capsule. Tartu, 2018, 116 p.
- 269. **Minh Son Nguyen**. Oral health status and prevalence of temporomandibular disorders in 65–74-year-olds in Vietnam. Tartu, 2018, 182 p.
- 270. **Kristian Semjonov**. Development of pharmaceutical quench-cooled molten and melt-electrospun solid dispersions for poorly water-soluble indomethacin. Tartu, 2018, 125 p.
- 271. **Janne Tiigimäe-Saar**. Botulinum neurotoxin type A treatment for sialorrhea in central nervous system diseases. Tartu, 2018, 109 p.
- 272. **Veiko Vengerfeldt**. Apical periodontitis: prevalence and etiopathogenetic aspects. Tartu, 2018, 150 p.
- 273. **Rudolf Bichele**. TNF superfamily and AIRE at the crossroads of thymic differentiation and host protection against *Candida albicans* infection. Tartu, 2018, 153 p.
- 274. **Olga Tšuiko**. Unravelling Chromosomal Instability in Mammalian Preimplantation Embryos Using Single-Cell Genomics. Tartu, 2018, 169 p.
- 275. **Kärt Kriisa**. Profile of acylcarnitines, inflammation and oxidative stress in first-episode psychosis before and after antipsychotic treatment. Tartu, 2018, 145 p.
- 276. **Xuan Dung Ho**. Characterization of the genomic profile of osteosarcoma. Tartu, 2018, 144 p.
- 277. **Karit Reinson**. New Diagnostic Methods for Early Detection of Inborn Errors of Metabolism in Estonia. Tartu, 2018, 201 p.

- 278. **Mari-Anne Vals**. Congenital N-glycosylation Disorders in Estonia. Tartu, 2019, 148 p.
- 279. **Liis Kadastik-Eerme**. Parkinson's disease in Estonia: epidemiology, quality of life, clinical characteristics and pharmacotherapy. Tartu, 2019, 202 p.
- 280. **Hedi Hunt**. Precision targeting of intraperitoneal tumors with peptideguided nanocarriers. Tartu, 2019, 179 p.
- 281. **Rando Porosk**. The role of oxidative stress in Wolfram syndrome 1 and hypothermia. Tartu, 2019, 123 p.
- 282. **Ene-Ly Jõgeda**. The influence of coinfections and host genetic factor on the susceptibility to HIV infection among people who inject drugs. Tartu, 2019, 126 p.
- 283. **Kristel Ehala-Aleksejev**. The associations between body composition, obesity and obesity-related health and lifestyle conditions with male reproductive function. Tartu, 2019, 138 p.
- 284. **Aigar Ottas**. The metabolomic profiling of psoriasis, atopic dermatitis and atherosclerosis. Tartu, 2019, 136 p.
- 285. **Elmira Gurbanova**. Specific characteristics of tuberculosis in low default, but high multidrug–resistance prison setting. Tartu, 2019, 129 p.
- 286. **Van Thai Nguyeni**. The first study of the treatment outcomes of patients with cleft lip and palate in Central Vietnam. Tartu, 2019, 144 p.
- 287. Maria Yakoreva. Imprinting Disorders in Estonia. Tartu, 2019, 187 p.
- 288. **Kadri Rekker**. The putative role of microRNAs in endometriosis pathogenesis and potential in diagnostics. Tartu, 2019, 140 p.
- 289. Ülle Võhma. Association between personality traits, clinical characteristics and pharmacological treatment response in panic disorder. Tartu, 2019, 121 p.
- 290. **Aet Saar**. Acute myocardial infarction in Estonia 2001–2014: towards risk-based prevention and management. Tartu, 2019, 124 p.
- 291. **Toomas Toomsoo**. Transcranial brain sonography in the Estonian cohort of Parkinson's disease. Tartu, 2019, 114 p.
- 292. **Lidiia Zhytnik**. Inter- and intrafamilial diversity based on genotype and phenotype correlations of Osteogenesis Imperfecta. Tartu, 2019, 224 p.
- 293. **Pilleriin Soodla**. Newly HIV-infected people in Estonia: estimation of incidence and transmitted drug resistance. Tartu, 2019, 194 p.
- 294. **Kristiina Ojamaa**. Epidemiology of gynecological cancer in Estonia. Tartu, 2020, 133 p.
- 295. **Marianne Saard**. Modern Cognitive and Social Intervention Techniques in Paediatric Neurorehabilitation for Children with Acquired Brain Injury. Tartu, 2020, 168 p.
- 296. **Julia Maslovskaja**. The importance of DNA binding and DNA breaks for AIRE-mediated transcriptional activation. Tartu, 2020, 162 p.
- 297. **Natalia Lobanovskaya**. The role of PSA-NCAM in the survival of retinal ganglion cells. Tartu, 2020, 105 p.

- 298. **Madis Rahu**. Structure and blood supply of the postero-superior part of the shoulder joint capsule with implementation of surgical treatment after anterior traumatic dislocation. Tartu, 2020, 104 p.
- 299. **Helen Zirnask**. Luteinizing hormone (LH) receptor expression in the penis and its possible role in pathogenesis of erectile disturbances. Tartu, 2020, 87 p.
- 300. **Kadri Toome**. Homing peptides for targeting of brain diseases. Tartu, 2020, 152 p.
- 301. **Maarja Hallik**. Pharmacokinetics and pharmacodynamics of inotropic drugs in neonates. Tartu, 2020, 172 p.
- 302. **Raili Müller**. Cardiometabolic risk profile and body composition in early rheumatoid arthritis. Tartu, 2020, 133 p.
- 303. **Sergo Kasvandik**. The role of proteomic changes in endometrial cells from the perspective of fertility and endometriosis. Tartu, 2020, 191 p.
- 304. **Epp Kaleviste**. Genetic variants revealing the role of STAT1/STAT3 signaling cytokines in immune protection and pathology. Tartu, 2020, 189 p.
- 305. Sten Saar. Epidemiology of severe injuries in Estonia. Tartu, 2020, 104 p.
- 306. **Kati Braschinsky**. Epidemiology of primary headaches in Estonia and applicability of web-based solutions in headache epidemiology research. Tartu, 2020, 129 p.
- 307. **Helen Vaher**. MicroRNAs in the regulation of keratinocyte responses in *psoriasis vulgaris* and atopic dermatitis. Tartu, 2020, 242 p.
- 308. **Liisi Raam**. Molecular Alterations in the Pathogenesis of Two Chronic Dermatoses Vitiligo and Psoriasis. Tartu, 2020, 164 p.
- 309. **Artur Vetkas**. Long-term quality of life, emotional health, and associated factors in patients after aneurysmal subarachnoid haemorrhage. Tartu, 2020, 127 p.
- 310. **Teele Kasepalu**. Effects of remote ischaemic preconditioning on organ damage and acylcarnitines' metabolism in vascular surgery. Tartu, 2020, 130 p.
- 311. **Prakash Lingasamy**. Development of multitargeted tumor penetrating peptides. Tartu, 2020, 246 p.
- 312. **Lille Kurvits.** Parkinson's disease as a multisystem disorder: whole transcriptome study in Parkinson's disease patients' skin and blood. Tartu, 2021, 142 p.
- 313. **Mariliis Põld.** Smoking, attitudes towards smoking behaviour, and nicotine dependence among physicians in Estonia: cross-sectional surveys 1982–2014. Tartu, 2021, 172 p.
- 314. **Triin Kikas**. Single nucleotide variants affecting placental gene expression and pregnancy outcome. Tartu, 2021, 160 p.
- 315. **Hedda Lippus-Metsaots**. Interpersonal violence in Estonia: prevalence, impact on health and health behaviour. Tartu, 2021, 172 p.

- 316. **Georgi Dzaparidze.** Quantification and evaluation of the diagnostic significance of adenocarcinoma-associated microenvironmental changes in the prostate using modern digital pathology solutions. Tartu, 2021, 132 p.
- 317. **Tuuli Sedman.** New avenues for GLP1 receptor agonists in the treatment of diabetes. Tartu, 2021, 118 p.
- 318. **Martin Padar.** Enteral nutrition, gastrointestinal dysfunction and intestinal biomarkers in critically ill patients. Tartu, 2021, 189 p.
- 319. **Siim Schneider.** Risk factors, etiology and long-term outcome in young ischemic stroke patients in Estonia. Tartu, 2021, 131 p.
- 320. **Konstantin Ridnõi.** Implementation and effectiveness of new prenatal diagnostic strategies in Estonia. Tartu, 2021, 191 p.
- 321. **Risto Vaikjärv.** Etiopathogenetic and clinical aspects of peritonsillar abscess. Tartu, 2021, 115 p.
- 322. **Liis Preem.** Design and characterization of antibacterial electrospun drug delivery systems for wound infections. Tartu, 2022, 220 p.
- 323. **Keerthie Dissanayake.** Preimplantation embryo-derived extracellular vesicles: potential as an embryo quality marker and their role during the embryo-maternal communication. Tartu, 2022, 203 p.
- 324. **Laura Viidik.** 3D printing in pharmaceutics: a new avenue for fabricating therapeutic drug delivery systems. Tartu, 2022, 139 p.
- 325. **Kasun Godakumara.** Extracellular vesicle mediated embryo-maternal communication A tool for evaluating functional competency of preimplantation embryos. Tartu, 2022, 176 p.
- 326. **Hindrek Teder.** Developing computational methods and workflows for targeted and whole-genome sequencing based non-invasive prenatal testing. Tartu, 2022, 138 p.
- 327. **Jana Tuusov.** Deaths caused by alcohol, psychotropic and other substances in Estonia: evidence based on forensic autopsies. Tartu, 2022, 157 p.
- 328. **Heigo Reima.** Colorectal cancer care and outcomes evaluation and possibilities for improvement in Estonia. Tartu, 2022, 140 p.