

MATHILDE FRÉDÉRIQUE E. ANDRÉ

New Guinea, a hotspot for Human  
evolution: settlement history  
and adaptation in northern Sahul



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Institute of Genomics and the Institute of Molecular and Cell Biology, University of Tartu, Estonia

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## LIST OF ORIGINAL PUBLICATIONS

- I. Nicolas Brucato, **Mathilde André**, Roxanne Tsang, Lauri Saag, Jason Kariwiga, Kylie Sesuki, Teppsy Beni, William Pomat, John Muke, Vincent Meyer, Anne Boland, Jean-François Deleuze, Herawati Sudoyo, Mayukh Mondal, Luca Pagani, Irene Gallego Romero, Mait Metspalu, Murray P Cox, Matthew Leavesley, François-Xavier Ricaut, Papua New Guinean Genomes Reveal the Complex Settlement of North Sahul, *Molecular Biology and Evolution*, Volume 38, Issue 11, October 2021, <https://doi.org/10.1093/molbev/msab238>
- II. Nicolas Brucato, **Mathilde André**, Georgi Hudjashov, Mayukh Mondal, Murray P.Cox, Matthew Leavesley, François-Xavier Ricaut, Chronology of natural selection in Oceanian genomes, *IScience*, Volume 25, Issue 7, June 2022, <https://doi.org/10.1016/j.isci.2022.104583>
- III. **Mathilde André**, Nicolas Brucato, Sébastien Plutniak, Jason Kariwiga, John Muke, Adeline Morez, Matthew Leavesley, Mayukh Mondal, François-Xavier Ricaut, Phenotypic differences between highlanders and lowlanders in Papua New Guinea, *PLOS ONE*, Volume 16, Issue 7, July 2021, <https://doi.org/10.1371/journal.pone.0253921>
- IV. **Mathilde André**, Nicolas Brucato, Georgi Hudjasov, Vasili Pankratov, Danat Yermakovich, Rita Kreevan, Jason Kariwiga, John Muke, Anne Boland, Jean-François Deleuze, Vincent Meyer, Nicholas Evans, Murray P. Cox, Matthew Leavesley, Michael Dannemann, Tõnis Org, Mait Metspalu, Mayukh Mondal, François-Xavier Ricaut, Positive selection in the genomes of two Papua New Guinean populations at distinct altitude levels, *bioRxiv*, [preprint], December 2022, <https://doi.org/10.1101/2022.12.15.520226>

## **AUTHOR'S CONTRIBUTION TO THE LISTED ARTICLES**

The author's contributions to the listed articles are as follows:

- Ref I:** I participated in the interpretation of the results and provided input for writing the paper.
- Ref II:** I participated in the selection dating analysis, the interpretation of the results and provided input for writing the paper..
- Ref III:** I curated and analysed phenotype measurements, generated figures, interpreted the results and led the manuscript writing team.
- Ref IV:** I generated selection scans, reconstructed allele frequencies, analysed association studies data and investigated candidate haplotypes for evidence of archaic introgression. In addition, I generated figures, interpreted the results and led the manuscript writing team.

## ABBREVIATIONS

<b>a.s.l.</b>	above sea level
<b>AMH</b>	anatomically modern human
<b>ARG</b>	ancestral recombination graph
<b>BMI</b>	body mass index
<b>bp</b>	base pair
<b>CI</b>	confidence interval
<b>cM</b>	centiMorgan
<b>CMS</b>	chronic mountain sickness
<b>DAF</b>	derived allele frequency
<b>DARC</b>	Duffy antigen receptor for chemokines
<b>EHH</b>	Extended Haplotype Homozygosity
<b>FEV1</b>	forced expiratory volume in 1 second
<b>F<sub>ST</sub></b>	fixation index
<b>FVC</b>	forced vital capacity
<b>GBP</b>	guanylate-binding protein
<b>GEMMA</b>	Genome-wide Efficient Mixed Model Association
<b>GEVA</b>	Genealogical Estimation of Variant Age
<b>GTE<sub>x</sub></b>	Genotype-Tissue Expression
<b>GWAS</b>	genome-wide association studies
<b>HIF</b>	Hypoxia-Inducible Factor
<b>HLA</b>	Human Leukocyte Antigen
<b>HMM</b>	Hidden Markov Model
<b>iHS</b>	integrated haplotype score
<b>IQR</b>	interquartile range
<b>kya</b>	thousand of years ago
<b>LD</b>	linkage disequilibrium
<b>LMM</b>	Linear Mixed Model
<b>LS</b>	Li and Stephen
<b>MCMC</b>	Markov Chain Monte Carlo
<b>MHC</b>	Major Histocompatibility Complex
<b>MRCA</b>	most recent common ancestor
<b>nSL</b>	number of segregating sites by length
<b>PBS</b>	Population Branch Statistic
<b>PCA</b>	Principal Component Analysis

<b>PEF</b>	peak expiratory flow
<b>PNG</b>	Papua New Guinea(n)
<b>SGDP</b>	Simons Genome Diversity Project
<b>SNP</b>	single nucleotide polymorphism
<b>TMRC</b>	time to the most recent common ancestor
<b>VEP</b>	Ensembl Variant Effect Predictor
<b>WHO</b>	World Health Organization
<b>XP-EHH</b>	cross-population extended haplotype homozygosity
<b>XP-nSL</b>	cross-population number of segregating sites by length
<b>YBP</b>	Years Before Present

# 1. INTRODUCTION

## 1.1. New Guinea: exploring the multi-layered diversity

New Guinea is the world's second-largest island. It is separated from Australia by the Arafura Sea and is shared between two countries. Indonesia governs the western half of the island, and Papua New Guinea (PNG), an independent country since 1975, constitutes the Eastern half of the island. The island of New Guinea has around 14 million inhabitants, with 4 million living in the Indonesian provinces in western Papua and 10 million in PNG. New Guineans and Papua New Guineans are used consistently throughout this thesis to name the inhabitants of the island of New Guinea or the country of Papua New Guinea, respectively. The term "Papuan" is exclusively used to describe the ancestry of the indigenous people of New Guinea, in contrast with the Austronesian ancestry or other ancestries brought to New Guinea by recent migrations.

New Guinea is known for its early settlement by anatomically modern humans (AMH) and unique bio-cultural diversity. After AMH left Africa around 100 thousand years ago (kya), they arrived in the ancient continent of Sahul, which included Australia, Tasmania and the current island of New Guinea, as early as 50 kya (O'Connell et al., 2018). The journey to Sahul must have been challenging as AMH faced various unfamiliar environments. After the initial settlement, New Guinea remained relatively isolated from outside influence until the Austronesian expansion 3 kya. Communication between the different regions of the island was limited for most of its history. This relative isolation has led to the unique cultural and biological diversity that arose in New Guinea over time. New Guinea's bio-cultural diversity makes it of great interest to many disciplines. For instance, New Guinea is the most linguistically diverse territory in the world, with more than 800 spoken languages (Palmer, 2017), making it a major area of study for linguistics. Alongside their linguistic diversity, New Guinean groups also exhibit strong genetic variation from one another (Bergström et al., 2017). New Guinean genomes stand out even more by harbouring one of the highest amounts of Denisovan ancestry in their genomes (Larena et al., 2021; Reich et al., 2010).

New Guinean territory is characterised by diverse environments, from the coastal lowlands to the highlands, which host some of the highest mountains in Oceania (4,884 meters above sea level (a.s.l.)). New Guinean highlanders and lowlanders followed different cultural trajectories for thousands of years. New Guinea highland territories located between 1,600 and 2,400 m a.s. l. have been permanently inhabited for 20 kya and are currently the most densely populated area of the country (Brookfield & Allen, 1989; Müller, Bockarie, Alpers, & Smith, 2003). The partial pressure of inspired oxygen decreases with increasing altitude, which results in hypoxia (i.e., reduced oxygen availability to the body). Various detrimental conditions, such as acute mountain sickness, can occur after exposure to hypoxia from elevation as low as 1500 m a.s.l. (Barry & Pollard, 2003). Hence,

New Guinean highlanders have been experiencing hypoxia and its detrimental effects for thousands of years. Some populations, like the Andean Calchaquíes who have lived at intermediate altitude (above 1,500 m a.s.l.), and the Tibetan population who have lived at high-altitude (above 2,500 m a.s.l.) for thousands of years, have adapted to hypoxia and carry genomic signatures of selection to this stressor (Cynthia M. Beall, 2014; Eichstaedt et al., 2015). In contrast, the New Guinean highlands, despite being the most densely populated areas of the island since the Holocene (11 kya) (Brookfield & Allen, 1989; Müller et al., 2003; Trájer, Sebestyén, & Domokos, 2020), have been overlooked in terms of adaptation to hypoxia as these other populations.

Pathogenic pressure is another strong environmental challenge in New Guinea, with pathogens infection being the leading cause of death among people of Papua New Guinea (PNGs) (GBD 2013 Mortality and Causes of Death Collaborators, 2015; Kitur, Adair, Riley, & Lopez, 2019; Naraq, Feling, & Leeder, 2003). Differences in pathogenic pressure between lowlanders and highlanders could explain why New Guinean highlands are more densely populated (Riley, 1983; Trájer, 2022; Trájer et al., 2020). For example, Malaria is highly prevalent in the lowlands while being rare in the highlands (Müller et al., 2003; Senn et al., 2010).

Population genetics has come a long way since the pioneering work of Cavalli-Sforza and colleagues, who provided the first insights into human populations' genetic diversity and structure (Cavalli-Sforza, Piazza, Menozzi, & Mountain, 1988). With the advent of affordable whole genome sequencing technologies, we have now entered the era of population genomics (Jobling, Hollox, Hurles, Kivisild, & Tyler-Smith, 2013; Pareek, Smoczynski, & Tretyn, 2011). The effort in large-scale sequencing of multiple populations worldwide (e.g., 1000 Genomes Project (The 1000 Genomes Project Consortium et al., 2015), Simons Genome Diversity Project (SGDP) (Mallick et al., 2016), Estonian Biocentre Human Genome Diversity Panel (Pagani et al., 2016) ) has further improved our understanding of human genetic diversity. New sequencing tools have also enabled the generation of genomes from ancient human populations (Orlando et al., 2021) and even from other hominin species like Neanderthal (Green et al., 2010; Prüfer et al., 2014) and Denisova (Reich et al., 2010). In the case of the Denisovans, there were so few remains that their existence was identified only through the analysis of ancient DNA (Krause et al., 2010). Together advancements in modern and ancient genomics have widened our understanding of the evolution of human genome diversity, both spatially and temporally. Furthermore, the recent recognition of Svante Pääbo's work with the 2022 Nobel Prize in Physiology or Medicine has brought worldwide attention to evolutionary genomics, highlighting its potential to illuminate the interplay between evolutionary history and human health ("The Nobel Prize in Physiology or Medicine 2022", 2022). Evolutionary genomics remains a hot topic that promises to advance our understanding of the biological mechanisms underlying human evolution and health.

Whole genome sequencing is particularly valuable for studying New Guineans' unique genetic diversity. Indeed, currently available single nucleotide polymorphism (SNP) array designs are missing important genetic components exclusively

found in New Guineans and then only provide a partial and biased picture of their genetic diversity. By analyzing whole genomes, we can better understand the evolutionary forces that have shaped this diversity and uncover unique signatures of selection that may be relevant to human health and disease. This thesis presents new whole genome sequences from PNG highlands and lowlands as well as measurements of diverse phenotypes in these two populations. The first reference aimed to describe the models for northern Sahul settlement based on genomic data (Ref I). The second reference investigated signatures of selection unique to New Guineans witnessing the selection pressure their ancestors faced immediately after settling in new environments when reaching Sahul (Ref II). The last two references explored phenotype differences (Ref III) and selection signatures (Ref IV) unique to PNG highlanders and lowlanders to detect if PNG populations might display signals of local adaptation to their specific environment.

## **2. LITERATURE OVERVIEW**

New Guinea is known for its early settlement by AMH and unique bio-cultural diversity. New Guineans are among the most genetically diverse populations and have lived in challenging environments for tens of thousands of years. Furthermore, their genomes harbour some of the highest levels of Denisovan DNA, an extinct hominin species. For all these reasons, New Guineans are a major interest in human evolution and adaptation studies.

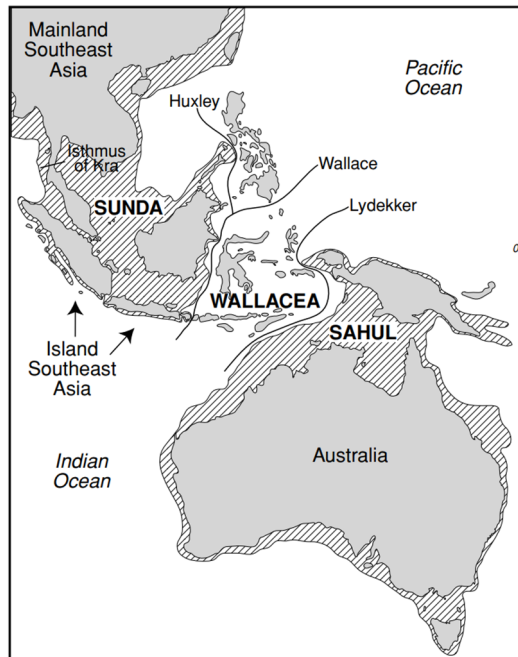
The following literature review provides an overview of the complex settlement of New Guinea and the different migratory waves that participated in the diversity of New Guineans. It also describes the key concepts in studying selection in human genomes and the methods used to detect and fine-map selection.

### **2.1. New Guinea: a unique settlement history and diversity**

#### **2.1.1. Crossing the sea: early human migration to New Guinea**

AMH appeared in Africa around 300 kya (Hublin et al., 2017), where they remained confined until 70 kya (Malaspinas et al., 2016; Mallick et al., 2016; Mondal et al., 2016). However, the time AMH expanded out of Africa remains debated, as some studies suggest an early out-of-Africa event around 100 kya (Grove et al., 2015; Kuhlwilm et al., 2016; Pagani et al., 2016).

When AMH left Africa, they began to spread across Eurasia and Southwest Asia. At this time, the sea level was lower, so some territories now separated by the sea were then part of the same landmass. There were two main continents in the South Pacific region: Sunda, which connected many current southeast Asian islands and mainland Asia; and Sahul, which included mainland Australia, Tasmania, and New Guinea. These two ancient continents were separated by Wallacea, many small islands separated by deep-water crossings that limited migrations between Sahul and Sunda (Wallace 1869) and created two distinct biogeographic areas (Figure 1) (Cámara-Leret et al., 2020). Despite this complex landscape, archaeological artefacts, ecological models and mitochondrial DNA indicate that the first settlers reached northern Sahul between 50 and 65 kya (Bradshaw et al., 2021; Clarkson et al., 2017; O'Connell & Allen, 2015; Pedro et al., 2020; Summerhayes et al., 2010). It has been suggested that New Guineans may even carry genomic traces from an early expansion of AMH out of Africa (Pagani et al., 2016). Nevertheless, the exact way how AMH crossed Wallacea and reached Sahul remains a topic of ongoing research and debate among scientists (Allen & O'Connell, 2020). Our current understanding relies on migration models that use paleogeological reconstructions of coastlines, environmental information or archaeological artefacts.



**Figure 1: Map of Australasia showing the extent of the Sunda and Sahul continental landmasses with Wallacea in between.** The contemporary geography of the region is shown in grey. Reprinted from Figure 1 (Harrison, Krigbaum, & Manser, 2006), with permission from Springer Nature. Copyright © 2006, Springer Science Business Media, LLC

#### 2.1.1.1. An accident or a deliberate effort?

There is debate over whether the crossing from Sunda to Sahul was intentional or accidental. Some studies have suggested that early humans accidentally reached Sahul using natural rafts of vegetation following currents (Bird et al., 2019). However, with a minimal founding population size of around 1,300 individuals, the accident hypothesis is unlikely (Allen & O’Connell, 2020; Bird et al., 2019; Bradshaw et al., 2019). The substantial size of the founding groups also suggests that they had cohesion and cognitive and technological abilities to plan a journey, supporting the idea of an intentional and directed migration (Bradshaw et al., 2019).

#### 2.1.1.2. The northern vs the southern route

Because Sunda and Sahul have never been connected by land, AMH must have used seafaring from one Wallacean island to another to reach Sahul. Two pathways were possible: the northern route through Sulawesi into New Guinea and the southern route through Bali and Timor into northwestern Australia (Smith, 2001). Different modelling approaches, using crossing distance as a proxy for crossing difficulty (Kealy, Louys, & O’Connor, 2018) and coastal viewshed analysis (i.e., identifying locations that are visible from the observer’s point of

view) (Bird et al., 2019; Norman et al., 2018), suggest that the northern path was the easiest and quickest route, while the southern path was longer and required travelling without having the destination in sight (Bird et al., 2019). However, older archaeological sites have been found along the southern route through the Arafura plain (around 46 kya) (Hawkins et al., 2017; Sutikna et al., 2018) (or even earlier, 65 kya in northern Australia (Clarkson et al., 2017)) than on the northern route (around 35 kya) (Latinis & Stark, 2005; O'Connor, Louys, Kealy, & Samper Carro, 2017). Nevertheless, these hypotheses cannot rely solely on archaeological evidence as the sea level has risen substantially since the first settlement, and many early sites may now be underwater (Summerhayes, Field, Shaw, & Gaffney, 2017). Finding older sites on the south path when models favour the northern arrival might suggest that first settlers rapidly spread along the coastal areas or a lack of archaeological investigation in western New Guinea (Bird et al., 2019).

Because the journey was intentional and repeatedly taken by several groups, it is also likely that the different groups used different routes during their trip to Sahul (Bird et al., 2019; Bradshaw et al., 2019). More recent models support multiple entry points, with an initial entry through the southern road into the north of modern Australia and a latter entrance via the northern route into New Guinea (Bradshaw et al., 2021). These groups of settlers may have already been genetically distinct (Bird et al., 2019). Indeed, the arrival of a unique human group is unlikely to explain the deep genetic diversity still observed in this region (Allen & O'Connell, 2020). However, though most of the studies on the Indigenous Australian and New Guinean Y chromosomes suggest a split around the time of the initial settlement of Sahul (Bergström et al., 2016; Tobler et al., 2017), there is also a suggestion of more recent connections of paternal lineages from these regions (Karmin et al., 2022).

## **2.1.2. Settlement within mainland New Guinea**

Since its settlement by AMH 50 kya (O'Connell et al., 2018; Summerhayes et al., 2010), New Guinea has remained relatively isolated from outside influences (Terrell, 2004). This isolation, coupled with the diverse landscape found in New Guinea (Pawley & Australian National University, 2005; Summerhayes et al., 2017), has resulted in exceptional cultural and genetic diversity on the island.

### **2.1.2.1. Facing multiple new environments**

The initial settlers of northern Sahul spread across the continent within a few thousand years (Bergström et al., 2016), encountering a diverse range of environments, including savannah grasslands, tropical rainforests, and montane landscapes.

After they entered Sahul, the first settlers rapidly accessed the mountain valleys. Indeed, one of the oldest sites in northern Sahul is located around 2000 m a.s.l. (Summerhayes et al., 2010). Coastal small hunter-gatherer groups were probably

penetrating the higher valleys as temporary foraging camps to access different resources than in the lowland. By the late Pleistocene (around 25kya), some groups shifted to a semi-permanent settlement in the highlands, probably thanks to the ameliorating climate (Gaffney, Ford, & Summerhayes, 2015). New Guinean highlanders have been residing in the highlands for at least 20,000 years (Haas et al., 2017; Madsen et al., 2017). The speed of their establishment suggests a rapid adaptation to the local flora and fauna for sustenance (Summerhayes et al., 2017).

Currently, the PNG highlands are the most densely populated area of the country, comprising approximately 3 million individuals, nearly 40% of the total population (Bourke & Allen, 2021). Many highlander groups have inhabited elevations between 1,600 and 2,400 m a.s.l. since the Holocene (Brookfield & Allen, 1989; Müller et al., 2003; Trájer et al., 2020). One reason for the uneven population distribution might be that intense pathogenic pressure in New Guinean lowlands that prompted the first inhabitant of New Guinea to access the highlands (Müller et al., 2003; Riley, 1983; Trájer, 2022; Trájer et al., 2020).

Since the late Pleistocene, the limited interaction between highland and lowland populations has facilitated the development of specific regional cultures. For instance, local plant cultivation practices appeared independently in the highlands around 10–7 kya (Denham, 2003; Golson, 2017) and have been associated with the demographic growth in that region (Bergström et al., 2017). In contrast with the highlands, the population size in the lowlands remained stable during the Holocene (Bergström et al., 2017), and agricultural practices were brought to the lowlands through the Austronesian influence around 3 kya (Bellwood et al., 2007; Skoglund et al., 2016).

#### 2.1.2.2. From environment diversity to genetic diversity

During the initial settlement of Sahul, Indigenous Australians and New Guineans were probably not yet distinct populations and may have only diverged around 30 kya (Bergström et al., 2016; Karmin et al., 2022; Malaspinas et al., 2016; Tobler et al., 2017). After the rapid peopling of Sahul, New Guineans remained genetically isolated from other populations leading to their current unique genetic makeup (Bergström et al., 2020, 2016; Malaspinas et al., 2016). New Guinean populations also show a striking genetic heterogeneity among themselves (Bergström et al., 2020). The genetic differentiation between New Guinean groups, as measured by the fixation index ( $F_{ST}$ ) between groups, is bigger than that of European or Asian groups separated by equivalent geographic distances. The high differentiation between New Guinean groups would support a limited dispersal and admixture of New Guinean populations over time (Bergström et al., 2017).

The first split between New Guinean populations is between New Guinea mainland inhabitants and Bismarck and Solomon islanders, shortly after northern Sahul's initial settlement (Bergström et al., 2017; Choin et al., 2021). The most diverging groups inside mainland New Guinea are lowlanders and highlanders. This genetic structure mostly lies in the isolation of these two groups during the Late Pleistocene, which also resulted in the different cultural practices mentioned above.

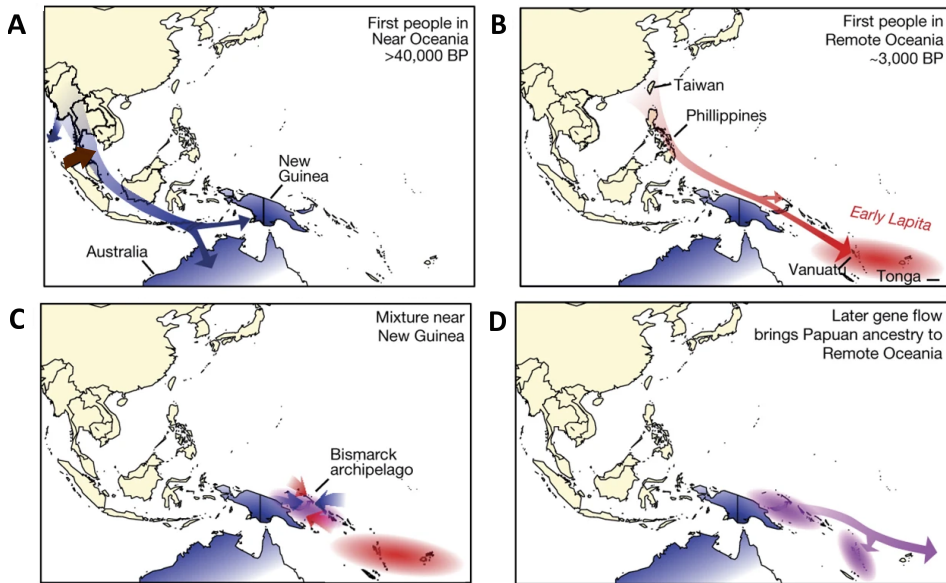
### 2.1.3. The Austronesian expansion

During the mid-Holocene (around 3 kya), Austronesians from Southeast Asia mixed with some Papuan populations, adding another layer of complexity to the New Guinean genetic diversity.

The Austronesian expansion, associated with the Lapita culture and Austronesian languages, started 5 kya from present-day Taiwan and reached the last inhabited territories in the Pacific Ocean (Bellwood et al., 2007; Patrick Vinton Kirch, 2002). This migration wave resulted in the modern Pacific population having a mixed Papuan and Austronesian ancestry (Kayser, 2010; Lipson et al., 2018).

The first wave of Austronesian-speakers from Taiwan to remote Oceania left minimal admixture trace within the inhabitants of Papua New Guinea (Figure 2B) (Bellwood et al., 2007; Hung & Carson, 2014; Skoglund et al., 2016). Another episode of gene flow with Papuans occurred around 3000 years ago in the Bismarck archipelago where the Austronesian-related ancestry was carried by the first settlers of Remote Oceania (P. V. Kirch & Hunt, 1988; Peter Bellwood & Eusebio Dizon, 2005; Skoglund et al., 2016) (Figure 2C). The secondary expansion to remote Oceania 2.3 to 2.9 kya included Papuan-Austronesian admixed populations (Lipson et al., 2018; Skoglund et al., 2016) (Figure 2D).

As a result of Austronesian admixture, some lowlanders in coastal New Guinea exhibit varying degrees of Southeast Asian genetic markers. Austronesian admixture in PNG is particularly important among Austronesian-speaking populations compared to non-Austronesian speakers (Bergström et al., 2017).



**Figure 2:** **A**, A model of population movements more than 40,000 years ago in which modern humans arrived in the Australia–New Guinea region (blue shading) and mixed with archaic Denisovans (brown arrow). **B**, A model of events before 3,000 years ago, in which the First Remote Oceanian population formed by the spread of a population of ultimate East Asian origin to a region including Vanuatu and Tonga, and experienced little or no mixture with the Papuans they encountered along the journey (red shading). Note that geographic routes are speculative. **C**, A model of populations of mixed Papuan–First Remote Oceanian ancestry in Near Oceania less than 3,000 years ago in a patchwork of islands with different proportions of First Remote Oceanian ancestry (pink shading). **D**, A model of secondary expansion of admixed populations bringing Papuan ancestry into Remote Oceania, which was still not complete in Tonga by the date of the Talasiu individual at 2,680–2,340 BP. Modified from Figure 3 (Skoglund et al., 2016) with permission from Springer Nature. Copyright © 2016, Macmillan Publishers Limited, part of Springer Nature. All rights reserved.

### 2.1.4. Interbreeding with archaic hominins

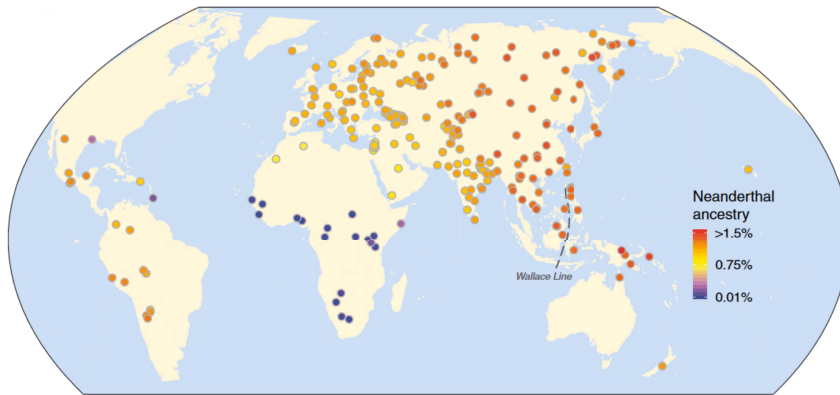
While AMH were still constrained to Africa, other hominin populations, such as Neanderthal and Denisova, were already present in Eurasia and East and South Asia, respectively. Both of these archaic hominins diverged from AMH around 520 kya and from each other around 390 kya (Meyer et al., 2016; Prüfer et al., 2017). When AMH left Africa, they met and admixed with other hominin species like Neanderthal (Green et al., 2010; Prüfer et al., 2014), Denisova (Reich et al., 2010) and potentially still unknown other archaic populations (Hammer, Woerner, Mendez, Watkins, & Wall, 2011; Mondal, Bertranpetit, & Lao, 2019; Mondal et al., 2016) (Figure 3). The remains of these admixture events can still be found in the genomes of current populations, including New Guineans. The admixture could have played a role in accelerating AMH adaptation to environments new to them but already inhabited by these archaic hominins for thousands of years (Dannemann & Kelso, 2017; Reilly, Tjahjadi, Miller, Akey, & Tucci, 2022; Sankararaman, Mallick, Patterson, & Reich, 2016; Vernot & Akey, 2014).



**Figure 3: Insights gleaned from Studying Archaic Admixture (A)** Schematic illustration of the out-of-Africa dispersal of modern humans (red). Blue and green shading roughly indicates the regions inhabited by Neanderthals and Denisovans, respectively. Although fossil evidence of Denisovans has been found only at Denisova Cave, the observed pattern of Denisovan admixture suggests they had a wide range that extended southeast. Reprinted from Figure 1.A (Vattathil & Akey, 2015) with permission from Elsevier. Copyright © 2015 Elsevier Inc. All rights reserved.

#### 2.1.4.1. Neanderthal introgression in New Guineans

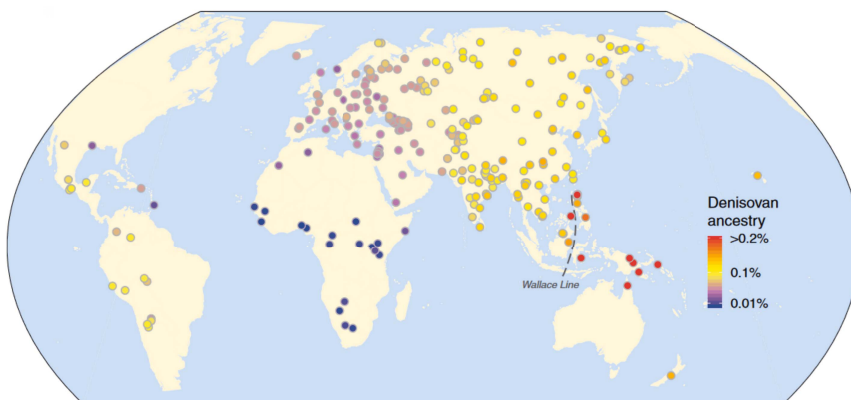
Non-African genomes carry around 2% of variants introgressed from Neanderthal (Bergström et al., 2020; Sankararaman et al., 2014). East Asians show a higher rate than Europeans (Figure 4), possibly due to a second pulse of admixture unique to the east Asian population (Vernot et al., 2016). Nonetheless, this enrichment of Neanderthal introgressed variants in East Asians remains relatively modest (Chen, Wolf, Fu, Li, & Akey, 2020). New Guineans would harbour a similar amount of Neanderthal ancestry than other non-African populations (around 2.2 to 2.9%) (Choin et al., 2021).



**Figure 4: Average proportions of Neanderthal introgression in modern humans.** Proportions of  $>1.5\%$  are presented as  $1.5\%$  for visualization. Reprinted from Figure 3.b (Yuan et al., 2021), licensed under Creative Commons CC BY license. Copyright © 2021, Yuan et al.

#### 2.1.4.2. Denisovan introgression in New Guineans

Only populations with Asian or Oceanian ancestry carry Denisovan introgression. New Guineans, along with Indigenous Australians and Philippine Ayta, stand out by carrying one of the highest Denisovan introgression rates (Larena et al., 2021; Reich et al., 2010) (Figure 5). Denisovan introgression in the region is complex because AMH have admixed independently with different Denisovan lineages (Jacobs et al., 2019). Previous studies have suggested that the current Denisovan ancestry within New Guineans results from at least two independent admixture pulses with different Denisovan lineages (Browning, Browning, Zhou, Tucci, & Akey, 2018; Choin et al., 2021; Jacobs et al., 2019). One of these admixture events likely happened before the separation of Near Oceanian and East Asian populations. The other would be more recent and private to New Guineans (Choin et al., 2021; Jacobs et al., 2019).



**Figure 5: Average proportions of Denisovan introgression in modern humans.** Proportions of  $>0.2\%$  are presented as  $0.2\%$  for visualization. Reprinted from Figure 3.a (Yuan et al., 2021), licensed under Creative Commons CC BY license. Copyright © 2021, Yuan et al.

## 2.2. Local adaptation

### 2.2.1. Survival of the fittest: how natural selection shaped human populations

Natural selection is one of the evolutionary forces that shape human diversity (Key, Fu, Romagne, Lachmann, & Andres, 2016). Humans have colonised various territories where they have experienced different environmental challenges. Under these adaptative pressures, some phenotypes might be more beneficial than others for survival. Consequently, natural selection will lead to changes in the allele frequency associated with the phenotype under selection. Different kinds of natural selection exist that can influence allele frequency in different manners.

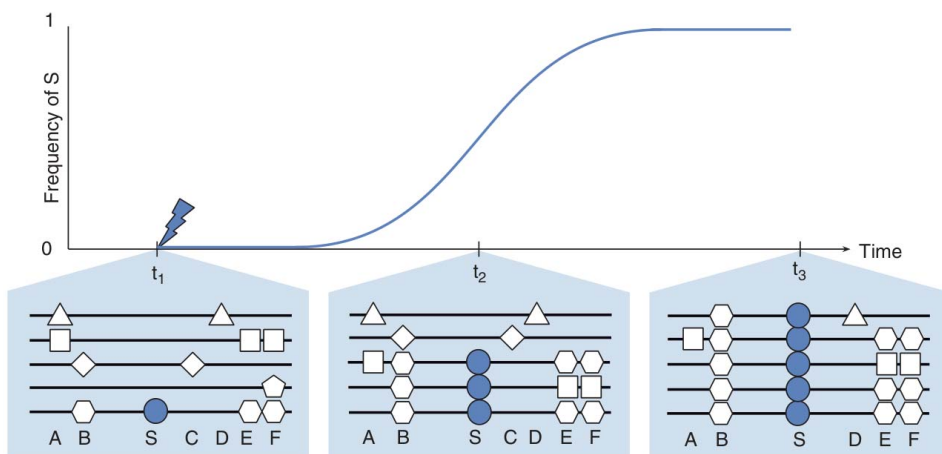
First, natural selection is called positive when acting on a genetic variant that confers an advantage in a specific environment. Individuals carrying the beneficial variant will pass it to their offspring. This offspring with higher fitness will have a higher survival rate. Carriers have then more descendants than non-carriers. The descendants of carriers will pass the beneficial variant to their offspring, which will also be more numerous. As a result, the selected variant frequency will increase in the population with time until its fixation (i.e., until it is the only remaining allele). While the selected genetic variant becomes predominant, the frequency of the other alleles on the same genomic site decrease. The loss of diversity as the selected variant frequency increases is called a selective sweep (Figure 6). A famous example of positive selection in the human population is lactase persistence, which is strongly positively selected in different populations (Bersaglieri et al., 2004). Why being able to digest milk is under such strong selective pressure is debated. It has been hypothesized that consuming milk improves survival by increasing the diet's energy content or vitamin D intake (Itan, Powell, Beaumont, Burger, & Thomas, 2009; Ségurel & Bon, 2017).

On the other hand, when natural selection leads to the removal of deleterious mutations, it is called negative or purifying selection. Negative selection decreases deleterious variants frequency in the population with time. Eventually, the deleterious variants will become rare or even disappear in the population, making negative selection hard to detect (Zhai, Nielsen, & Slatkin, 2009). Negative selection plays a major role in limiting polymorphism in loci involved in essential cellular or developmental functions (e.g., protein synthesis) and constrains the modification of the associated genome sequences to maintain them identical (conserved) through generations (Jobling et al., 2013). However, slightly deleterious mutations (i.e., nearly neutral variants) might still reach fixation when selection strength is weaker than the genetic drift, for example, when the population size is small (Ohta, 2017).

Neutral mutations, which do not confer any selective advantages or disadvantages, are the most common type of mutation found throughout the genome. Positive and negative selection indirectly impact neutral mutations through linkage disequilibrium (LD), a phenomenon in which variants located near each other on the genome are more likely to be transmitted together to the next generation. As

a result, a neutral mutation may be transmitted along with advantageous variants positively selected or removed with disadvantageous variants under negative selection. A selective sweep (i.e., variant frequency increases in the population because of positive selection, which lowers the diversity at the locus) generates a loss of polymorphism in the surrounding genomic region of the selected variant. Because of LD, the variants that co-occur nearby the selected one on the same chromosome will be transmitted simultaneously to the offspring (Kaplan, Hudson, & Langley, 1989). This process is called genetic hitchhiking. If selection is strong enough, the frequency of the selected haplotype (i.e., the set of genetic variants inherited together with the selected variant) increases faster than reverse processes, such as recombination or mutations, which break the haplotype or increase haplotype diversity (Figure 6). When a *de novo* mutation is under selection, the linked haplotype will rapidly rise in frequency in the population. This is called a hard sweep. On the other hand, a soft sweep happens when selection acts on a mutation already existing in the population, which is then likely to be included in multiple haplotypes. In this case, all haplotypes carrying the mutation will increase in frequency, which makes it harder to be detected. Similarly, because of LD, the removal of deleterious variants by negative selection results in background selection, which is the loss of non-deleterious variants linked to the variant negatively selected. Because the deleterious variant is not transmitted to the next generation, linked variants nearby this variant is less likely to be transmitted too (D. Charlesworth, Charlesworth, & Morgan, 1995).

While both positive and negative selection reduce genetic variability by favouring or removing genetic variants, respectively, another kind of selection, called balancing selection, maintains multiple alleles (polymorphisms) (Deborah Charlesworth, 2006). The high frequency of sickle-cell disease in Sub-Saharan regions has extensively been used to illustrate balancing selection (Flint, Harding, Boyce, & Clegg, 1998). This disease is caused by a mutation in the *HBB* gene that encodes the  $\beta$ -globin. People carrying the mutated gene, HbS, display an atypical haemoglobin (haemoglobin S) that creates deformed red blood cells (sickle cell). Because homozygotes carrying two copies of HbS suffer from detrimental sickle-cell anaemia, we would imagine that negative selection would have removed the HbS variant. However, heterozygotes carrying only one copy of the mutated gene will have a reduced risk of severe malaria (heterozygote advantage). Because of this advantage, heterozygotes, and then the HbS variant, are maintained in the population (Ackerman et al., 2005).



**Figure 6: Genetic variation changes as a sweep progresses due to hitch-hiking.** Individual haplotypes ( $n = 5$ ) are denoted using a different shape for each haplotype in the sample at  $t_1$ , to keep track of recombination events during the sweep. At  $t_1$ , the selected allele  $S$  mutates into the population in the  $\circ$  background. At this stage, there are six neutral polymorphisms (A–F) in the sample. At  $t_2$ , the sweep is ongoing and the frequency of the selected allele is intermediate (incomplete selective sweep). Relative to a neutral allele, fewer recombination events have occurred around the selected allele by the time it reaches this frequency, due to its rapid increase in frequency driven by selection. As a result, diversity within haplotypes carrying  $S$  is much reduced compared to haplotypes carrying the disfavored ancestral allele, that is, there has been an increase in haplotype homozygosity within the allelic class carrying  $S$ . Note that at this stage, two recombinations have occurred, both between  $\square$  and  $\circ$ . At  $t_3$ ,  $S$  has swept to fixation, along with the  $B$  allele. Another recombination event has occurred between  $\triangle$  and  $\circ$ . Note the increase in high-frequency derived alleles and overall reduced levels of variability at  $t_3$  relative to  $t_1$  and  $t_2$ , exemplified by the loss of diversity at three sites (B, C, S). Furthermore, the sample is IBD for the entire tract  $B - S$ . Reprinted from Figure 14.1 (Stern & Nielsen, 2019) with permission from John Wiley and Sons. Copyright © 2019 John Wiley & Sons Ltd.

## 2.2.2. Genomic approaches to detecting positive natural selection

While negative and balancing selection (Fijarczyk & Babik, 2015; Zhai et al., 2009) are complex to identify, multiple methods exist for detecting positive selection at the genome level. These techniques screen for genomic patterns compatible with signatures associated with positive selection (Rees, Castellano, & Andrés, 2020).

### 2.2.2.1. Haplotype-based selection scans

One signature associated with genomic regions that have been a target of positive selection is a reduced diversity on a haplotype. Estimates of haplotype homozygosity have therefore been used to scan for such regions in a population. The Extended Haplotype Homozygosity (EHH) metric is one of the statistics that

estimates such signals. The EHH score reflects the probability that two randomly chosen haplotypes are identical around a SNP up to a distance  $x$ . Because of recombination, EHH decreases when the distance  $x$  from the SNP increases. Under selection, there is less variation around the SNP – haplotypes carrying the selected SNP are more similar – and EHH will decrease more slowly (Sabeti et al., 2007; Stern & Nielsen, 2019).

The integrated Haplotype Score (iHS) is a later modification of EHH that considers the local recombination rate. Using genetic distance instead of physical distance corrects for recombination rate. Indeed, regions with low recombination tend to display more haplotype homozygosity (Voight, Kudaravalli, Wen, & Pritchard, 2006). EHH can also be modified into cross-population extended haplotype homozygosity (XP-EHH) that detects selective sweep that has occurred in one population but not another (Sabeti et al., 2007). XP-EHH detects selective sweeps in which the selected allele has achieved fixation in one population (complete sweep) but remains polymorphic in the overall human population.

Another haplotype-based statistic calculates the number of segregating sites by length (nSL) and is a measure that does not require a genetic map. Instead, it computes the length of haplotype homozygosity between two haplotypes with respect to the number of mutations within the other haplotypes from the studied dataset (Ferrer-Admetlla, Liang, Korneliussen, & Nielsen, 2014). Similarly to XP-EHH, a cross-population approach using nSL – called XP-nSL – exists that compares haplotypes between two populations. This approach offers a favourable method for detecting soft sweeps compared to XP-EHH (Ferrer-Admetlla et al., 2014; Szpiech, Novak, Bailey, & Stevison, 2021).

#### 2.2.2.2. Allele frequency-based selection scan: Population Branch Statistic

The population branch statistic (PBS) is used to detect another type of genomic signature of positive selection. While XP-EHH or XP-nSL are based on haplotype homozygosity, PBS uses allele frequency differences between populations to identify signatures of selection. Three populations are needed to compute PBS for a target population: a target population, a reference population and an out-group population (Yi et al., 2010). A population's PBS value represents the amount of allele frequency change at a given genomic position in the history of a target population.

The allele frequency will be used to estimate the fixation index ( $F_{ST}$ ) between each population at each SNP.  $F_{ST}$  is as common neutrality test that characterizes the genetic differentiation between two populations. There are multiple ways to estimate  $F_{ST}$ , but they all reflect the amount of genetic variation between populations ( $V_B$ ) as a proportion of the total variation ( $V_T$ ) that includes within population and between populations variations (eq. 1) (Beaumont & Wang, 2019).

$$F_{ST} = V_B/V_T \quad (\text{eq. 1})$$

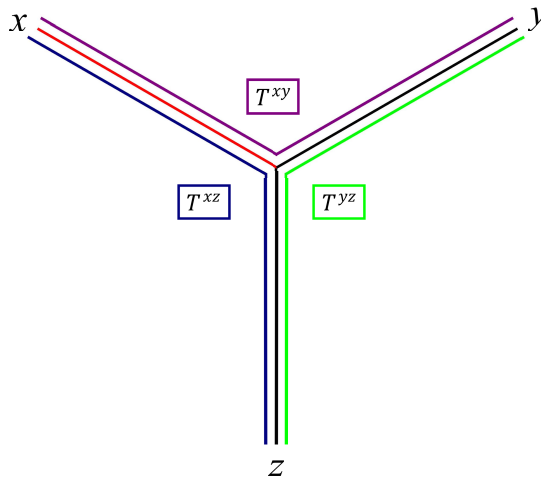
When the allele frequency of one variant highly differs between two populations, the corresponding  $F_{ST}$  value will be close to 1, while it will be equal to 0 when the allele frequency is identical between two populations. High genetic differentiation of a SNP (high  $F_{ST}$ ) between two populations might result from selection (Lewontin & Krakauer, 1973; Stern & Nielsen, 2019). However, the  $F_{ST}$  value alone cannot decipher within which population the variant is under selection. Using an outgroup population (i.e., a population as distant to the target as the reference population) for the PBS calculation allows for the directional assessment of selection (Yi et al., 2010). When picturing a tree with three branches (one for each population) (Figure 7),  $F_{ST}$  is used to estimate the branch length (or the population divergence time  $T^{xy}$ ) between every population pair (eq. 2). Or for population  $x$  and  $y$ :

$$T^{xy} = -\log(1-F_{ST}(x,y)) \quad (\text{eq. 2})$$

The branch length between each pair of populations is used to calculate the branch length from the inner node of the tree to the target population  $x$ . This inner branch length is the PBS.

$$\text{PBS} = \frac{T^{xy} + T^{xz} - T^{yz}}{2} \quad (\text{eq.3})$$

The frequency change of a genetic variant under selection is often more dramatic than that observed in variants that are not under selection, which increases its PBS score (Figure 7) (Yi et al., 2010).



**Figure 7: Computing locus-specific population branch statistic (PBS) for the target population  $x$ , considering a reference population  $y$  and an outgroup  $z$ .** Log transformed pairwise  $F_{ST}$  are used to estimate  $T^{xy}$ ,  $T^{xz}$  and  $T^{yz}$  (eq. 2), the branch length (or the divergence time) between the target and reference populations (in purple), the target and outgroup populations (in blue) and the reference and outgroup populations respectively. Pairwise branch lengths are used to estimate PBS (in red), the length of the branch to the target population  $x$  since the divergence from the reference population  $y$  and the outgroup population  $z$ , following eq. 3.

### 2.2.3. Challenges and limitations in interpreting selection

Although selection scans offer insights into the genomic regions under selection in a population, establishing a link between the associated genomic region and a potential target phenotype is challenging.

One way to identify the biological consequences of the candidate variants for selection is by testing the variants with functional assays to determine their function in a non-human model organism. Still, this approach is limited by the reduced transferability of a gene function between organisms (i.e., from the model organism to humans) (Rees et al., 2020). The growing number of large-scale genome-wide association studies (GWAS) have provided another source to test genomic variants for their phenotypic effects (e.g., UKbiobank (Pan-UKB team, 2020)). Nonetheless, GWAS are performed on a specific population, and the transferability of the summary statistic between populations is limited (Mathieson, 2021).

An additional challenge is that a similar phenotypic adaptation can occur due to different genetic variants, meaning that populations under identical selection pressures can display the same phenotype but with distinct genetic causes (= convergent evolution). For instance, high-altitude populations have independently adapted to low oxygen availability through genes impacting the Hypoxia-Inducible Factor (HIF) pathway. HIFs are transcription factors regulating the hypoxic response (Bigam & Lee, 2014).

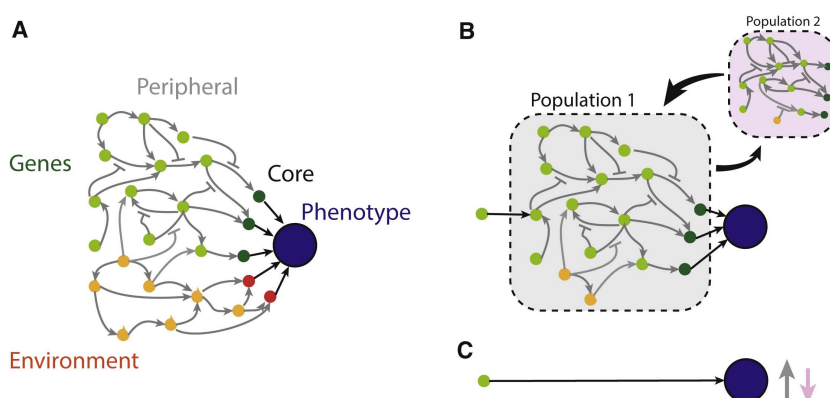
It is also important to note that most phenotypes are polygenic, meaning they are impacted by many genetic variants, each of which only has a limited effect. When natural selection acts on these complex phenotypes, the frequency of each genetic variant that impacts the phenotype will only shift by a small degree. This subtle frequency shift makes detecting polygenic selection difficult (Sella & Barton, 2019).

Pleiotropy is another phenomenon, which relates to single genes or single genetic variants, that can have an effect on multiple unrelated traits, which makes it challenging to establish the relationship between a gene that is under selection and a phenotype that may be the target of selection (Stearns, 2010).

Finally, because the environment might have changed since the time of selection, identifying the initial environmental pressure that acted on a population remains complex. An example is the Duffy-null phenotype, protecting against *P. vivax*, found in high frequency in Africa, while *P. vivax* is not endemic in Africa anymore (McManus et al., 2017)). In addition, a phenotype detrimental to health might still benefit survival under specific pressure (ex: sickle cell disease) and maintained in the population by balancing selection (Fan, Hansen, Lo, & Tishkoff, 2016).

The recent omnigenic model elegantly put together these multiple concepts by suggesting there exist only a few “core” genes that directly affect a phenotype (Boyle, Li, & Pritchard, 2017). Most of the genes would be “peripheral” and influence the phenotype indirectly through a network of interactions with multiple peripheral and core genes. Because of the dense interconnection between genes, a single variant may affect various traits. This process is called network

pleiotropy. Nonetheless, phenotypes do not only result from genetics but are also impacted by various environmental factors. For example, obesity is associated with multiple genes and environmental factors like diet or smoking (Loos & Yeo, 2022). The omnigenic model can be extended by adding core and peripheral environment effects to the network to take account of the environmental influence on a phenotype. Between populations, the network will vary: core genes are conserved, but interactions between peripheral genes and environmental effects vary. Different omnigenic networks between populations explain the differences in effect size between GWAS in different populations (Figure 8) (Mathieson, 2021). Identifying the phenotypic consequences of variants with signatures of selection in the omnigenic network is challenging and exemplifies why translating selection signatures to their phenotypic targets is often difficult.



**Figure 8: The omnigenic model** (A) Schematic of the omnigenic model (after Liu, Li, & Pritchard, 2019). Genetic factors are in green and environmental factors are in orange, with arrows showing interactions. Peripheral factors are lightly shaded, whereas core factors with direct effects on phenotype are darkly shaded. (B) A gene's-eye perspective. A single peripheral gene's effect on the phenotype is filtered through part of the network (grey box). (C) The GWAS perspective. The effect of a causal variant as measured by an association study in a single population (grey arrow in C) is the expected value of its effect, with respect to the distribution of genetic (i.e., allele frequencies) and environmental factors in the population. In a different population (pink box in B), the weights and possibly structure of the network change, leading to a different expected effect size (pink arrow in C). Reprinted from Figure 1 (Mathieson, 2021) with permission from Elsevier. © 2021 American Society of Human Genetics.

#### 2.2.4. Enhancing fitness through archaic interbreeding: the role of adaptive introgression

Neanderthals inhabited a vast territory from Southwest Europe to South Siberia from around 300 kya until around 40 kya (Higham et al., 2014; J. J. Hublin, 2009). There is a lack of fossil records for Denisovans, but indirect evidence suggests that their territory might have spread from Siberia (Reich et al., 2010) to Sulawesi (van den Bergh et al., 2016). They might have inhabited these territories between

100 and 200 kya (Douka et al., 2019; van den Bergh et al., 2016). The extended time that archaic humans, such as Neanderthals and Denisovans, were exposed to their environments gave selection time to act on their genome and increase their fitness to environmental challenges. When AMH interbred with these archaic groups, their genomes inherited some of these selected archaic variants that might have helped them to enhance their fitness faster than under a selective process acting on a *de novo* mutation (Racimo, Sankararaman, Nielsen, & Huerta-Sánchez, 2015). A noteworthy example is a variant of the *EPAS1* gene introgressed from Denisovans, found in high frequencies in Tibetans, and associated with reduced haemoglobin concentration. The archaic variants at this gene locus might have helped Tibetan to counteract high-altitude hypoxia's detrimental effects (Huerta-Sánchez et al., 2014). Nonetheless, selection might have happened significantly after the admixture event and would not constitute a real adaptative introgression but rather a selection on a standing variant introgressed earlier (X. Zhang et al., 2021). Adaptative introgression has also notably contributed to the modern human immune system and skin pigmentation (Dannemann, Andrés, & Kelso, 2016; Mendez, Watkins, & Hammer, 2012; Reilly et al., 2022; Sams et al., 2016; Sankararaman et al., 2016; Vernot & Akey, 2014).

On the other hand, background selection has also been shown to play an important role in introgressed archaic DNA. The smaller effective population sizes of Neanderthals and Denisovans, compared to AMH, meant that selection against harmful gene variants was weak in these archaic hominin populations. As a result, their genomes accumulated weak deleterious alleles. However, after admixture with AMH, which had a larger effective population size, selection against these introgressed deleterious variants became stronger (Harris & Nielsen, 2016; Juric, Aeschbacher, & Coop, 2016). Negative selection has particularly targeted introgressed variants in conserved regulatory regions (Petr, Pääbo, Kelso, & Vernot, 2019). In addition, there are also large genomic regions in present-day people that are devoid of introgressed Neanderthal and Denisovan variants, also referred to as archaic deserts. These regions are enriched for genes expressed in the brain and testis. They are also enriched on the X chromosome. While it is difficult to pinpoint the exact target of selection, these results suggest that sex-biased admixture processes, potentially associated with incompatibilities in reproduction or brain-related traits, might have driven the removal of archaic DNA in these regions (Sankararaman et al., 2014; Vernot & Akey, 2014; Vernot et al., 2016).

#### 2.2.4.1. Adaptative introgression in New Guineans

New Guineans with Indigenous Australians and Philippine Ayta carry both Neanderthal introgressions and have one of the highest Denisovan introgression rates (Larena et al., 2021; Reich et al., 2010). Introgressed loci in New Guineans might have enabled them to face the environmental pressure in New Guinea by shaping their immune system (Vespasiani et al., 2022). For example, *TNFAIP3* (Gittelman et al., 2016; Zammit et al., 2019), *STAT2* (Mendez et al., 2012; Yermakovich et al., 2022), and the *GBP* locus (Vernot et al., 2016) are three

introgressed archaic loci candidates for selection in New Guineans that are involved in the interferon-induced cell-autonomous host defence (MacMicking, 2012). Moreover, archaic introgression might also have contributed to New Guinean dietary adaption through changes in the metabolism (e.g., *FADS1* (Jacobs et al., 2019), *SUMF1* (Choin et al., 2021; Jacobs et al., 2019)).

## 2.2.5. Adaptation to extreme environments

Since they arrived in northern Sahul, New Guinean populations have experienced various selection pressures associated with the variety of environments they have settled in. New Guinean highlanders and lowlanders face different environmental challenges, with the strongest challenges being hypoxic stress and pathogenic pressure, respectively.

### 2.2.5.1. Altitude pressure

#### 2.2.5.1.1. *Altitude pressure worldwide*

Altitude is a severe environmental pressure inflicted on human beings (Fan et al., 2016). The human body's biggest challenge at altitude is hypoxia– the lower oxygen availability to body tissues. Hypoxia not only occurs during summit ascent but is also a condition common to various diseases (e.g., heart failure, lung diseases, anaemia, cancer, covid-19) (Lee, Chandel, & Simon, 2020; Rahman et al., 2021). Hypoxia is also a common pregnancy complication (Piešová & Mach, 2020). The hypoxia-inducible factor (HIF) pathway orchestrated most cellular mechanisms that preserve oxygen homeostasis in the hypoxic organism (Lee et al., 2020). But despite its importance in human health, the body's responses to hypoxia are still not fully understood (Lee et al., 2020). Many studies conducted in high-altitude populations have enhanced our knowledge of the human body's strategy to counteract the detrimental effect of hypoxia.

For example, Tibetans, Andeans, and Ethiopian Amharas display phenotypic traits that seem to counterbalance the decrease in oxygen in the blood because of hypoxia (Moore, 2017). Andeans have a wider chest, larger lung capacity, and higher haemoglobin levels compared to their corresponding lowlanders (C. M. Beall et al., 1998; Brutsaert, Soria, Caceres, Spielvogel, & Haas, 1999; Mueller, Schull, Schull, Soto, & Rothhammer, 1978), while Tibetans show lower haemoglobin levels due to higher plasma volume (C. M. Beall et al., 1998; Stembridge et al., 2019). At first glance, because haemoglobin is the molecule transporting oxygen in the blood, lower haemoglobin concentration can seem maladaptive for living at high-altitude. Indeed, in response to the lower oxygen levels, lowlanders at high-altitude will experience an increase in haemoglobin concentration to enhance oxygen delivery to the body tissues. Nonetheless, an increase in haemoglobin concentration also results in higher blood viscosity, which is detrimental in the long term. Higher blood viscosity puts more stress on the cardiovascular system and results in cardiovascular and pregnancy complications. The high

haemoglobin levels of Andean highlanders might even explain their high susceptibility to chronic mountain sickness (CMS) (Villafuerte & Corante, 2016). Ethiopian highlanders also show increased haemoglobin levels and a larger chest (Alkorta-Aranburu et al., 2012; Hoit et al., 2011; Scheinfeldt et al., 2012), although not to the same extent as Andeans. However, not all studies have confirmed the increased haemoglobin levels in Ethiopians (Cynthia M. Beall et al., 2002; Cheong et al., 2016; Huerta-Sánchez et al., 2013).

Genomic studies revealed that many genes considered for high-altitude adaptation are associated with the HIF pathway (Bigam & Lee, 2014). Among these genes are *EGLN1*, which is under selection in both Andean and Tibetan highlanders, and *PRKAA1*, *EPAS1* and *HLHE41*, which are candidates for selection in Andean, Tibetan and Ethiopian Amhara highlanders, respectively (Bigam & Lee, 2014; Huerta-Sánchez et al., 2013). *EPAS1*– associated with the characteristic lower haemoglobin concentration level in Tibetans (Simonson et al., 2010; Yi et al., 2010) – is particularly noteworthy as its selected Tibetan variant is associated with an introgressed Denisovan haplotype (Huerta-Sánchez et al., 2014). Nonetheless, it has been suggested that the selection of this variant postdates the introgression event (X. Zhang et al., 2021). Another matter of discussion is the phenotype targeted by selection in Tibetans. It has been shown that selection might target a higher plasma volume instead of a higher haemoglobin concentration (Stembridge et al., 2019). Moreover, selection to hypoxia also acted on intermediate altitude populations such as Andean Calchaquíes living around 2,300 m a.s.l. that show a strong signal of selection for *PRKG1*, suggesting adaptation of the nitric oxide pathway (Eichstaedt et al., 2015).

Although natural selection may explain part of the specific phenotypes of highlanders worldwide, these traits may result from other non-mutually exclusive factors such as genetic adaptation to other high-altitude stressors (e.g., cold) (Payne, Kumar Bc, Pomeroy, Macintosh, & Stock, 2018), ancestry, high-altitude exposure during growth (Frisancho, 2013), lifestyle (Cynthia M. Beall, 2014; Leonard, 1989), and socioeconomic status (Greksa, 2006).

#### 2.2.5.1.2. Altitude pressure in New Guinea highlands

New Guinea has more than 40 summits above 4000 m a.s.l. and is the most mountainous region of the Sahul continent. The first settlers entered New Guinean highlands for semi-permanent settlement as early as 25 kya (Summerhayes et al., 2017). However, despite the extended time they have inhabited the highlands, New Guineans highlanders have been understudied regarding adaptation to hypoxia. Since they permanently settled in the highlands 20 kya (Malaspinas et al., 2016; Mountain, 1991; O’Connell et al., 2018; Summerhayes et al., 2010), New Guinean highlanders remained relatively genetically isolated through time, with only minor influences from outside (Bergström et al., 2017). Nowadays, the most densely populated places in PNG are located at intermediate altitude (above 1500 m a.s.l.) between 1,600 and 2,400 m a.s.l. (Brookfield & Allen, 1989; Müller et al., 2003). Previous studies on the health of PNG populations showed

that highlander groups display an increase in ventilatory lung function and haemoglobin concentration (Cotes, Anderson, & Patrick, 1974; Senn et al., 2010; Woolcock, Colman, & Blackburn, 1972). While these data were not explicitly studied in the context of adaptation to hypoxia, they strongly suggest that PNG highlanders could display similar phenotypic traits as those observed in other intermediate and high-altitude populations worldwide.

#### 2.2.5.2. Pathogenic pressure

As AMH expanded across new territories, they were exposed to different pathogens that exerted strong selective pressure leading to the selection of protective genes (Fumagalli et al., 2011). At the same time, pathogens evolved to counteract the humans' newly acquired adaptations, leading to new selective pressures. Consequently, the immune system has been a prime target of natural selection throughout human evolution (Siddle & Quintana-Murci, 2014). Moreover, environmental changes (e.g., climate change) and cultural practices (e.g., agriculture, medicine) created constantly shifting pathogenic pressures. This dynamic interplay of adaptive traits and selective pressures is theorized by the Red Queen hypothesis (Van Valen, 1973). It explains how pathogenic pressure and diseases are significant and constant driving forces in human evolution. This remains true today, as demonstrated by the recent emergence of novel pathogens such as SARS-CoV-2, the virus responsible for COVID-19 (Souilmi et al., 2021). These constant challenges left distinct signatures of selection in the genomes of human populations. For example, pathogenic pressure has driven the diversity of Human Leukocyte Antigen (HLA) class I genes in humans (Prugnolle et al., 2005). More specifically, some populations show adaptation to a specific disease, like the above-mentioned widespread sickle-cell allele HbS protecting sub-Saharan populations from malaria (Ackerman et al., 2005). Additionally, archaic introgression may have contributed to immune system adaptations in AMH (Dannemann et al., 2016; Enard & Petrov, 2018; Mendez et al., 2012; Quach et al., 2016; Sams et al., 2016; Vespasiani et al., 2022).

##### 2.2.5.2.1. Malaria pressure worldwide

Malaria is the most common parasitic disease in humans (White, 2018) and one of the leading causes of death for children under five years old (World Health Organization, 2022b). This infection of the red blood cells by a protozoan parasite of the *Plasmodium* group is transmitted through bites of infected *Anopheles* mosquitoes. *Plasmodium falciparum* and *Plasmodium vivax* are the leading cause of malaria in humans (Haldar & Mohandas, 2009). Malaria infection induces the destruction of parasitised and unparasitized red blood cells and bone marrow dysfunction (Totino, Daniel-Ribeiro, & Ferreira-da-Cruz, 2016; White, 2018). Malaria is of major health concern worldwide. The World Health Organization (WHO) recorded 247 million malaria cases and 619 000 deaths from malaria in

2021. Most of these cases were in sub-Saharan countries (World Health Organization, 2022a).

The most common effect of malaria is anaemia – a decrease in haemoglobin concentration – which causes substantial infant mortality in the affected territories (Haldar & Mohandas, 2009; White, 2018). Another common side effect of malaria is decreased platelet count (thrombocytopenia) (O’Sullivan, Preston, O’Regan, & O’Donnell, 2016). Malaria can reach more severe stages like cerebral anaemia, when the infected red blood cells stick to the brain endothelium from *P. falciparum* infection (Bafaro, Liu, Xu, & Dempski, 2017; Haldar & Mohandas, 2009). Platelets have been shown to play a crucial role in this process by mediating the adhesion of infected red blood cells together and to the endothelium (O’Sullivan et al., 2016; Pain et al., 2001). This haemostatic dysfunction can even result in vascular occlusion and increase the risk of mortality from *P. falciparum* or *P. vivax* malaria (Lampah et al., 2015). However, platelets also have beneficial effects in regulating malaria symptoms by binding to infected red blood cells and contributing to *Plasmodium* killing (Kho et al., 2018). Thus, platelets play a complex and essential role in the pathogenesis of malaria, but their complete contribution to disease outcomes is subject to ongoing research (McMorran, 2019).

While most of the deaths and severe symptoms are attributable to *P. falciparum*, *P. vivax*, the most widespread type of malaria outside of Africa (Gething et al., 2012), can still lead to severe anaemia (Haldar & Mohandas, 2009; White, 2018). Moreover, *P. vivax* might induce heart pathologies (Bammigatti, Shetty, Shetty, & Kumar, 2011; Bhat, Kumar, & Alva, 2013; Gupta et al., 2021; Holm, Gomes, Biering-Soerensen, Silvestre, & Brainin, 2020).

Because of its high mortality rate among children, malaria exerts strong selective pressures on the human genome (Kwiatkowski, 2005). Malaria-induced selective pressure is linked to the emergence of high-frequency haemoglobinopathies and red blood cell polymorphisms in impacted regions (Flint et al., 1998). A striking example we have addressed in the chapter above is the frequency of the HbS allele, causing deformed red blood cells. While HbS homozygotes suffer from sickle-cell anaemia that can often be fatal, heterozygotes are protected against severe forms of malaria (Ackerman et al., 2005). The heterozygote advantage maintains HbS at around 10% in malaria-endemic regions thanks to balancing selection (Flint et al., 1998). But malaria has kept a huge variety of other red blood cell polymorphisms, such as the Duffy blood group-negative phenotype that offer a complete resistance against malaria caused by *P. vivax*. Duffy-null red blood cells do not carry the Duffy antigen receptor for chemokines (DARC) that *P. vivax* uses to infect the red blood cell. The allele associated with this phenotype is called FY\*O, which is near fixation in western and central Africa but carries the signature of strong selection (McManus et al., 2017).

#### 2.2.5.2.2. Pathogenic pressure in New Guinea lowlands

New Guineans are exposed to a variety of pathogens (e.g., malaria, dysentery and pneumonia, tuberculosis, HIV, etc.) that are the leading cause of death in PNG (GBD 2013 Mortality and Causes of Death Collaborators, 2015; Kitur et al., 2019; Naraqi et al., 2003). Nevertheless, the environmental diversity in PNG leads to a unique situation regarding the epidemiology of malaria. We previously discussed the uneven population distribution in PNG, with most people living between 1600 and 2400 m a.s.l. (Brookfield & Allen, 1989; Müller et al., 2003) since the Holocene (Trájer et al., 2020). A possible explanation for this uneven distribution could be the migration of early settlers of New Guinea towards the highlands due to the high pathogenic pressure in the lowlands (Riley, 1983; Trájer, 2022; Trájer et al., 2020). Malaria might be one of the leading causes of this pathogenic pressure, even though we do not know when it first appeared in the lowlands (Trájer et al., 2020). Indeed, malaria is widespread in New Guinea: PNG accounted for nearly 87% of the malaria cases and 94% of malaria deaths in the Western Pacific Region in 2021 (World Health Organization, 2022a). Malaria estimated mortality rate of children under 10 is between 4 and 17% depending on the region in PNG (Cleary, Hetzel, & Clements, 2022). Despite these numbers, New Guinean highlands are nearly malaria-free (Müller et al., 2003; Senn et al., 2010). An association between altitude level and lower malaria prevalence has also been observed in other populations like Tanzanians and Kenyans (Akhwale et al., 2004; Drakeley et al., 2005).

New Guineans lowlanders carry polymorphisms associated with blood-related traits that might have been selected to face this endemic malaria pressure (Müller et al., 2003), like G6PD deficiency,  $\alpha$ -thalassaemia, and the Gerbich (Ge)-negative blood group, that are absent or rare in PNG highlands (Müller et al., 2003). Interestingly, PNG highlanders affected by malaria have a higher risk of thrombocytopenia (decrease in the platelet count) than lowlanders, suggesting the absence of selection in PNG highlanders (Lampah et al., 2015).

## 2.3. Estimating genealogies

Inferring genealogies, considering coalescent and recombination events for a set of sequences, has become an essential tool in statistical population genomics. However, estimating genealogies for large whole genome datasets can be computationally challenging. This has led to the development of various statistical methods for inferring genealogies, providing insight into the evolutionary history of populations and the forces that have shaped their genomes.

### 2.3.1. The neutral Wright-Fisher model

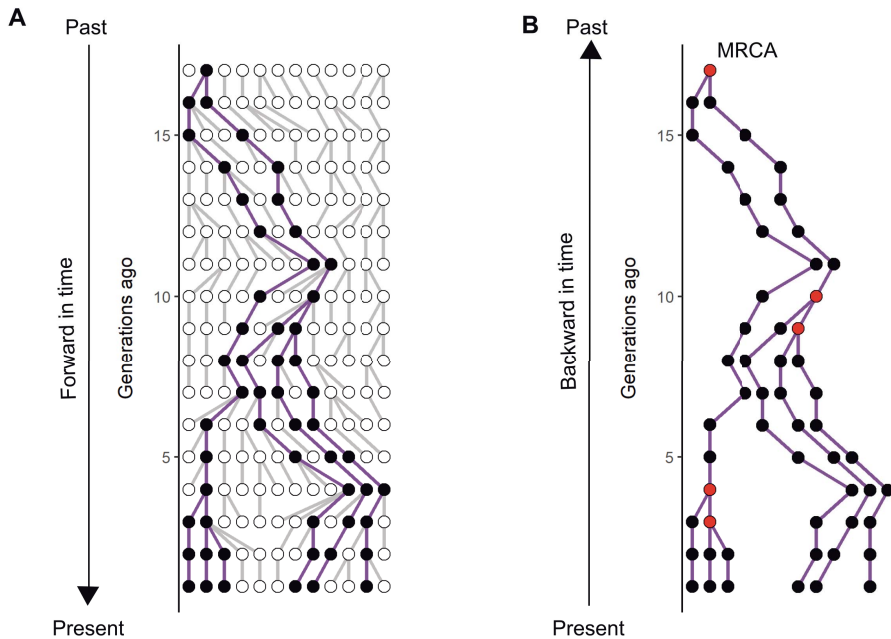
To draw meaningful evolutionary inferences from a dataset, it's crucial to understand the genealogy that has led to its diversity. The neutral Wright-Fisher model is widely used to build such genealogy (Hein, Schierup, & Wiuf, 2005). This

model represents the effects of genetic drift and random neutral mutation at a given locus of the studied samples. The Wright-Fisher model is used to infer demographic and evolutionary parameters that have led to the current genomic diversity (Nordborg, 2019). It is an idealized representation of how a population evolves over time and relies on several assumptions that simplify the simulation of genealogies. The first assumption is neutrality, meaning that selection has not influenced the allele frequency in the studied dataset. This assumption implies that no mutation in the population has been selected positively or negatively. Therefore, no mutation in the population will affect the number of offspring produced by carriers of that mutation. Panmixia, or random mating, is a second assumption of this model. Under this assumption, each individual in a given generation is considered to have randomly inherited its genes from the previous generation's pool of parents. In other words, each parent in the previous generation has an equal chance of being the genetic contributor for any given individual in the next generation. Consequently, the model can be applied to a subset of a population without knowledge about the whole population. Finally, the Wright-Fisher model assumes discrete and non-overlapping generations, an absence of migration and constant population size at every generation. Individuals at each generation are obtained by sampling with replacement from the parents (Hein et al., 2005; Nordborg, 2019). Besides its simplicity, the Wright-Fisher model becomes computationally challenging when the population size is much larger than the size of the dataset being studied, as it is usually the case.

### 2.3.2. The coalescent theory

The coalescent model approximates the Wright-Fisher model for large populations (Kingman, 1982). A key feature of the coalescent theory is that the genealogy is built backwards in time. When we think of a family tree, we instinctively imagine it is going forward in time, from the ancestors to the descendants. Lineages of this tree split into separate new lineages whenever an individual has multiple offspring and end when there are no more descendants. However, coalescent trees are modelled in reverse: they start with the dataset being studied and go backwards to the ancestors. In a coalescent tree, lines converge (coalesce) when two or more individuals share a common ancestor (Figure 9). Because coalescent trees are built backwards in time, lineages without descendants at the basis of the tree are excluded. Not having to do calculations for these extinct lineages enable more efficient computing than the Wright-Fisher model.

Coalescent trees keep track of coalescence events between lineages until, eventually, the most recent common ancestor (MRCA) to the sample is found; this gives the time to the most recent common ancestor (TMRCA) or the first time when the sample had one ancestor only (Hein et al., 2005; Nordborg, 2019). Mutations seen in the dataset are superimposed on the tree branches in a forward-in-time manner. The more mutations there are on a tree branch, the longer the branch is and the time to the next coalescent event (Hein et al., 2005; Nordborg, 2019).



**Figure 9: Genealogy of a sample of 6 individuals inferred under the Wright-Fisher model and the coalescent model.** **A.** Genealogy for a population of size 12 under the neutral Wright-Fisher model for 17 generations. The Wright-Fisher model works forward in time, from the past to the present. Individuals that are ancestors to a sample of 6 individuals in the final generation are represented as black dots. The genealogy of the sample is shown in purple. **B.** Tree of the sample inferred with the coalescent model. Because the coalescent theory has a backwards-in-time perspective, the tree only tracks the ancestors that have contributed to the studied sample. This process is more efficient than the Wright-Fisher model – especially when the population size is larger than the studied sample – because it does not have to track individuals that did not contribute to the study sample. Coalescent events are shown in red. The last coalescent event of this tree corresponds to the sample’s most recent common ancestor (MRCA).

The coalescent theory is used to understand the statistical properties of a sample of genomes from a population. It is particularly useful to study biological phenomena that affect the genealogical process but do not affect the mutation process (e.g., population subdivision). It is also used to understand how model parameters (mutation rate, generation time, etc.) affect polymorphism data by understanding how they affect genealogies (Nordborg, 2019).

### 2.3.3. Coalescent with recombination

During gamete formation, a diploid cell in humans containing 23 pairs of chromosomes undergoes a division that produces four haploid cells, each with 23 chromosomes. During this cell division, called meiosis, variants located on different chromosomes will be transmitted independently to different gametes. In contrast, variants on the same chromosome will be inherited together. However, two

chromosomes from the same pair (homologous chromosomes) can crossover during meiosis and exchange some of their genetic material. This crossover creates a recombination event: variants initially located on the same chromosome are not inherited together (Jobling et al., 2013). The more physical distance there is between two variants, the more likely the recombination event will happen between them. In contrast, variants close to each other remain more likely to be co-inherited, resulting in LD between these variants (Altshuler, Donnelly, & The International HapMap Consortium, 2005).

The frequency of recombination between two variants is not linearly associated with the physical distance between these variants. Indeed, the recombination rate is non-uniform along the genome: recombination events are more frequent toward the telomere but less frequent toward the centromere. Moreover, there are recombination hotspots, small genomic regions with a high recombination rate, or coldspots where the recombination rate is way lower (Crawford et al., 2004; McVean et al., 2004; Myers, Bottolo, Freeman, McVean, & Donnelly, 2005). The recombination rate also differs between sexes, populations and individuals (Kong et al., 2010). Because the physical distance, measured in base pair (bp), cannot reflect the frequency of recombination between two variants, a genetic distance has been defined and is expressed in centiMorgan (cM). One cM between two variants equals a recombination event frequency of 1% between these two variants per generation. Genetic distance measurements along the genome are stored in a genetic map (or recombination map) (Haldane, 1919).

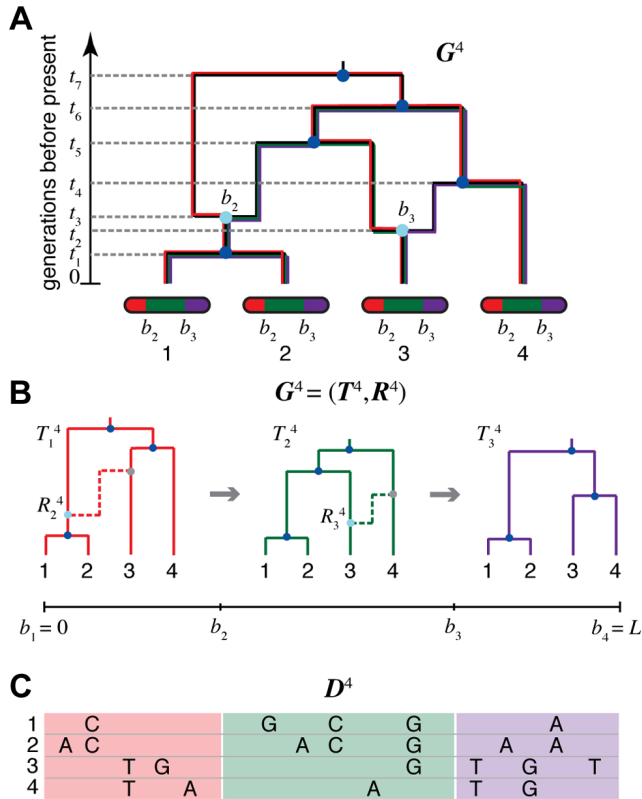
Recombination can break down the ancestral relationships between haplotypes, leading to more complex genealogical histories. The classical coalescent theory assumes no recombination between loci, and the genealogy of haplotypes follows a simple branching pattern back to a common ancestor. However, since considering recombination leads to a more complex genealogy, it becomes necessary to consider the probability of different genealogical histories. Therefore, incorporating recombination events into coalescent models is essential to accurately infer the genealogical relationships between haplotypes and their evolutionary history. Considering recombination is particularly important in population genetics research, where understanding the genealogy of haplotypes is essential for inferring demographic and evolutionary processes.

#### 2.3.3.1. Representing coalescent with recombination: Ancestral Recombination Graph

The addition of recombination boosts the complexity of the coalescent model. When going backwards in time, coalescent events are represented as two sequences merging into one common ancestor, while recombination events is a sequence that splits into two different ancestors (Figure 10). Because of these recombination events along the genome, different genomic loci can have different genealogies, and a genomic sequence cannot be represented with a unique tree anymore (Hein et al., 2005; Nordborg, 2019). Instead, one tree will exist for each site of the sequences (local trees). The genealogy of the whole sequence will be

a collection of these local trees in the form of a complex graph called the ancestral recombination graph (ARG) (Griffiths & Marjoram, 1997; Hudson, 1983).

The ARG is precious for studying evolutionary scenarios where recombination has occurred, especially when analyzing whole-genome data. ARG can be used to visualize relationships among lineages, estimate recombination and mutation rate, date mutation and recombination events, extract information about the demography history of the sample or detect selection (Arenas, 2013; Hubisz & Siepel, 2020).



**Figure 10: An ancestral recombination graph (ARG) for four sequences.** (A) Going backwards in time (from bottom to top), the graph shows how lineages that lead to modern-day chromosomes (bottom) either “coalesce” into common ancestral lineages (dark blue circles) or split into the distinct parental chromosomes that were joined (in forward time) by recombination events (light blue circles). Each non-recombining interval of the sequences (shown in red, green, and purple) corresponds to a “local tree” embedded in the ARG (shown in matching colours). Recombinations cause these trees to change along the length of the sequences, making the correlation structure of the data set highly complex (B) Representation of the ARG in terms of a sequence of local trees  $T^4$  and recombination events  $R^4$ . (C) An alignment of four sequences,  $D^4$ , corresponds to the linearized ARG shown in (B). Reprinted from Figure 1 (M. D. Rasmussen, Hubisz, Gronau, & Siepel, 2014), licensed under Creative Commons CC BY 4.0 license. Copyright © 2014 Rasmussen et al.

### 2.3.3.2. Inferring coalescent with recombination with the Hidden Markov model

Coalescent models with recombination can be easily simulated (Hudson, 1983) but using these models on genome-wide data remains computationally demanding. However, some models can now approximate coalescence with recombination on sequenced data. One such model is the Li and Stephens (LS) model, which invokes a Hidden Markov Model (HMM) (Li & Stephens, 2003; Lunter, 2019; M. D. Rasmussen et al., 2014). An HMM models a sequence of values (called states) considering two fundamental properties. First, a future state depends only on the current state, so there is no need to know the full chain of events. Secondly, the states are hidden, meaning they cannot be observed directly but are associated with observable variables. An HMM uses observable states to estimate hidden states. The LS model infers the coalescent and recombination events (i.e., the hidden states) that generated the sampled sequences using the genetic data (i.e., the observable states). In other words, the LS model reconstructs a target haplotype of an individual at a given locus, the hidden state, as a mosaic of all the remaining haplotypes in the dataset. This is an imperfect mosaic because the LS model considers mutation rate, allowing some variation between the reference and target haplotypes. Although the LS model is a significant step in inferring coalescent and recombination events at the whole genome level, the number of generated states remains high, which means it cannot be used on large cohorts (Lunter, 2019; M. D. Rasmussen et al., 2014).

### 2.3.4. Estimating genome-wide genealogies for large datasets

While full likelihood methods that incorporate all the information stored in ARGs may be up-and-coming, they, unfortunately, remain computationally intractable for large genomic datasets. However, several methods have been developed that infer ARGs using various approximations and assumptions (Kelleher et al., 2019; M. D. Rasmussen et al., 2014; Speidel, Forest, Shi, & Myers, 2019), which can be used to quantify selection or other population genomics statistics. Some methods even infer population genomics statistics under a coalescent model without generating trees (Albers & McVean, 2020).

#### 2.3.4.1. RELATE: inferring marginal trees

The RELATE software addresses the ARG computing problem by inferring marginal coalescence trees instead of full ARG. First, RELATE estimates the tree topology at the first SNP at the 5' end of a chromosome using a hierarchical clustering algorithm that will define the order of coalescence event between haplotype pairs from distance matrices calculated with a modified Li and Stephens HMM (Li & Stephens, 2003). To reduce computing, RELATE does not estimate a tree at every SNP but instead builds a new tree only if a mutation on a SNP cannot be mapped on a unique branch of the previous tree (marginal tree). Once trees have

been assigned to each SNP, RELATE estimates the branch lengths jointly with the coalescence rate with a Markov Chain Monte Carlo (MCMC) algorithm.

RELATE can perform different downstream analyses using these marginal trees as input. It can, for example, estimate within-population and cross-population coalescence rates, two estimates used to understand populations' separation history. RELATE also includes a selection test based on estimating the speed of spread of the lineage carrying the derived allele relative to the non-carriers lineages.

#### 2.3.4.2. Genealogical Estimation of Variant Age

Genealogical Estimation of Variant Age (GEVA) is a method to estimate the age and origin of genetic variants. This method relies on the fact that, under some given assumptions, the time of mutation events is delimited by the TMRCA of lineages carrying the mutation and the most recent time these carrier lineages coalesce with non-carrier sequences. GEVA will first select random concordant pairs (pairs of sequences carrying the mutation) and random discordant pairs (one sequence carrying the mutation and one sequence that does not). GEVA uses an HMM to estimate the region where the MRCA remains unchanged for concordant and discordant pairs. Specifically, the HMM calculates the distance to the closest recombination event on either side of the focal position along the sequence. The inferred sequence is also used to determine the number of mutations that have occurred on the branch to the MRCA of the haplotype. Secondly, GEVA will infer the pairwise TMRCA between lineages (concordant and discordant) pairs for three probabilistic models that account for mutation rate, recombination rate or both. Finally, information from the pairs is combined within a composite-likelihood framework to obtain an approximate posterior distribution on the age of the derived allele.

#### 2.3.5. CLUES: Inferring selection using marginal trees

Selection tests based on RELATE have several limitations: they use single-point estimates of the branch length and do not take uncertainty into account. Moreover, they assume that the tree is derived from a population of constant size, which is usually not true. The Coalescent Likelihood Under Effects of Selection (CLUES) method is an approximate full-likelihood method for inferring selection that has been developed to overcome these limitations (Stern, Wilton, & Nielsen, 2019). CLUES uses the local tree generated with RELATE containing coalescence times for a SNP of interest to compute the ratio of the likelihood of the SNPs being under selection given a coefficient of selection versus the likelihood of the SNP being under neutrality. CLUES also infers the change in derived allele frequency (DAF) over time for a model with recombination and selection using only modern data. This DAF trajectory is an asset because it allows estimating strength and timing selection from modern data, which previously required genomes from different time periods.

### **3. AIMS OF THE STUDIES**

The settlement of Sahul results from one of the most ancient human migrations of AMH. The paths AMH took across Wallacea, before reaching northern Sahul, are still debated. The location of the entrance point to Sahul and the dynamic of settlements within Sahul are also still areas of investigation. Because they faced new environments, the first settlers encountered new adaptative pressures once they reached New Guinea. This thesis aims to investigate New Guinea populations' genetic and phenotypic diversity using a combination of new whole-genome sequences and phenotypic measurements. The first reference explores genomic models for migrations from Sunda to Sahul and the settlement within New Guinea. The second study concentrates on the chronology of selection events during the migration of AMH across Wallacea and their settlement in northern Sahul using published genomes from Wallacea, New Guinea and the Bismarck Archipelago. The third study compares phenotypic differences between PNG highlanders and lowlanders. The fourth study investigates selection signals in the same PNG highlanders and lowlanders. These studies shed light on New Guinean populations' genetic and phenotypic diversity and evolutionary history.

#### **3.1. Aims of the first study (Ref I)**

The exact migratory routes taken by the initial settlers of Sahul remain unclear (Allen & O'Connell, 2020). In this study, we sought to address multiple scenarios by reconstructing the key genetic events that led to the settlement of New Guinea using a genomic model. We first investigated whether one or several human groups conducted the migration to Sahul and which was the most likely migratory route. We also explored how New Guineans settled the diverse environments of the current island and the dynamics of this settlement. To accomplish this, we sequenced 58 new whole genome sequences sampled in Port Moresby, filling geographical gaps in previous samplings of PNG sequences and providing the first genomic model for the settlement of northern Sahul, considering one or two migrations from Wallacea.

#### **3.2. Aims of the second study (Ref II)**

When AMH migrated through Wallacea and settled in northern Sahul, they had to adapt rapidly to unfamiliar environments. This study aimed to establish the chronology of selection events during this settlement. We analysed selection signals in whole-genome sequences previously published for three populations: Wallaceans, New Guineans, and Bismarck Islanders. We chose these populations due to their relative isolation, as we aimed to identify selection signals specific to each stage of the northern Sahul settlement. We used cross-population selection

tests, PBS, XP-EHH and XP-nSL (Sabeti, 2006; Szpiech & Hernandez, 2014; Yi et al., 2010), to identify unique selection signals in these three populations. By detecting such selection events, we established a relative chronology of the adaptation. We complemented these efforts with GEVA (Albers & McVean, 2020) to compute the coalescence time of the variants under selection to obtain an absolute chronology of these events. Finally, we explored the contribution of Denisovan introgression to the loci under selection.

### **3.3. Aims of the third study (Ref III)**

Multiple populations world-wide have adapted to living at high-altitude and exhibit different phenotypes than their lowlander counterparts. However, despite exposure to hypoxia for the last 20,000 years, limited information is available on the impact of selection and phenotypic variation on PNG highlanders. We hypothesised that PNG highlanders differ from PNG lowlanders in phenotypes that have helped them cope with the lower oxygen availability in the highlands. To test this hypothesis, we explored how 13 newly measured phenotypes related to cardiovascular health, body proportions, and lung capacities differ between PNG highlanders and lowlanders. We also compared our findings to those made in high-altitude populations from the Andes, Ethiopia, and Tibet.

### **3.4. Aims of the fourth study (Ref IV)**

PNG highlanders and lowlanders have faced different environmental challenges since they separated from their shared ancestral population. PNG highlanders have been exposed to low oxygen levels for the last 20,000 years, while PNG lowlanders have encountered a unique pathogenic environment. We hypothesized that PNGs might show distinct selection signals depending on their environmental niche. We further hypothesized that these selection signals would be associated with phenotypic differences defined by Ref III (André et al., 2021). To explore this hypothesis, we sequenced and analysed 128 new whole-genomes from PNG lowlands and highlands. We detected unique selection candidates for PNG lowlanders and highlanders using PBS and XP-EHH selection scores (Sabeti, 2006; Yi et al., 2010). We used CLUES (Stern et al., 2019) to identify the most likely SNP to drive selection signal in each region under selection in PNG highlanders and lowlanders. We looked for associations of these candidate SNPs with phenotypes from the UKBiobank (Pan-UKB team, 2020) or measured in PNGs in Ref III (André et al., 2021).

## 4. MATERIAL AND METHODS

### 4.1. Ref I

This study includes 58 newly sequenced Papua New Guinean participants sampled in Port Moresby. These participants originate from different parts of the country and then increase the geographical coverage of New Guinea, which is under-represented in previously published whole-genome data sets. We combined the new PNG sequences with published genomes from Indonesia (Jacobs et al., 2019), northeast Australia (Mallick et al., 2016), and the Bismarck Archipelago (Vernot et al., 2016), and with genotyping data from North Maluku Islands located on the northern route of Wallacea (Kusuma et al., 2017) and imputed genotyping data from PNG (Bergström et al., 2017). We used fineSTRUCTURE to identify the regional groups within our dataset (Lawson, Hellenthal, Myers, & Falush, 2012). Because our study focuses on migrations movements before the Lapita people reached New Guinea, we masked the Asian genetic ancestry in our dataset using haplotypic information with PCAdmix (Brisbin et al., 2012). We further explored the diversity of our dataset by performing ADMIXTURE (Alexander, Novembre, & Lange, 2009) and PCA analysis, as well as computing the  $F_{ST}$  between each regional group pair. We estimated splits and admixture events in our dataset using TREEMIX. To identify the most likely route taken by the first settlers of Sahul (i.e., southern vs northern route), we computed outgroup  $f_3$ -statistics (Patterson et al., 2012) to show how genetically close New Guinean, northeast Indigenous Australian, and Bismarck Archipelago islander subgroups are to Wallacean subgroups located on the southern or the northern route, using African Yoruba as an outgroup. Additionally, we performed  $f_4$ -statistic (Patterson et al., 2012) to test for gene flow from Wallacean subgroups located on the southern route and Wallacean subgroups located on the northern route to New Guinean, northeast Indigenous Australian and Bismarck Archipelago islander, using African Yoruba as an outgroup again. To explore the likelihood of different migration hypotheses, we tested several models using qpGraph (Patterson et al., 2012). We tested models that consider migrations from Sunda to Sahul along either the southern or the northern route and different possibilities of admixture between Wallacean and New Guinean groups. We also modelled demographic scenarios with one or two migrations from Sunda to Sahul. To improve the resolution of our demographic models, we incorporated a group from the Bismarck Archipelago and tested its relationship with New Guineans and northeast Indigenous Australians. Finally, to determine the most likely entry point into New Guinea, we used qpGraph to determine the best scenario for the settlement within New Guinea, fitting data from the five New Guinean groups and African Yoruba outgroups, with or without admixture events. To refine the various scenario models described in this paper, we also inferred the effective population size of these groups and the divergence dates between them using MSMC2 (Schiffels & Durbin, 2014).

## 4.2. Ref II

All data used in this publication has been obtained from the literature. It used whole genome sequences from 239 Oceanians. In addition to the 58 PNG sequences from Ref I (Brucato et al., 2021), this study also included populations from Wallacea, New Guinea, and the Bismarck Archipelago (Jacobs et al., 2019; Malaspina et al., 2016; Mallick et al., 2016; Vernot et al., 2016). To increase the sample size of the New Guinean population, we also included previously published imputed genomes from PNG (Bergström et al., 2017). We added 330 whole-genome sequences from non-Oceanian populations (Jacobs et al., 2019; Mallick et al., 2016) to compare our dating results from GEVA with the initial publication. We used GEVA v1.beta (Albers & McVean, 2020) (Genealogical Estimation of Variant Age; described in the literature review) to estimate the distribution of coalescence ages of SNPs. We performed XP-EHH and XP-nSL analysis using Selscan v.1.3.0 (Szpiech & Hernandez, 2014). To detect signatures of selection unique to New Guineans, Bismarck Islanders, and Wallaceans, we generated these selection scans for three target and reference population pairs: New Guinea compared to Wallacea, Bismarck Archipelago compared to Wallacea and the Bismarck Archipelago compared to New Guinea. PBS scores (Yi et al., 2010) were generated from  $F_{ST}$  distances inferred with vcfTools v0.1.15 (Danecek et al., 2011). We considered New Guinea or the Bismarck Archipelago as the target population, Wallacea as the reference population and used African genomes from the SGDP dataset as an outgroup. We also perform a PBS analysis defining the Bismarck Archipelago as the target population, New Guinea as the reference population and Africa as the outgroup. Finally, we combined the XP-EHH, XP-nSL and PBS for each SNP in a Fisher Score (Lopez et al., 2019). We estimated the significance of enrichment of selection signals in 1000 years time frames using coalescent time estimates from GEVA for the SNPs with outlier Fisher Score. Finally, we estimated the archaic introgression in the genomes from Wallacea, New Guinea and the Bismarck Archipelago using Skov's HMM method (Skov et al., 2018), which does not require an archaic reference. We also analysed adaptive introgression using the Q95 statistics (Choin et al., 2021; Racimo et al., 2015).

## 4.3. Ref III

This paper relies on phenotypes measurements performed on PNG highlanders living in Mount Wilhelm between 2,300 and 2,700 m a.s.l. and PNG lowlanders living on Daru island around 100 m a.s.l. Phenotypes were collected on healthy adults. The studied phenotypes were standing height, weight, body mass index (BMI), waist circumference, minimal and maximal chest depth, haemoglobin concentration, heart rate, and diastolic and systolic blood pressure. Spirometer measurements reflecting lung capacities were also recorded: the volume of air

that is forcibly blown out in the first second after full inspiration (forced expiratory volume in 1 second, FEV1), the maximal flow achieved during the maximal forced expiration initiated at full inspiration (peak expiratory flow, PEF) and the volume of air that is forcibly blown out after complete aspiration (forced vital capacity, FVC). To consider only PNG highlanders with minimal lowland admixture and reciprocally, we excluded all individuals with at least one parent or grandparent not originating from the same altitude level. After this step, we kept 70 highlander and 86 lowlander individuals. We corrected all the phenotypes for age and sex by computing residuals for the corresponding multilinear regression. We also regressed minimal and maximal chest depth, waist circumference, weight and spirometer measurements for height that could be a covariate to these phenotypes. Finally, we compared the residuals of the phenotypes between PNG highlanders and lowlanders with a Mann-Whitney U test, a non-parametric test. The measured phenotypic traits analysed in this study are reportedly adaptive in other highlander groups worldwide (Cynthia M. Beall, 2013; Bigham & Lee, 2014). We then compared our results with previous work on other highlander populations.

#### 4.4. Ref IV

In this publication, we used newly sequenced whole genomes from three villages in Mount Wilhelm located in PNG highlands (n=54, 2300 to 2700 meters a.s.l.) and from Daru Island (n=74, 100 meters a.s.l.). We aimed to detect genomic regions showing signs of positive selection in PNG highlanders and lowlanders. To achieve this, we used three different scores: XP-EHH (Sabeti et al., 2007), PBS (Yi et al., 2010), and a Fisher Score, which combines XP-EHH and PBS scores (Lopez et al., 2019). We used PNG highlanders as the target population and PNG lowlanders as the reference population and vice versa for each score. We included Yorubas from the 1000 Genomes project (The 1000 Genomes Project Consortium et al., 2015) as the outgroup when computing the two PBS scores. With this approach, we identified signatures of selection unique to PNG highlanders and lowlanders. We considered ten genomic regions of interest with the highest scores for each score. Next, we inferred ancestral recombination graphs for the full dataset, using the two PNG populations and the 58 sequences from Ref I (Brucato et al., 2021) and Asian, European and African genomes from The 1000 Genomes Project (The 1000 Genomes Project Consortium et al., 2015) with RELATE (v1.1.8) (Speidel, Forest, Shi, & Myers, 2019). We extracted the local tree for each SNP of the genomic regions under selection from PNG highlanders and lowlander subtrees. We used these local trees as input for CLUES (Stern et al., 2019). CLUES assigned a likelihood ratio (logLR) which indicates the support of the non-neutral model for each tested SNP.

The log LR was computed five times for each SNP, and the average of the five runs was taken. To decide between the top five SNPs, the logLR was generated 50 more times, and the SNP with the highest average logLR (a.k.a. candidate

SNP) was considered the most likely SNP to drive selection within the region of interest. Defining the candidate SNPs allowed us to look for the associated phenotypes in the UK biobank (Pan-UKB team, 2020). We extracted the p-value and the beta for European ancestry from the UK biobank summary statistics for the candidate SNPs and 1,931 phenotypes with more than 10,000 samples. When candidate SNPs were not listed in the UK biobank, we looked for summary statistics for the closest SNP from a 1kb upstream and 1kb downstream region. We considered the association of a candidate SNP with a phenotype of interest if the  $\log_{10}(\text{p-value})$  is lower than  $-11.29$  to correct for multiple testing. We chose this threshold based on the standard significance p-value threshold in GWAS of  $5e-8$  (Dudbridge & Gusnanto, 2008) and adjusted it for the 1931 different phenotypes we used in the summary statistics following the Bonferroni correction guideline (Shaffer, 1995).

Moreover, we compared these associations from the UK biobank with associations found for phenotypes measured directly in this publication's two studied PNG populations and published in Ref III (André et al., 2021). We tested if the candidate SNPs were related to any PNG phenotypes using a statistical model called Linear Mixed Model (LMM) with the Genome-wide Efficient Mixed Model Association (GEMMA) (v0.98.4) software (X. Zhou & Stephens, 2012). To increase our sample size of genotypes and phenotypes, we performed these association tests using PNG highlanders, PNG lowlanders, and PNG samples from Port Moresby published with Ref I (Brucato et al., 2021). We incorporated a centred relatedness matrix generated with GEMMA for all the PNG sequences into the LMM to correct for population stratification. We adjusted the p-values of the SNPs to account for multiple tests using the Benjamini-Hochberg procedure (Benjamini & Hochberg, 1995). We grouped the phenotype into the five categories of highly correlated phenotypes defined in Ref III (André et al., 2021): body proportion group (height, weight, waist circumference, BMI), pulmonary function group (chest depth, spirometer measurements), haemoglobin concentration, blood pressure (diastolic and systolic blood pressure) and heart rate. We adjusted the p-value threshold with Bonferroni correction for the five independent phenotype groups tested ( $\text{p-value}_{\text{adjusted}}=0.05/5$ ) to determine statistical significance.

As an additional effort to decipher the function of the candidate SNPs, we looked for significant eQTLs for each candidate SNP using the Genotype-Tissue Expression (GTEx) Portal (Lonsdale et al., 2013). Moreover, we used the Ensembl Variant Effect Predictor (VEP) (McLaren et al., 2016) on the regions under selection to detect missense variants in these regions.

Finally, we used haplostrips (v1.3) (Marnetto & Huerta-Sánchez, 2017) to explore similarities between PNG haplotypes and archaic haplotypes for the genomic regions under selection in PNG highlanders and lowlanders. We also analysed archaic allele frequencies in the PNG from the SGDP dataset (Mallick et al., 2016) in the regions of interest with introgressed haplotypes in PNG highlanders and lowlanders.

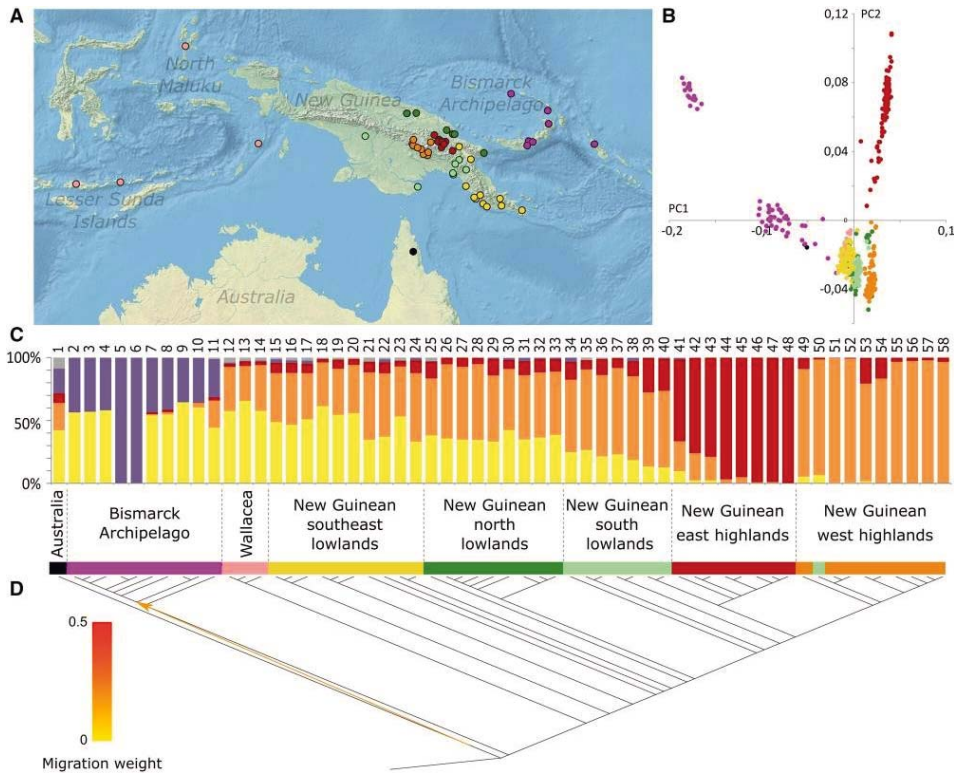
## **5. RESULTS AND DISCUSSION**

This section summarises the main results and discussions for the four scientific publications constituting this dissertation's original part. The following is a compressed summary of the main results and discussions. More detailed information can be found in the original articles and their respective supplementary information.

### **5.1. Models for northern Sahul settlement based on genomic data (Ref I)**

#### **5.1.1. Genetic structure characterization:**

We identify our dataset's main regional genetic groups using fineSTRUCTURE (Lawson et al., 2012). We found that individuals in our dataset clustered into seven main groups, corresponding to their respective geographical locations, including the New Guinean southeast lowlands, New Guinean north and south lowlands, New Guinean east highlands, New Guinean west highlands, northeast Australia, Bismarck Archipelago, and Wallacea. PCA, Admixture and  $F_{ST}$  results reflect a strong geographical structure of the human genetic diversity in Sahul. However, they also indicate a relative genetic continuity between New Guinea and Wallacea, implying that the latter was not an insurmountable barrier (Figure 11B).



**Figure 11 : Genetic diversity of individuals from New Guinea, Wallacea, the Bismarck Archipelago, and northeast Australia.** (A) Geographic locations of all 58 genetic subgroups defined by the fineSTRUCTURE analysis. (B) Principal component analysis representing 26.7% of the total variability of all individual genomes from the 58 subgroups, masked for Asian genetic ancestry. The two first components are represented (PC1 and PC2). (C) ADMIXTURE plot for eight genetic components summarized by genetic subgroups (based on  $K = 8$ ). The grey component combines four components mainly present in African and Eurasian groups. (D) TREEMIX plot for all 58 subgroups. All four panels used the same colour scheme to identify the 58 genetic subgroups. Reprinted from Figure 1 (Brucato et al., 2021), licensed under Creative Commons CC BY license. Copyright © 2021, Brucato et. al.

### 5.1.2. Identification of gene flow

We obtained an optimal TREEMIX model (Pickrell & Pritchard, 2012) when including five gene flows, one of which is from ancestors of northeast Indigenous Australians to a group ancestral to most Bismarck Archipelago islanders. (Figure 11D). This gene flow might explain the component shared between Indigenous Australians and Bismarck Archipelago Islanders observed in the ADMIXTURE plot (Figure 11C). The tree also shows that all the New Guinean subgroups branch together while its root lies in the genetic diversity of New Guinean southeast lowlanders (Figure 11D). Additionally, we observed that subgroups from the New Guinean north and south lowlands constituted two distinct genetic clusters that had genetically diverged from the New Guinean southeast lowlanders (Figure 11D). These results suggest that New Guinean north and south lowlanders originated from separate migrations that were initiated from the southeast region of New Guinea.

### 5.1.3. Modelling the genetic scenario of the migration in Wallacea

$F_3$ -statistics (Patterson et al., 2012) results show that all subgroups from New Guinea, northeast Australia, and the Bismarck Archipelago are equally distant to any Wallacean subgroups located on the southern route to Sahul. However,  $f_3$ -statistics increase when using a Wallacean subgroup located on the northern route instead of Wallacean subgroups from the southern route. These results suggest a stronger genetic link of the Wallacean subgroup located on the north route with all studied subgroups than those on the southern route.  $F_4$ -statistics (Patterson et al., 2012) support that most populations from New Guinea and the Bismarck Archipelago are significantly genetically closer to the Wallacean population located on the northern Route ( $f_4(\text{Yoruba, X; southern Wallacea, northern Wallacea}) > 0.001$ ,  $Z \text{ score} > 3$ ). Finally, this genetic link is particularly pronounced in New Guineans:  $f_3$ -statistics and  $f_4$ -statistics values were higher and more significant for New Guineans with north Wallacean subgroups than for the Bismarck Archipelago and northeast Australia.

Two non-exclusive hypotheses might explain the more important genetic link between the north Sahul populations and northern Wallacea subgroups. The first hypothesis is that the northern route was the most likely path for the settlement of Sahul. Alternatively, there may have been secondary gene flows between north Wallacea and northern Sahul groups. To explore the likelihood of these hypotheses, we tested multiple migration and admixture scenarios using qpGraph (Patterson et al., 2012). We obtained two models fitting the data (Figure 12). The first model considers a unique migration from Sunda to Sahul ( $Z = -2.558$ ). In this model, New Guineans and Indigenous Australians share a common ancestor more recently than Wallaceans. The split from Wallaceans is followed by two important gene flows from New Guinea to Wallacea.

The second model fitting the data displays two migrations from Sunda to Sahul ( $Z = 2.337$ ). In this model, northern Indigenous Australians and New Guineans descend from groups that have migrated to Sahul independently. These migrations are followed by a gene flow of the ancestors of New Guineans to northern Wallacean groups. This model suggests that New Guineans' ancestors have taken the northern route, which is consistent with previous modelling studies (Bird et al., 2019; Bradshaw et al., 2019; Kealy et al., 2018).

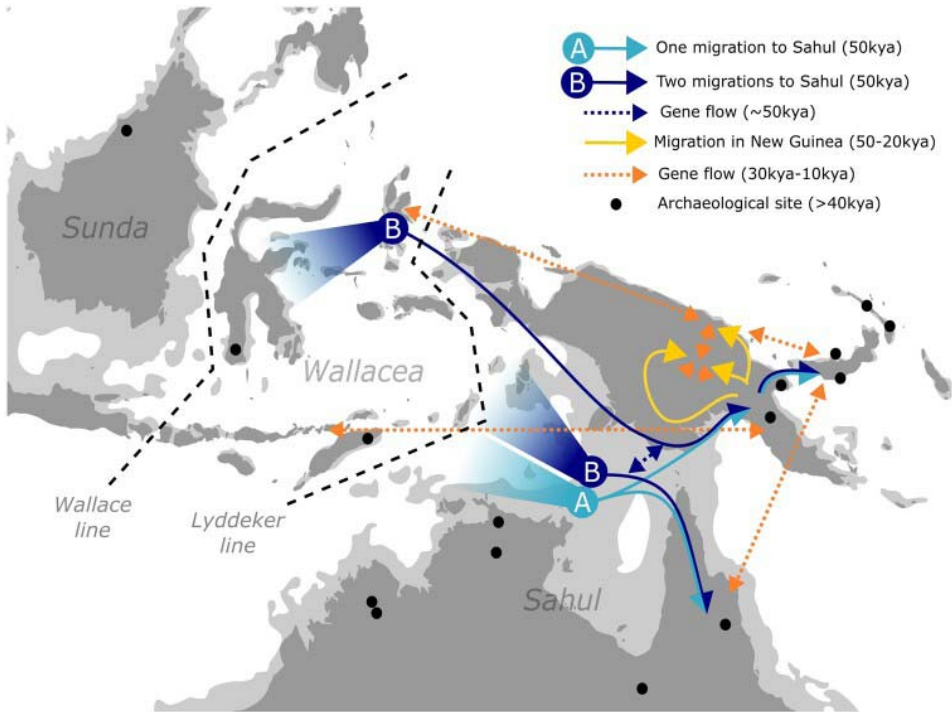
#### 5.1.4. Modelling the genetic scenario of the settlement of Sahul

We improved the previously described qpGraph models for the settlement of Sahul by incorporating a subgroup from the Bismarck Archipelago into the previous two scenarios. For both models, with one (model A,  $Z=-2.570$ ) or two migrations (model B,  $Z=2.301$ ) to Sahul, we obtained models fitting the data when the Bismarck Archipelago group results from a separate admixture event between ancestral New Guineans and northeast Indigenous Australian (Figure 12) which correspond to the gene flow between ancestral indigenous Australians group and ancestral Bismarck Archipelago group detected by Treemix. This might again support the continuity hypothesis between groups from Wallacea to New Guinea but stronger genetic barriers between New Guinea and Australia and around the Bismarck Archipelago.

#### 5.1.5. Modelling the genetic scenario of the settlement of New Guinea

We computed qpGraph models to determine the best model for the settlement of New Guinea using the five New Guinean groups (southeast lowlanders, south lowlanders, north lowlanders, east highlanders, and west highlanders) and the African Yoruba outgroup. The unique model fitting our data ( $Z$  score  $=-0.06$ ) presents the southeast lowlands as the most likely entry point to northern Sahul. When adding the west and east highlander groups, the best-fitting model places west highlanders with New Guinean south lowlanders ( $Z$  score  $= 2.759$ ) (Figure 12).  $F_3$ -statistics show a closer link between New Guinean west highlanders and New Guinean south lowlanders. In addition, New Guinean east highlanders are almost at a similar genetic distance to New Guinean south and north lowlanders, which suggests that New Guinean east highlanders came from a place between south and north lowlands. This final model identifies the southeast lowlands as the entry point into New Guinea. Interestingly, New Guinean southeast lowlands correspond to the location of the oldest known archaeological site in New Guinea (45–49kya) (Summerhayes et al. 2010). This supports an initial migration via the ancient plain currently under the Arafura Sea. Our results indicate that this initial migration was followed by two migrations to the south and north lowlands. Later, the New Guinean highlands were settled by two separate migratory waves: one migration from the east lowlands to the east highlands and another from the south

lowlands to the west highlands. We completed this scenario of settlement of New Guinea using MSMC2 (Schiffels & Durbin, 2014). We found a similar divergence time between New Guinean populations (7.6–12.8kya) than previously published results (Bergström et al., 2016). These relatively recent divergence times probably reflect long-term contact between New Guinean populations after the initial settlement.



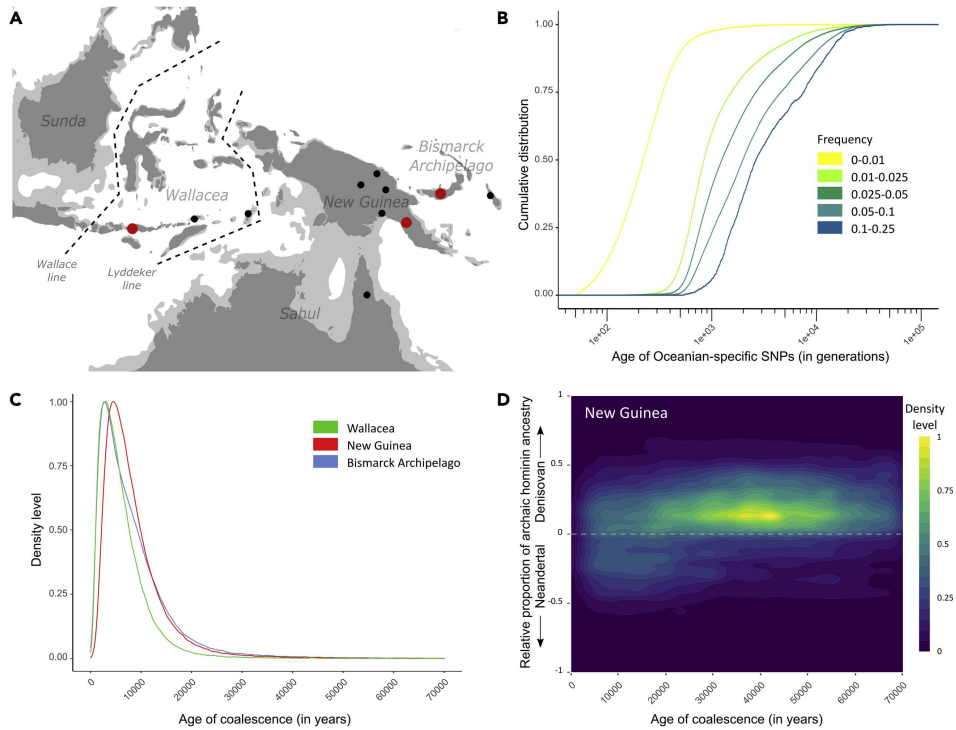
**Figure 12: Genomic scenario of the human dynamics in north Sahul during the Upper Pleistocene.** Light blue arrows represent the settlement of north Sahul following a scenario with one migration to Sahul (model A). The dark blue arrows represent the settlement of north Sahul following a scenario with two migrations to Sahul (model B). The dotted dark blue arrow represents a gene flow following a scenario with two migrations to Sahul (model B). Yellow arrows represent the settlement of New Guinea. The dotted orange arrows represent gene flows after the initial settlement of Sahul. The black dots represent approximate locations of archaeological sites older than 40 ka (O’Connor, 2007; Summerhayes et al., 2017). The dashed black lines represent ecological lines in Wallacea. The dark grey areas represent the current land area, with light grey areas representing the estimated land area around 50,000 years ago based on paleogeological reconstructions (Coller, 2009). Reprinted from Figure 4 (Brucato et al., 2021), licensed under Creative Commons CC BY license. Copyright © 2021, Brucato et. al.

## 5.2. Time of selection in Oceanian genomes (Ref II)

### 5.2.1. Dating the Oceanian genetic diversity

First, we showed that the age of coalescence estimates obtained using GEVA on SGDP whole-genomes from Africa, Asia, and Europe, strongly correlated with the previously published estimates (Albers & McVean, 2020; Mallick et al., 2016). To limit the influence of recent Asian ancestry, we defined the ages of coalescences only for SNPs specific to New Guinea, Bismarck Archipelago or Wallacea populations by excluding SNPs found in the other populations in our dataset with several Asian genomes. The median age of the SNPs specific to Oceanians with a frequency of 0.25 (78,795 Years Before Present (YBP) IQR: 47,383-225,075 YBP, Figure 13B) is older than the median age of private SNPs at the same frequency in other non-African populations. This result is coherent with previous studies, including Ref I (Brucato et al., 2021), on the divergence time of Oceanians from Africans and reflects the extended time these populations have been isolated from other continents (Malaspinas et al., 2016; Pagani et al., 2016).

Most SNPs private to populations from New Guinea, Bismarck Archipelago or Wallacea appeared in the last 10,000 years (Figure 13C), which might correspond to the demographic expansion observed worldwide during the Last Glacial Maximum era (Bergström et al., 2020). We observed that this increase in diversity occurs slightly earlier in New Guineans. This earlier increase in diversity in New Guineans might result from their early farming practices.



**Figure 13: Distribution of coalescence ages in Oceanian genomes.** (A) Map of analyzed populations. Red dots localize the three studied Oceanian regional groups in Wallacea (Flores Island), New Guinea (Papua New Guinea southeast coast), and the Bismarck Archipelago (New Britain). Black dots represent other groups included in our dataset. (B) Cumulative distribution of coalescence ages estimated with GEVA for Oceanian-specific SNPs binned by frequency. (C) Density plot of coalescence ages for SNPs specific to each Oceanian regional group. (D) Density plot of coalescence ages for SNPs present in archaic fragments in New Guinean genomes. The relative proportion of archaic ancestry, estimated with Skov’s HMM method, is calculated as  $(\text{Denisovan ancestry} - \text{Neanderthal ancestry}) / (\text{Denisovan ancestry} + \text{Neanderthal ancestry})$ . Reprinted from Figure 1 (Brucato et al., 2022), licensed under Creative Commons CC BY license. Copyright © 2022, Brucato et. al.

### 5.2.2. Relative and absolute dating of the signal of selection

We defined SNPs significantly under selection in New Guineans and Bismarck islanders with a Fisher Score (Fxp) (Lopez et al., 2019) combining XP-EHH (Sabeti et al., 2007), XP-nSL (Szpiech & Hernandez, 2014) and PBS (Yi et al., 2010). We used Wallaceans as the reference population and African genomes from SGDP as the outgroup population for PBS. Because we used Wallaceans as the reference population, the Fisher Score of New Guineans and Bismarck Islanders characterise genomic regions under selection present only in New Guineans or Bismarck Islanders, respectively, but not in Wallaceans. In other words, we used Wallaceans as the reference population to mask signatures of selection in New Guineans and Bismarck islanders dating before the initial settlement of north Sahul. We used this approach to define a relative timing for selection between the crossing of Wallacea and the settlement of northern Sahul (Figure 14C, D). We also performed the absolute dating of the SNPs under selection using GEVA (Figure 14A, B).

When coupling these two dating approaches, we observed a significant enrichment of selection signals for SNPs dating between 52,000 and 54,000 YBP in both New Guinean and Bismarck Archipelago genomes ( $Z > 6$ ,  $p$ -value  $< 10^{-9}$ , Figure 14). This timing coincides with the supposed time of the first settlement of Sahul (Summerhayes et al., 2017). This convergence in the time period with enrichment for SNPs under selection in New Guineans and Bismarck, despite the two populations being highly divergent, would suggest that common ancestors of New Guineans and Bismarck Archipelago Islanders faced strong selection pressure around this time period.

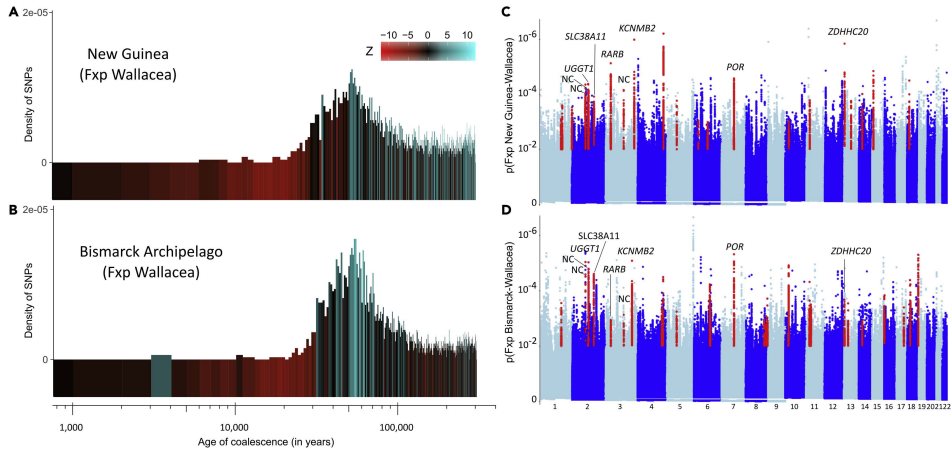
The results of the selection scans on the genomes of New Guineans and Bismarck Islanders suggest that selection during the settlement of Sahul mainly affected genes related to the immune system and diet. One of the strongest selection signal in both populations overlaps with the protein-coding gene *ZDHHC20*, which is involved in antiviral response (McMichael et al., 2017). Such selection signals in New Guineans and Bismarck Islanders might result from an ancient viral pathogenic pressure.

Other significant signals of selection in the genomes of New Guineans and Bismarck Islanders overlap genes related to food intakes, such as *RARB*, *KCNMB2*, and *POR*, suggesting that diet (e.g., intake of Vitamin A and folic acid) was a major selective force. The diet of modern New Guineans and Bismarck Islanders considerably relies on the local plant. Selection acting on the first settlers of northern Sahul probably favoured genes supporting the use of the local plant resources of these new territories.

We also observed a significant enrichment for SNPs with signal of selection unique to Bismarck Islanders between 42,000 and 43,000 YBP ( $Z > 6$ , Figure 14B) when using this population as the target population and New Guineans or Wallaceans as the reference population. This corresponds to the first settlement of the archipelago (Summerhayes et al., 2017).

Moreover, New Guinean and the Bismarck Islanders, but not the Wallaceans, have a significantly lower number of selection signals for SNPs dating between 10,000 and 30,000 YBP ( $Z < -6$ ,  $p\text{-value} < 10^{-9}$ , Figure 14A, B). This lower enrichment would be specific to the descendants of the first settlers of north Sahul and would correspond to the reduced demographic dynamics during the Last Glacial Maximum (Pedro et al., 2020; Summerhayes et al., 2017)

Overall, these results show that the first settlers of northern Sahul were exposed to strong selective pressure, likely associated with exposure to a new diet, rapidly after they reached New Guinea and Bismarck.



**Figure 14: Distribution of coalescence ages for SNPs under selection in the genomes of New Guineans and Bismarck Archipelago Islanders. (A and B)** Density plot of coalescence *ages* for all significant Fxp signals of selection in New Guinea (A) and the Bismarck Archipelago (B), using Wallacea as reference ( $p < 0.01$ ), binned by 1,000 years. The blue-red gradient represents the Z score of enrichment of SNPs with a significant Fxp signal of selection in comparison to the rest of the genome (resampled 1,000 times) for each 1,000-year bin. (C and D) Manhattan plots of the Fxp p-values for each SNP in the analyses: (C) Fxp New Guinea vs Wallacea and (D) Bismarck Archipelago vs. Wallacea. Red dots represent variants showing significant Fisher scores in 100kb windows enriched for significant scores ( $>0.7$ ). Gene names represent loci including at least one SNP with a coalescence age between 52,000 and 54,000 YBP. Reprinted from Figure 2 (Brucato et al., 2022), licensed under Creative Commons CC BY license. Copyright © 2022, Brucato et. al.

### 5.2.3. Introgression

All New Guinean, Bismarck and Wallacean populations show a higher rate of Denisovan than Neanderthal introgression. Most SNPs introgressed from Denisovan have a coalescence time falling between 35,000 and 45,000 YB. This coalescence age is in the time frame of the admixture events with Denisovans previously reported for the Oceanian region (30,000–46,000 YBP) (Jacobs et al., 2019; Malaspinas et al., 2016). When refining our approach, we noticed that many SNPs introgressed from Denisovans and private to New Guineans have an even later coalescence time of around 42,000 YBP. Nonetheless, most archaic SNPs in any of these three Oceanian populations have a coalescence time of about 10,000 years, suggesting that they were not inherited from Neanderthals or Denisovans.

### 5.2.4. Introgression and selection

Although the SNPs significantly under selection in New Guineans, Bismarck Islanders, and Wallaceans did not show an overall enrichment for Denisovan ancestry ( $Z < 2$ ,  $p$ -value  $> 10^{-2}$ ), we found the highest proportion of Denisovan ancestry in SNPs under selection between 52,000–53,000 YBP in New Guineans and 39,000–40,000 YBP in Bismarck Islanders. These results suggest that the SNPs under selection were present in Denisovan introgressed fragments during the main phases of settlement in the New Guinea region.

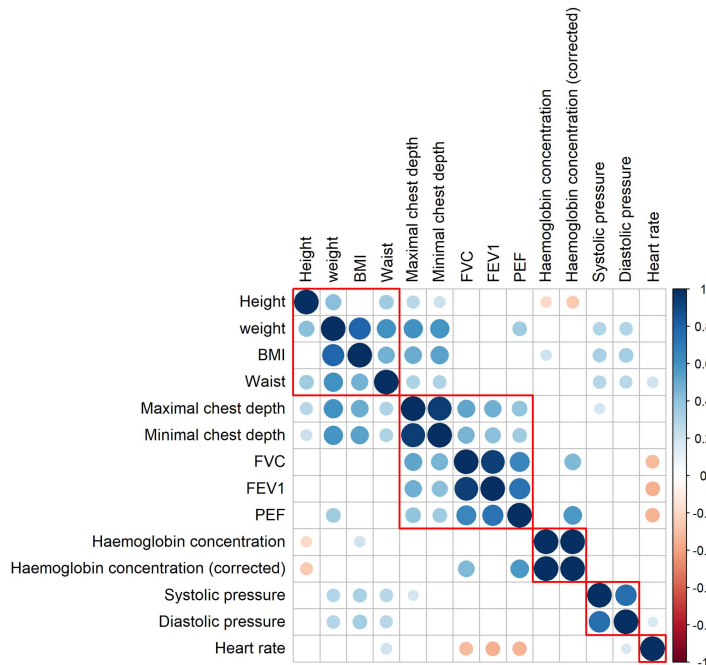
Of the loci enriched for Denisovan ancestry and under selection in Wallacea, New Guinea and Bismarck populations (e.g., *TNFAIP3*, *IFNGR1*, the *GBP* locus and the *SIGLEC* locus), most have already been reported previously (Jacobs et al., 2019; Sankararaman et al., 2016; Vernot et al., 2016). Interestingly, none of the three populations shows an elevated Fisher score for any of these loci, suggesting that their selection predated the initial settlement of northern Sahul.

It is to be noted that we found a strong signal of selection in a locus enriched for Denisovan ancestry and overlapping with the gene *RARB*. The gene *RARB* encodes a receptor which is a vitamin A derivative and is involved in adaptive intestinal immunity (Bang et al., 2021; Mora et al., 2006). This suggests that diet and, more specifically, the intake of vitamin A were major selective forces.

## 5.3. Phenotype differences between PNG highlanders and lowlanders (Ref III)

### 5.3.1. Multitesting correction

We compared 13 phenotypes between PNG lowlanders and highlanders using the Mann-Whitney U test. Because we were performing multiple comparisons, which multiple the risk of false positive results, we adjusted the significance threshold of 0.05 for the number of independent tests performed following the Bonferroni correction to control for type I error. Among the 13 studied phenotypes, some were highly correlated, meaning that tests for the correlated phenotype were not independent. Adjusting the p-value threshold for the 13 phenotypes would lead to a loss of power (Shaffer, 1995). We defined five groups of highly correlated phenotype using the Spearman correlation coefficient (Figure 15): Body proportion group (height, weight, waist circumference, BMI), Pulmonary function group (chest depth, spirometer measurements), haemoglobin concentration, blood pressure (diastolic and systolic blood pressure) and heart rate. We considered that differences between PNG highlanders and lowlanders were significant if the p-value was lower than 0.05 after Bonferroni correction for five independent tests giving an adjusted p-value of 0.01.



**Figure 15: Correlation matrix between the studied phenotype measurements.** Spearman correlation was used. Only significant correlations (p-value <0.05) are shown. Five groups of correlated phenotypes are enclosed in red boxes. Reprinted from Figure S1 (André et al., 2021), licensed under Creative Commons CC BY license. Copyright © 2021, André et. al.

### 5.3.2. Exploring phenotypic differences between PNG highlanders and lowlanders

PNG highlanders are significantly shorter (mean: 164.6 cm [95% CI 163.21–166.03]) than lowlanders (mean: 168.8 cm [95% CI 166.84–170.72]) while having significantly deeper chests ( $p\text{-value}_{depth\ min} = 2.37e^{-5}$ ,  $p\text{-value}_{depth\ max} = 3.20e^{-4}$ ). Both these phenotypes have been shown to differ in a similar direction between Andean, Tibetan and Ethiopian highlanders and the respective lowlander population they are traditionally compared with (Table 1) (Brutsaert et al., 1999; Droma et al., 1991; Eichstaedt et al., 2015; G. A. Harrison et al., 1969; Mueller et al., 1978; Rupert & Hochachka, 2001; Weitz, Garruto, Chin, & Liu, 2004). The combination of larger chest size and shorter height has been associated with enhanced lung capacities in Tibetans and Andeans (Hall & Guyton, 2011). Similarly, we also observed a significantly higher FVC ( $p\text{-value} = 0.008$ ), a proxy for lung volume, in PNG highlanders (mean: 4.09 L [95% CI 3.78–4.40]) in comparison with PNG lowlanders (mean: 3.32 L [95% CI 3.02–3.63]). The significantly increased pulmonary volume observed in highlanders from Papua New Guinea may suggest that they have adapted to lower oxygen availability in the highlands. We also reproduced results from another study showing significantly greater haemoglobin levels ( $p\text{-value} = 5.51e^{-4}$ ) in PNG highlanders (mean: 14.45 g/dl [95% CI 13.84–15.06]) than in PNG lowlanders (mean: 12.63 g/dl [95% CI 12.07–13.20]) (Senn et al., 2010). We further show that this observation remains true ( $p\text{-value} = 3.36e^{-4}$ ) even when excluding individuals suffering from anaemia following the WHO standard cut-offs (Department of Nutrition for Health and Development (NHD), 2011; Silubonde et al., 2020). This difference then does not result from the difference in malaria incidence between the highlands and the lowlands. While Andean and Ethiopian highlanders also display elevated haemoglobin levels, Tibetans show a reduced haemoglobin concentration. Moreover, high haemoglobin concentration is also observed in acclimatised lowlanders living at high-altitude and is associated with increased blood viscosity, which is maladaptive in the long term. An increased level of haemoglobin concentration is observed in individuals born with low-altitude ancestry and living at high-altitude and facing lower levels of oxygen saturation caused by hypoxia. Surprisingly, PNG highlanders do not differ from lowlanders for any cardiovascular phenotypes. Because native lowlanders usually show increased heart rate and blood pressure at high-altitude (Brito et al., 2007), the absence of such an increase in PNG highlanders may be another adaptive trait counteracting the detrimental effect of hypoxia.

**Table 1 : Phenotypic comparison between PNG highlanders and lowlanders.** Modified from Table 1 (André et al., 2021), licensed under Creative Commons CC BY license. Copyright © 2021, André et. al.

	PNG lowlanders	PNG highlanders	Mann-Whitney U test residuals		
	N= 86	N= 70	raw data	age, sex	age, sex, height
	Mean [95% CI]	Mean [95% CI]			
<i>Body proportions</i>					
<b>Height (cm)</b>	<b>168.8</b> [166.84–170.72]	<b>164.6</b> [163.21–166.03]	<b>0.001*</b>	<b>0.001*</b>	–
Weight (kg)	68.46 [65.86–71.15]	64.82 [63.2–66.53]	0.137	0.198	0.87
BMI (m <sup>2</sup> /kg)	24.04 [23.19–24.97]	24 [23.33–24.76]	0.661	0.425	–
<b>Waist circumference (cm)</b>	<b>91.77</b> [89.44–94.16]	<b>83.54</b> [81.99–85.17]	<b>&lt;.001*</b>	<b>&lt;.001*</b>	<b>0.001*</b>
<i>Respiratory capacities</i>					
<b>Minimal chest depth (cm)</b>	<b>19.16</b> [18.66–19.68]	<b>19.91</b> [19.43–20.41]	<b>0.033</b>	<b>&lt;.001*</b>	<b>&lt;.001*</b>
<b>Maximal chest depth (cm)</b>	<b>20.50</b> [20.00–21.01]	<b>21.01</b> [20.47–21.56]	<b>0.146</b>	<b>0.003*</b>	<b>&lt;.001*</b>
Forced vital capacity (FVC) (L)	3.32 [3.02–3.63]	4.09 [3.78–4.40]	0.005§	0.020§	0.008
Forced expiratory volume after 1 second (FEV1) (L)	2.72 [2.45–3.00]	3.31 [3.01–3.61]	0.008§	0.177§	0.124
Peak expiratory flow (PEF) (L/min)	6.09 [5.28–6.92]	6.32 [5.39–7.34]	0.747§	0.638§	0.962
<i>Circulatory system</i>					
<b>Haemoglobin concentration (g/dL)</b>	<b>12.63</b> [12.07–13.20]	<b>14.45</b> [13.84–15.06]	<b>&lt;.001*</b>	<b>&lt;.001*</b>	–
<b>Haemoglobin concentration** (g/dL)</b>	<b>14.81</b> [14.31–15.33]	<b>16.07</b> [15.65–16.49]	<b>0.001§</b> *	<b>&lt;.001§</b> *	–
Systolic pressure (mmHg)	131.76 [126.14–137.23]	123.94 [117.48–129.93]	0.071	0.169	–
Diastolic pressure (mmHg)	85.29 [82.33–88.26]	81.11 [77.21–84.99]	0.156	0.196	–
Heart rate (bpm)	67.77 [65.49–70.09]	64.38 [61.67–67.2]	0.042	0.03	–

Significant results are in bold. \*: Significant Mann-Whitney U test with Bonferroni correction for 14 multiple tests (adjusted p-value = 0.004). §: data corrected to avoid ties. \*\*: People suffering from anaemia were removed following the World Health Organization (WHO) standard cut-offs (exclusion of non-pregnant women < 12g/dl, men <13g/dl) (Department of Nutrition for Health and Development (NHD), 2011).

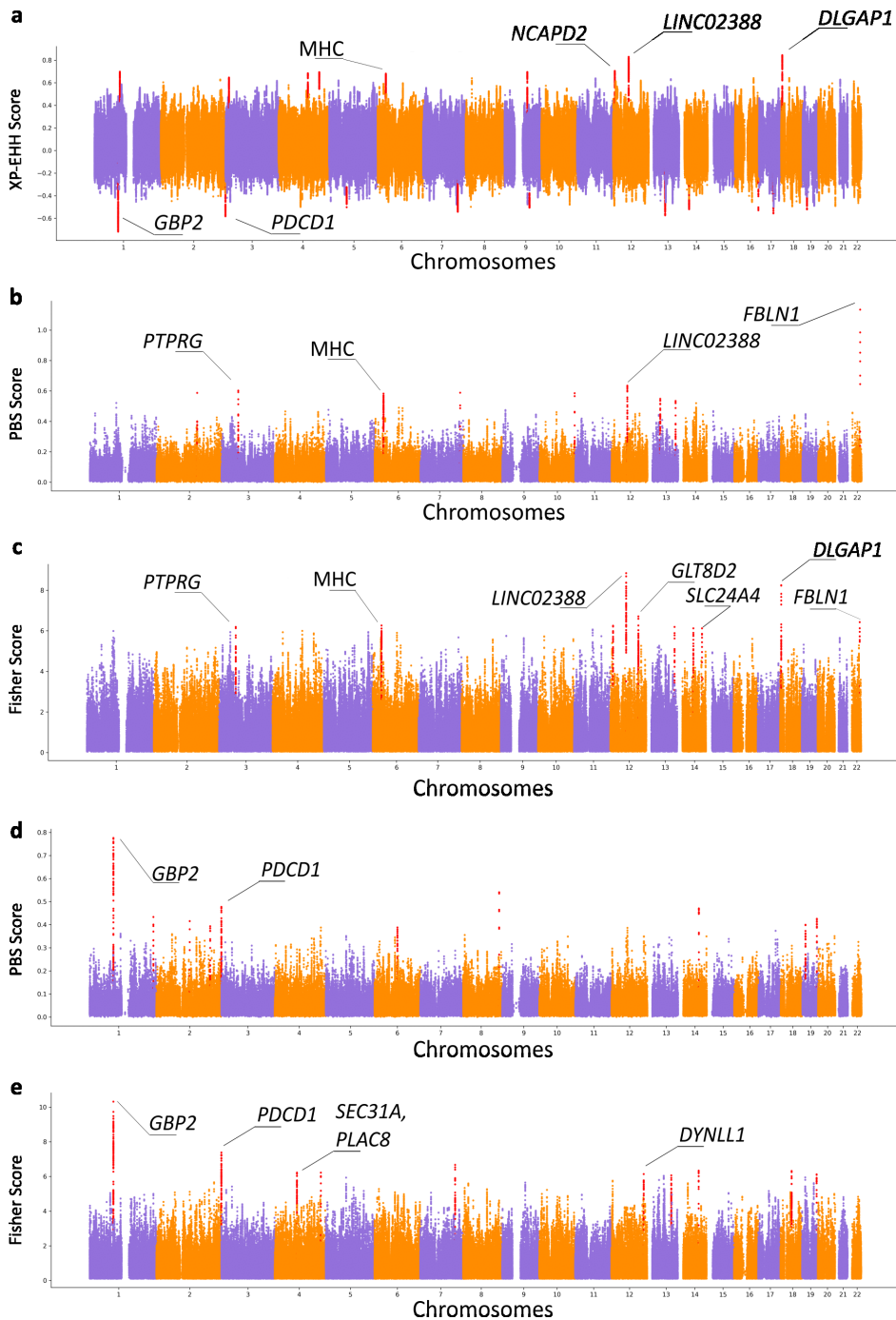
## 5.4. Differential positive selection in PNG highlanders and lowlanders (Ref IV)

### 5.4.1. Genomic regions under selection in PNG highlanders

PNG highlanders face several challenges related to hypoxia, including increased blood viscosity, a high risk of stroke, and neurodevelopmental disorders. To identify genomic regions with signatures of selection in PNG highlanders, we used PBS, XP-EHH and a Fisher Score, which combines the two first scores, with PNG highlanders as the target population, PNG lowlanders as the reference population and Yorubans from the 1000 Genomes as the outgroup population in PBS. We explored further the top ten regions in each of these scores. When merging the overlapping regions in these 30 top regions, we ended up with 23 regions under selection in PNG highlanders that overlap with genes that regulate platelet adhesion, HIF pathway, or neurodevelopment (Figure 16a,b,c). For example, the *FBLN1* gene regulates platelet adhesion (Godyna, Diaz-Ricart, & Argraves, 1996) and thrombosis (Tran et al., 1995). Selection for *FBLN1* in PNG highlanders might counteract the higher risk of thrombosis with increasing altitude because of increased blood viscosity (Ortiz-Prado & Dunn, 2011). The region with the highest Fisher Score overlaps the *LINC02388* intergenic RNA that may play a role in the formation of new blood vessels and increase available oxygen to tissues through regulation of the HIF-pathway (Gudjonsson et al., 2022; Peng et al., 2021; H. Zhou et al., 2021). Several genes regulating brain activity and causing various neurodevelopmental disorders when malfunctioning, such as *DLGAPI* (A. H. Rasmussen, Rasmussen, & Silaharoglu, 2017), are also under positive selection in PNG highlanders. These genes might protect the brain from hypoxic damage. Additionally, we identified candidate genes under selection related to the immune system, including the Major Histocompatibility Complex (MHC) genomic region that contains HLA genes, which might play a central role in the adaptation of PNG highlanders' immune system.

### 5.4.2. Genomic regions under selection in PNG lowlanders

We explored 21 regions with signature of positive selection in PNG lowlanders. These regions were in the top 10 regions with the highest PBS, XP-EHH or Fisher Score, which combines the two first scores, when using PNG lowlanders as the target population, PNG highlanders as the reference population and Yorubans from the 1000 Genomes as the outgroup population in PBS. PNG lowlanders show strong selection in immunity-related regions, specifically in the guanine-binding protein (GBP) family locus. The genomic region overlapping these genes shows the highest XP-EHH, PBS and Fisher Score in PNG lowlanders (Figure 16a,d,e).



**Figure 16: Manhattan plots for the three selection scans among PNG highlanders and lowlanders.** Candidate genes discussed in the paper are shown. (a) XP-EHH scores using PNG highlanders as the target population and PNG lowlanders as the reference population. Genomic regions with the highest score indicate selection in PNG highlanders. Genomic regions with the lowest score indicate selection in PNG lowlanders.

(b) PBS scores using PNG highlanders as the target population, PNG lowlanders as the reference population, and Yorubas from 1000G as the outgroup. (c) Fisher Scores combining the PBS and XP-EHH scores of PNG highlanders. (d) PBS scores using PNG lowlanders as the target population, PNG highlanders as the reference population, and Yorubas from 1000G as the outgroup. (e) Fisher Scores combining the PBS and XP-EHH scores of PNG lowlanders. Reprinted from Figure 1 (André et al., 2022), licensed under Creative Commons CC BY license. Copyright © 2022, André et. al.

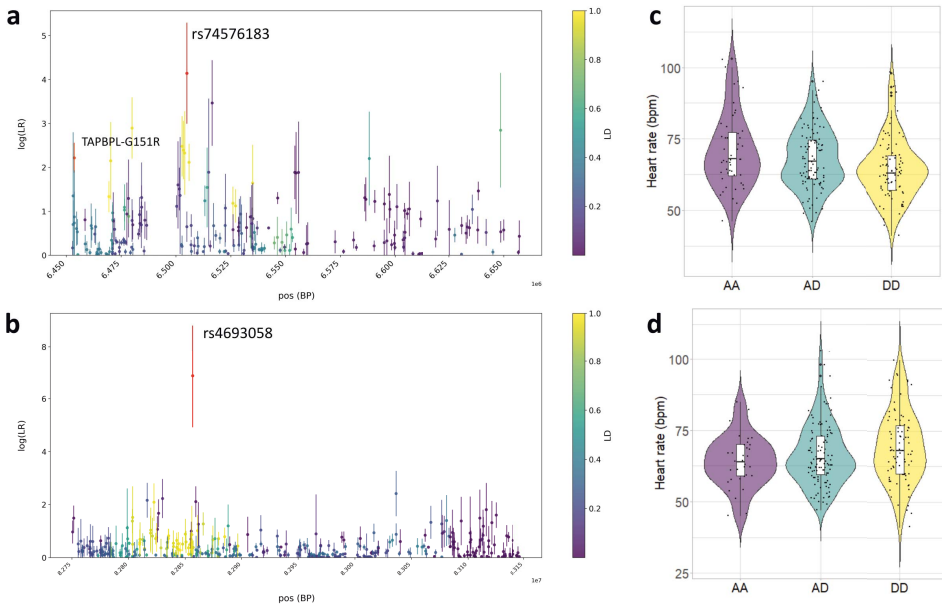
These genes protect against diverse pathogens (Tretina, Park, Maminska, & MacMicking, 2019). For example, one of the *GBP7* variants is associated with the strength of malaria symptoms in Cameroon (Apinjoh et al., 2014). *NRF1*, a regulator of intracellular redox homeostasis (Hu et al., 2022; Y. Zhang & Xiang, 2016), may also have been selected in PNG lowlanders due to pathogenic pressure. Increased oxidative stress during the intraerythrocytic life cycle of the malaria parasite protects the host against severe malaria (Rahbari et al., 2017). We hypothesize these genes might have helped PNG lowlanders to face malaria infection.

#### 5.4.3. Associations of the candidate SNPs with phenotypes in UK biobank and PNG populations

Seven candidate SNPs of PNG highlanders exhibit significant associations with at least one phenotype in the UK Biobank. Among these, three SNPs are significantly associated with haematological phenotypes. Similarly, eight candidate SNPs from PNG lowlanders demonstrate significant associations with phenotypes in the UK Biobank, with four associated with haematological phenotypes. Furthermore, we reported two candidate SNPs associated with heart rate measured in the PNG population (Figure 17). Specifically, the derived G-allele of rs74576183, an intronic variant of *NCAPD2* under selection in PNG highlanders, is associated with a slower heart rate ( $p\text{-value}_{\text{adjusted}} = 0.041$ ,  $\beta = -2.981$ ). On the other hand, the derived allele-T of rs4693058, an intronic variant of *SEC31A* under selection in PNG lowlanders, is associated with a faster heart rate ( $p\text{-value}_{\text{adjusted}} = 0.041$ ,  $\beta = 3.137$ ). However, these associations did not survive the correction for the number of phenotypes ( $p\text{-value}_{\text{adjusted}} > 0.01$ ). It's worth noting that both these SNPs showed significant association with different type of haematological phenotype in the UK biobank, pointing out that the suggestive associations with heart rate may reflect an association with other haematological components that were not measured in our PNG samples. We hypothesize that selection acted on various haematological traits to optimize oxygen transport in PNG highlanders and to counteract the detrimental effects of malaria in PNG lowlanders, which also affects the red blood cells.

### 5.4.4. Functional roles of the candidate SNPs

We investigated the putative regulatory role and effects on the protein structure of the candidate SNPs under selection in PNG highlanders and lowlanders. Five candidate SNPs in PNG highlanders and three in PNG lowlanders, including the two candidate SNPs associated with heart rate, show significant eQTLs in various GTEx tissues. In addition, one of the regions under selection in PNG highlanders overlaps with one missense variant with an exceptionally high DAF in PNG highlanders (0.7) but below 0.12 in African, Asian or European populations. Moreover, this missense variant is in high LD ( $R^2=0.952297$ ) with the candidate SNP, rs74576183, associated with a slower heart rate (Figure 15).



**Figure 17: Clues log(LR) and violin plots of the heart rate distribution depending on the genotype of the candidate SNP for the regions chr12:6452552-6662260 and chr4:82750503-83146792. (a, b) log(LR) for SNPs in regions under selection after five runs of CLUES or 50 runs of CLUES for each of the five top SNPs in the candidate region. The candidate SNP driving selection for the region is shown in red. The colour scale indicates linkage disequilibrium with the candidate SNP. (a) Region chr12:6452552-6662260, which is under selection in PNG highlanders. The candidate SNP for the region is rs74576183. The missense variant (rs7295376, TAPBPL-G151R) in high LD with rs74576183 is shown in orange. (b) Region chr4:82750503-83146792, which is under selection in PNG lowlanders. Candidate SNP is rs4693058. (c, d) Violin plots of the heart rate distribution in PNG depending on their genotype for the candidate SNPs (A = ancestral allele, D = derived allele (under selection)) (c) rs74576183, AA=AA, AD=AG, DD=GG and (d) rs4693058, AA=CC, AD=CT, DD=TT. Reprinted from Figure 2 (André et al., 2022), licensed under Creative Commons CC BY license. Copyright © 2022, André et. al.**

The GBP locus under selection in PNG lowlanders also has a missense variant absent in non-PNG populations, with a DAF of 82% in PNG lowlanders. This variant is also in LD ( $R^2=0.57$ ) with the candidate SNP for the region (rs368120563). While we expect CLUES top results to be enriched for the causal SNPs of selection, we suggest that for these two cases where the candidate SNPs are in LD with a missense variant, the linked missense variant modifying protein sequence is likely to be the real target of selection for the genomic region.

#### 5.4.5. Archaic introgression in the genomic regions under selection

Within PNG highlanders, the region with the strongest signature of selection carries an haplotype sharing similarities with the Altai neanderthal. This region overlaps with *LINC02388*, which might regulate the HIF/VEGF pathway and is under selection in PNG highlanders. Rs74576183, the candidate SNP associated with a slower heart rate and under selection in PNG highlanders, is located in a region carrying a Denisovan-like haplotype.

The six regions under selection in PNG lowlanders and with evidence for archaic introgression include the immunity-related GBP locus, which exhibited the highest selection peak in PNG lowlanders and showed haplotypes with similarities to Denisovan and Altai Neanderthal.

Interestingly, two candidate SNPs for each studied PNG population (a total of four SNPs) are found on introgressed haplotypes and absent on non-archaic haplotypes, but they are not found on the archaic genomes. These observations suggest that the selected mutations appeared after the introgression event, and the selection of the mutation led to an increase in the introgressed haplotype in PNG populations. Another hypothesis might be that the archaic hominin might show polymorphism for this site but that the currently available whole genome sequences for Denisova or Neanderthal do not carry this candidate variant and that the PNG population would carry both the segregating haplotypes.

## 6. CONCLUSIONS

The four references in this thesis contributed to a deeper understanding of the evolutionary history and the genetic and phenotypic diversity of New Guinean populations from both a global perspective of the Sahul settlement and a local perspective of the diverse environments within PNG.

**Ref I** – We present the first models of northern Sahul and New Guinea settlement based on whole-genome data. We described two likely models for the migration across Wallacea to northern Sahul. The first model supports a unique migration from Sunda to Sahul by the ancestors of Indigenous Australians and New Guineans. In contrast, the second model suggests an early split of this ancestral group. We explored additional models describing New Guinea's settlement after the AMH's arrival in northern Sahul. The settlement of New Guinea was probably initiated from its southeast region and followed by two migrations into the highlands.

**Ref II** – By combining absolute and relative chronologies, we show that the first settlers of northern Sahul, the ancestors of New Guineans and Bismarck Islanders, overcame a strong selection almost immediately after reaching this new territory. Selection mostly acted on genes related to the immune response (e.g., *ZDHHC20*), the diet (e.g., *KCNMB2*, *POR*) or both (e.g., *RARB*). Indeed, Sahul's first settlers encountered novel pathogenic organisms and faced the challenge of exploiting unfamiliar food resources, potentially requiring adaptations in nutrient absorption.

**Ref III** – PNG highlanders significantly differ from PNG lowlanders for multiple phenotypes. Specifically, we found that PNG highlanders have a shorter standing height, smaller waist, deeper chest, and higher haemoglobin concentration. There might be likely several processes other than natural selection contributing to these differences. However, given that most of these phenotype differences are similar to those found in other high-altitude populations, we suggest they are part of an adaptive strategy of PNG highlanders to cope with the lower oxygen levels in the highlands. Further genomic analysis has been pursued to investigate this hypothesis in Ref III.

**Ref IV** – This study provides evidence of genomic selection signatures specific to PNG highlanders and lowlanders resulting from distinct environmental pressures such as hypoxia and pathogenic pressure. Our findings suggest that natural selection has shaped genes associated with HIFs, brain development, blood composition, and immunity in PNG highlanders and genes related to immunity and blood composition in lowlanders. Furthermore, we identified one SNP driving selection in highlanders that lower the heart rate, whereas one in lowlanders increases the heart rate. Moreover, we observed that both SNPs are associated with blood composition phenotypes in the UK biobank. Our results suggest that selection acted on haematological components in PNG highlanders and lowlanders and that heart rate might be a proxy. Finally, we also highlighted the potential role of archaic introgression in local adaptation in PNG populations.

This study suggests that natural selection in response to hypoxia has resulted in distinct genetic adaptations in PNG highlanders compared to other high-altitude populations. Nevertheless, our findings indicate that, consistent with observations in other high-altitude populations, cardiovascular phenotypes remain a target of selection in PNG highlanders. PNG lowlanders also showed a strong signal of local adaptation to pathogenic pressure. Our study provides important insights into the genetic mechanisms underlying local adaptation in two PNG populations and highlights the complex interplay between genetic and environmental factors shaping human genetic diversity.

The studies presented in this thesis offer a comprehensive perspective on the interplay between the unique diversity in the populations of PNG and its diverse environment. They emphasize the complexity of the settlement within New Guinea. This thesis also highlights the crucial role of natural selection in shaping genetic variation, which has acted on PNG genomes since the early settlement of Sahul. Moreover, it sheds light on the differences in selection signals between PNG populations using newly sequenced whole-genome sequences and phenotypic measurements. These studies stress the complex interplay between genetic variation, environmental pressures, and local adaptation in PNG populations.

## SUMMARY IN ESTONIAN

Anatoomiliselt kaasaegse inimese evolutsioon Uus-Guineas: asustamise, arhailiste inimlastega segunemise ja kohastumuste ajalugu

Uus-Guinea on üks pikimat aega järjest inimese poolt asustatud piirkondadest väljaspool Aafrikat (O'Connell et al., 2018). Esmaasustamise ajal, kui anatoomiliselt kaasaegsed inimesed (AKI) Aafrikast välja rändasid, oli merepind praegusega võrreldes oluliselt madalam, mistõttu mõned alad, mis tänapäeval on merega eraldatud, moodustasid ühtseid maismaamassiive. Vaikse-Ookeani lõunaosas oli kaks peamist kontinenti: Sunda, mis ühendas paljusid tänaseid Kagu-Aasia saari Aasia kontinendiga, ja Sahul, mis ühendas Austraalia, Tasmaania ja Uus-Guinea. Need kaks iidset kontinenti olid üksteisest eraldatud Wallacea üleminekupiirkonnaga, paljude väikeste saartega pikitud sügava merega, mis takistas inimese liikumist Sunda ja Sahuli vahel (Wallace 1869). Vaatamata neile takistustele jõudsid esimesed inimesed Sahuli pinnale 65 kuni 50 tuhat aastat tagasi (Bradshaw et al., 2021; Clarkson et al., 2017; Pedro et al., 2020). Kuidas täpselt esimesed AKId Wallacea veelahtme ületasid ja Sahuli jõudsid, on siiani vaidlusalane uurimisteema. Ilmselt kavandati teekond ette ja seda tehti mitmes jaos. On ka tõenäoline, et erinevad grupid jõudsid Sahuli eri teid pidi (Bird et al., 2019; Bradshaw et al., 2019).

Lisaks varasele asustamisele kaasaegse inimese poolt paistab Uus-Guinea silma ka ainulaadse bioloogilise ja kultuurilise mitmekesisusega. Kuna suurema osa ajast viimase 50 000 aasta jooksul olid uusguinealased eraldatud nii Euroopast kui Aasiast, on seal praeguseks välja kujunenud rikkalik kohalik geneetiline mitmekesisus (Bergström et al., 2020, 2016; Malaspinas et al., 2016). Üksteisest geneetiliselt kõige eristunud inimgrupid Uus-Guineas on mäestike ja tasandike elanikud. Nende eristumine algas holotseeni ajal, mil grupid jäid üksteise suhtes isolatsiooni (Bergström et al., 2017). Selle aja jooksul on nad pidanud kohanema ka väga erinevate keskkonnamõjudega. Uus-Guineas asuvad Okeania kõrgeimad tipud, mis ulatuvad kuni 4900 meetrini merepinnast (Summerhayes et al., 2017).

Uus-Guinea mäestikuelanikud asusid mägedesse elama vähemalt 20 000 aastat tagasi (Haas et al., 2017; Madsen et al., 2017). Vaatamata kõrgusest tekkivale hapnikuvaegusele e hüpoksiaale, mis on väga tugev väliskeskonnast tulenev stressiallikas, on alates holotseenist mäestikualad kõige tihedamini asustatud Uus-Guinea piirkonnad (Brookfield & Allen, 1989; Müller et al., 2003; Trájer et al., 2020). Vaatamata pikale asustamisajale, pole uuritud sealsete rahvaste kohastumust hüpoksiaga. Üks hüpotees Paapua Uus-Guinea rahvastiku väga ebaühtlase asustusjaotusele on väga tugev patogeeni surve tasandikel (Riley, 1983; Trájer, 2022; Trájer et al., 2020). Malaaria võib olla üks peamisi selle surve põhjustajaid.

Veel üheks uusguinealaste eripäraks on nende genoomides arvukalt säilinud jäljed hiljutisest segunemisest tänaseks väljasurnud hominiini – denisi inimesega (Larena et al., 2021; Reich et al., 2010). Arhailiste inimestega segunemise tulemusel saadud geneetiline materjal võis kiirendada Uus-Guineasse jõudnud AKIde

kohastumist võrreldes *de novo tekkinud* mutatsioonidele mõjuvate valikuprotsessidega (Racimo et al., 2015). Näiteks on tiibetlastel seostatud denisi inimeselt saadud *EPASI* geeni varianti madalama hemoglobiini kontsentratsiooniga veres, mis võib olla kohastumus hüpoksia kahjustava mõju tasakaalustamiseks (Huerta-Sánchez et al., 2014).

Kui AKId liikusid Sundast Sahuli ning jõudsid ka praeguse Uus-Guinea aladele, seisid nad silmitsi paljude uute keskkonnatingimuste ning maastikega. See töö analüüsib Paapua Uus-Guinea populatsioonide geneetilist ja fenotüüpilist mitmekesisust, kombineerides täisgenoomide sekveneerimist ja fenotüüpide mõõtmist. Esimene uurimus kirjeldab genoomikaandmetel põhinevat esmast Põhja-Sahuli asustamise mudelit. Teise uurimuse keskmes on loodusliku valiku kronoloogia AKI rändel läbi Wallacea ja Põhja-Sahuli asustamisel. Kolmas uurimus võrdleb fenotüüpilisi erinevusi Paapua Uus-Guinea mäestiku- ja tasandikuelanike vahel. Neljandas töös uuritakse loodusliku valiku mõju Paapua Uus-Guinea mäestiku- ja tasandikuelanikele. Käesoleva dissertatsiooni uuringud avavad Paapua Uus-Guinea elanike evolutsioonilise ajaloo uued tahud nii laiemalt Sahuli asustamise seisukohast kui ka kitsamalt, keskendudes Paapua Uus-Guinea inimeste kohastumistele sealses mitmekesisuses keskkonnas. Teadustöö tulemused toovad tähelepanu keskmesse keeruka vahekorra geneetilise varieeruvuse, keskkonnamõjude ja kohastumuslike vastuste vahel, mis kõik mõjutavad inimese evolutsiooni.

Esimeses uuringus (Ref I) tuvastasime genoomiandmete põhjal koostatud mudelite abil Uus-Guinea asustamise ajal toimunud peamised geneetilised sündmused. Kirjeldasime meie andmestikuga sobivat kahte Sahuli asustamise mudelit. Esimene mudel toetab Austraalia ja Uus-Guinea põlisasukate eellaste rännet Sundast Sahuli. Teine mudel näitab Austraalia ja Uus-Guinea põlisasukate eellasgruppide varast lahknemist, võimalik, et Wallaceas viibimise ajal. See mudel toetab ka põhjateed kui ühte võimalikku inimeste rändeteed Sahuli. Uurisime ka lisamudeleid, mis kirjeldavad Uus-Guinea asustamise kulgu peale esimeste AKIde saabumist Põhja-Sahuli. Uus-Guinea asustamine sai alguse ilmselt saare loodeosast, levis sealt rännetega lõunas ja põhjas asuvatele madalamatele aladele jõudes lõpuks lääne ja ida mäestikupiirkondadesse.

Teises uuringus (Ref II) seadsime eesmärgiks tuvastada loodusliku valiku ülegenoomseid mustreid. Taastasime valikusündmuste kronoloogia AKI rändel läbi Wallacea ja Põhja-Sahuli asustamise. Tuvastasime loodusliku valiku geneetilisi mustreid Wallaceast, Uus-Guineast ja Bismarcki saarestikust pärit inimeste varem avaldatud täisgenoomidest. Valisime need populatsioonid omavahelise suhtelise eraldatuse tõttu, mis võimaldab tuvastada erinevatele rände- ja asustamisetappidele iseloomulikke valikusignaale. Igale populatsioonile unikaalsete valikusignaale tuvastamiseks kasutasime populatsioonidevahelisi valikuteste: PBS, XP-EHH ja XP-nSL (Sabeti, 2006; Szpiech & Hernandez, 2014; Yi et al., 2010). Tuvastades valikusignaale, mis olid omased vaid ühele populatsioonile kolmest, saime reastada suhtelise valikusündmuste kronoloogia. Valikus olevate geneetiliste variantide koalestsentsiaegade arvutamine GEVA meetodiga (Albers & McVean, 2020) võimaldas paika panna valikusündmuste absoluutse ajaskaala. Kombineerides suhtelist ja absoluutset ajaskaalat, näitasime, et Sahuli esmaasukad – uusguinea-laste ja bismarklaste esivanemad, jäid väga tugeva loodusliku valiku surve alla

peaaegu kohe peale uuele territooriumile jõudmist. Valik mõjus peamiselt geenidele, mis on seotud immuunvastusega (*ZDHHC20*), toitumisega (*KCNMB2*, *POR*) või mõlemaga (*RARB*). Sahuli esmaasukad olid silmitsi uute patogeenidega ning pidid kasutusele võtma ka uusi ja tundmatuid toiduallikaid, mis võisid vajada kohastumist toitainete imendumisel.

Kolmandas uuringus (Ref III) püstitasime hüpoteesi, et Paapua Uus-Guinea mäestikuelanikud erinevad sealsetest tasandikuelanikest teatud fenotüüpide poolest, mis võimaldavad neil hakkama saada mäestiku madalama hapnikusisaldusega. Hüpoteesi testimiseks analüüsisime, kuidas 13 selle uuringu jaoks mõõdetud fenotüüpi on seotud südameveresoonekonna tervise, keha proportsioonide ja kopsu mahuga, ning mil moel erinevad need Paapua Uus-Guinea mäestiku- ja tasandikuelanike vahel. Võrdlesime oma leide ka varem avaldatud vaatlustulemustega Andide, Etioopia ja Tiibeti mäestikuelanikelt. Leidsime, et Paapua Uus-Guinea mäestikuelanikud on lühemad, väiksema vööümbermõdduga, sügavama rinnakorviga ning madalama hemoglobiinitasemega veres võrreldes tasandikuelanikega. Neid erinevusi võivad põhjustada ka mitmed muud protsessid lisaks looduslikule valikule. Samas, enamik neid fenotüübierinevusi on sarnased teistele maailma mäestikupiirkondade elanikele, seega on tõenäoline, et need omadused on osa Paapua Uus-Guinea mäestikuelanike kohastumisest madala hapnikutasemega.

Neljanda uuringus (Ref IV) püstitasime hüpoteesi, et Paapua Uus-Guinea mäestiku- ja tasandikuelanike genoomides võib leida erinevaid loodusliku valiku signaale vastavalt nende keskkonnanišile. Lisahüpotees oli, et need signaalid on seotud varem näidatud fenotüüpide erinevustega (Ref III, André et al., 2021). Nende hüpoteeside uurimiseks sekveneerisime üle 200 Paapua Uus-Guinea mäestiku- ja tasandikuelaniku genoomiproovi, millest tuvastasime populatsioonidele iseloomulikud loodusliku valiku signaalid PBS ja XP-EHH valikuskooridega (Sabeti, 2006; Yi et al., 2010). Tuvastamiseks SNPsid, mis neis populatsioonides on kõige tõenäolisemalt valiku all, kasutasime CLUES meetodit (Stern et al., 2019). Lõpuks seostasime need SNPd konkreetsete fenotüüpidega UK Biopanga andmebaasist (Pan-UKB team, 2020) ja paapua-uusguinealastel mõõdetutega (Ref III, André et al., 2021). Uuring tuvastas vastavalt Paapua Uus-Guinea mäestiku- ja tasandikuelanike genoomidest neile iseloomulikud positiivse loodusliku valiku signaalid, mis on põhjustatud erinevatest keskkonnatingimustest nagu näiteks hüpoksia mäestikus ja patogeenide surve tasandikul. Meie tulemused näitavad, et Paapua Uus-Guinea mäestikuelanikel on looduslik valik mõjutanud genee, mis on seotud HIF1 (hüpoksiast indutseeritud faktor), aju arengu, vere koostise ja immuunsusega, samas kui tasandikuelanikel vaid immuunsuse ja vere koostisega. Lisaks tuvastasime ühe SNP, mis põhjustab mäestikuelanikel madalamat südame löögisagedust ja teise SNP, mis alandab tasandikuelanikel südame löögisagedust. Mõlemad SNPd on UK Biopangas seostatud verekoostise fenotüüpidega. Meie tulemused viitavad, et meie poolt mõõdetud südame löögisagedus on asendustunnus, ja tegelikult mõjutas valik hematoloogilisi komponente ka Paapua Uus-Guinea mäestiku- ja tasandikuelanikel. Samuti hindasime uuringus arhailistelt inimlastelt pärit geneetilise materjali potentsiaalset mõju Paapua Uus-Guinea populatsioonide kohastumisel keskkonnatingimustega.

## REFERENCES

- Ackerman, H., Usen, S., Jallow, M., Sisay-Joof, F., Pinder, M., & Kwiatkowski, D. P. (2005). A comparison of case-control and family-based association methods: The example of sickle-cell and malaria. *Annals of Human Genetics*, *69*(Pt 5), 559–565. <https://doi.org/10.1111/j.1529-8817.2005.00180.x>
- Akhwale, W. S., Lum, J. K., Kaneko, A., Eto, H., Obonyo, C., Björkman, A., & Kobayakawa, T. (2004). Anemia and malaria at different altitudes in the western highlands of Kenya. *Acta Tropica*, *91*(2), 167–175. <https://doi.org/10.1016/j.actatropica.2004.02.010>
- Albers, P. K., & McVean, G. (2020). Dating genomic variants and shared ancestry in population-scale sequencing data. *PLOS Biology*, *18*(1), e3000586. <https://doi.org/10.1371/journal.pbio.3000586>
- Alexander, D. H., Novembre, J., & Lange, K. (2009). Fast model-based estimation of ancestry in unrelated individuals. *Genome Research*, *19*(9), 1655–1664. <https://doi.org/10.1101/gr.094052.109>
- Alkorta-Aranburu, G., Beall, C. M., Witonsky, D. B., Gebremedhin, A., Pritchard, J. K., & Rienzo, A. D. (2012). The Genetic Architecture of Adaptations to High Altitude in Ethiopia. *PLOS Genetics*, *8*(12), e1003110. <https://doi.org/10.1371/journal.pgen.1003110>
- Allen, J., & O'Connell, J. F. (2020). A different paradigm for the initial colonisation of Sahul. *Archaeology in Oceania*, *55*(1), 1–14. <https://doi.org/10.1002/arco.5207>
- Altshuler, D., Donnelly, P., & The International HapMap Consortium. (2005). A haplotype map of the human genome. *Nature*, *437*(7063), 1299–1320. <https://doi.org/10.1038/nature04226>
- André, M., Brucato, N., Hudjasov, G., Pankratov, V., Yermakovich, D., Kreevan, R., ... Ricaut, F.-X. (2022, décembre 15). *Positive selection in the genomes of two Papua New Guinean populations at distinct altitude levels* (p. 2022.12.15.520226). p. 2022.12.15.520226. bioRxiv. <https://doi.org/10.1101/2022.12.15.520226>
- André, M., Brucato, N., Plutniak, S., Kariwiga, J., Muke, J., Morez, A., ... Ricaut, F.-X. (2021). Phenotypic differences between highlanders and lowlanders in Papua New Guinea. *PLOS ONE*, *16*(7), e0253921. <https://doi.org/10.1371/journal.pone.0253921>
- Apinjoh, T. O., Anchang-Kimbi, J. K., Njua-Yafi, C., Ngwai, A. N., Mugri, R. N., Clark, T. G., ... in collaboration with The MalariaGEN Consortium. (2014). Association of candidate gene polymorphisms and TGF-beta/IL-10 levels with malaria in three regions of Cameroon: A case-control study. *Malaria Journal*, *13*(1), 236. <https://doi.org/10.1186/1475-2875-13-236>
- Arenas, M. (2013). The importance and application of the ancestral recombination graph. *Frontiers in Genetics*, *4*. Accessed at <https://www.frontiersin.org/articles/10.3389/fgene.2013.00206>
- Bafaro, E., Liu, Y., Xu, Y., & Dempski, R. E. (2017). The emerging role of zinc transporters in cellular homeostasis and cancer. *Signal Transduction and Targeted Therapy*, *2*, 17029. <https://doi.org/10.1038/sigtrans.2017.29>
- Bammigatti, C., Shetty, S., Shetty, S., & Kumar, A. (2011). Benign tertian malaria – a misnomer? *Tropical Doctor*, *41*(3), 168–169. <https://doi.org/10.1258/td.2011.110025>
- Barry, P. W., & Pollard, A. J. (2003). Altitude illness. *BMJ: British Medical Journal*, *326*(7395), 915–919.
- Beall, C. M., Brittenham, G. M., Strohl, K. P., Blangero, J., Williams-Blangero, S., Goldstein, M. C., ... Gonzales, C. (1998). Hemoglobin concentration of high-altitude Tibetans and Bolivian Aymara. *American Journal of Physical Anthropology*, *106*(3), 385–400. [https://doi.org/10.1002/\(SICI\)1096-8644\(199807\)106:3<385::AID-AJPA10>3.0.CO;2-X](https://doi.org/10.1002/(SICI)1096-8644(199807)106:3<385::AID-AJPA10>3.0.CO;2-X)

- Beall, Cynthia M. (2013). Human adaptability studies at high altitude : Research designs and major concepts during fifty years of discovery. *American Journal of Human Biology*, 25(2), 141–147. <https://doi.org/10.1002/ajhb.22355>
- Beall, Cynthia M. (2014). Adaptation to High Altitude : Phenotypes and Genotypes. *Annual Review of Anthropology*, 43(1), 251–272. <https://doi.org/10.1146/annurev-anthro-102313-030000>
- Beall, Cynthia M., Decker, M. J., Brittenham, G. M., Kushner, I., Gebremedhin, A., & Strohl, K. P. (2002). An Ethiopian pattern of human adaptation to high-altitude hypoxia. *Proceedings of the National Academy of Sciences*, 99(26), 17215–17218. <https://doi.org/10.1073/pnas.252649199>
- Beaumont, M., & Wang, J. (2019). Conservation Genetics. In *Handbook of Statistical Genomics* (p. 457–40). John Wiley & Sons, Ltd. <https://doi.org/10.1002/9781119487845.ch16>
- Bellwood, P., Gamble, C., Blanc, S. A. L., Pluciennik, M., Richards, M., & Terrell, J. E. (2007). *First Farmers : The Origins of Agricultural Societies*, by Peter Bellwood. Malden (MA): Blackwell, 2005; ISBN 0-631-20565-9 hardback £60; ISBN 0-631-20566-7 paperback £17.99, xix+360 pp., 59 figs., 3 tables. *Cambridge Archaeological Journal*, 17(1), 87–109. <https://doi.org/10.1017/S0959774307000078>
- Benjamini, Y., & Hochberg, Y. (1995). Controlling the False Discovery Rate : A Practical and Powerful Approach to Multiple Testing. *Journal of the Royal Statistical Society. Series B (Methodological)*, 57(1), 289–300.
- Bergström, A., McCarthy, S. A., Hui, R., Almarri, M. A., Ayub, Q., Danecek, P., ... Tyler-Smith, C. (2020). Insights into human genetic variation and population history from 929 diverse genomes. *Science*, 367(6484). <https://doi.org/10.1126/science.aay5012>
- Bergström, A., Nagle, N., Chen, Y., McCarthy, S., Pollard, M. O., Ayub, Q., ... Tyler-Smith, C. (2016). Deep Roots for Aboriginal Australian Y Chromosomes. *Current Biology: CB*, 26(6), 809–813. <https://doi.org/10.1016/j.cub.2016.01.028>
- Bergström, A., Oppenheimer, S. J., Mentzer, A. J., Auckland, K., Robson, K., Attenborough, R., ... Tyler-Smith, C. (2017). A Neolithic expansion, but strong genetic structure, in the independent history of New Guinea. *Science*, 357(6356), 1160–1163. <https://doi.org/10.1126/science.aan3842>
- Bersaglieri, T., Sabeti, P. C., Patterson, N., Vanderploeg, T., Schaffner, S. F., Drake, J. A., ... Hirschhorn, J. N. (2004). Genetic signatures of strong recent positive selection at the lactase gene. *American Journal of Human Genetics*, 74(6), 1111–1120. <https://doi.org/10.1086/421051>
- Bhat, S., Kumar, M., & Alva, J. (2013). Malaria and the conducting system of the heart. *BMJ Case Reports*, 2013, bcr2012007462. <https://doi.org/10.1136/bcr-2012-007462>
- Bigham, A. W., & Lee, F. S. (2014). Human high-altitude adaptation : Forward genetics meets the HIF pathway. *Genes & Development*, 28(20), 2189–2204. <https://doi.org/10.1101/gad.250167.114>
- Bird, M. I., Condie, S. A., O'Connor, S., O'Grady, D., Reepmeyer, C., Ulm, S., ... Bradshaw, C. J. A. (2019). Early human settlement of Sahul was not an accident. *Scientific Reports*, 9, 8220. <https://doi.org/10.1038/s41598-019-42946-9>
- Bourke, M. (Mike), & Allen, B. (2021). *Estimating the population of Papua New Guinea in 2020* (SSRN Scholarly Paper N° ID 3770356). Rochester, NY: Social Science Research Network. Accessed at Social Science Research Network website: <https://papers.ssrn.com/abstract=3770356>

- Boyle, E. A., Li, Y. I., & Pritchard, J. K. (2017). An expanded view of complex traits : From polygenic to omnigenic. *Cell*, *169*(7), 1177–1186. <https://doi.org/10.1016/j.cell.2017.05.038>
- Bradshaw, C. J. A., Norman, K., Ulm, S., Williams, A. N., Clarkson, C., Chadœuf, J., ... Saltré, F. (2021). Stochastic models support rapid peopling of Late Pleistocene Sahul. *Nature Communications*, *12*(1), 2440. <https://doi.org/10.1038/s41467-021-21551-3>
- Bradshaw, C. J. A., Ulm, S., Williams, A. N., Bird, M. I., Roberts, R. G., Jacobs, Z., ... Saltré, F. (2019). Minimum founding populations for the first peopling of Sahul. *Nature Ecology & Evolution*, *3*(7), 1057–1063. <https://doi.org/10.1038/s41559-019-0902-6>
- Brisbin, A., Bryc, K., Byrnes, J., Zakharia, F., Omberg, L., Degenhardt, J., ... Bustamante, C. D. (2012). PCAdmix : Principal Components-Based Assignment of Ancestry Along Each Chromosome in Individuals with Admixed Ancestry from Two or More Populations. *Human Biology*, *84*(4), 343–364. <https://doi.org/10.3378/027.084.0401>
- Brito, J., Siqués, P., León-Velarde, F., De La Cruz, J. J., López, V., & Herruzo, R. (2007). Chronic Intermittent Hypoxia at High Altitude Exposure for over 12 Years : Assessment of Hematological, Cardiovascular, and Renal Effects. *High Altitude Medicine & Biology*, *8*(3), 236–244. <https://doi.org/10.1089/ham.2007.8310>
- Brookfield, H., & Allen, B. (1989). High-Altitude Occupation and Environment. *Mountain Research and Development*, *9*(3), 201–209. JSTOR. <https://doi.org/10.2307/3673510>
- Browning, S. R., Browning, B. L., Zhou, Y., Tucci, S., & Akey, J. M. (2018). Analysis of Human Sequence Data Reveals Two Pulses of Archaic Denisovan Admixture. *Cell*, *173*(1), 53–61.e9. <https://doi.org/10.1016/j.cell.2018.02.031>
- Brucato, N., André, M., Hudjashov, G., Mondal, M., Cox, M. P., Leavesley, M., & Ricaut, F.-X. (2022). Chronology of natural selection in Oceanian genomes. *iScience*, 104583. <https://doi.org/10.1016/j.isci.2022.104583>
- Brucato, N., André, M., Tsang, R., Saag, L., Kariwiga, J., Sesuki, K., ... Ricaut, F.-X. (2021). Papua New Guinean Genomes Reveal the Complex Settlement of North Sahul. *Molecular Biology and Evolution*, (msab238). <https://doi.org/10.1093/molbev/msab238>
- Brutsaert, T. D., Soria, R., Caceres, E., Spielvogel, H., & Haas, J. D. (1999). Effect of developmental and ancestral high altitude exposure on chest morphology and pulmonary function in Andean and European/North American natives. *American Journal of Human Biology*, *11*(3), 383–395. [https://doi.org/10.1002/\(SICI\)1520-6300\(1999\)11:3<383::AID-AJHB9>3.0.CO;2-X](https://doi.org/10.1002/(SICI)1520-6300(1999)11:3<383::AID-AJHB9>3.0.CO;2-X)
- Cámara-Leret, R., Frodin, D. G., Adema, F., Anderson, C., Appelhans, M. S., Argent, G., ... van Welzen, P. C. (2020). New Guinea has the world's richest island flora. *Nature*, *584*(7822), 579–583. <https://doi.org/10.1038/s41586-020-2549-5>
- Cavalli-Sforza, L. L., Piazza, A., Menozzi, P., & Mountain, J. (1988). Reconstruction of human evolution: Bringing together genetic, archaeological, and linguistic data. *Proceedings of the National Academy of Sciences*, *85*(16), 6002–6006. <https://doi.org/10.1073/pnas.85.16.6002>
- Charlesworth, D., Charlesworth, B., & Morgan, M. T. (1995). The pattern of neutral molecular variation under the background selection model. *Genetics*, *141*(4), 1619–1632. <https://doi.org/10.1093/genetics/141.4.1619>

- Charlesworth, Deborah. (2006). Balancing Selection and Its Effects on Sequences in Nearby Genome Regions. *PLOS Genetics*, 2(4), e64. <https://doi.org/10.1371/journal.pgen.0020064>
- Chen, L., Wolf, A. B., Fu, W., Li, L., & Akey, J. M. (2020). Identifying and Interpreting Apparent Neanderthal Ancestry in African Individuals. *Cell*. <https://doi.org/10.1016/j.cell.2020.01.012>
- Cheong, H. I., Janocha, A. J., Monocello, L. T., Garchar, A. C., Gebremedhin, A., Erzurum, S. C., & Beall, C. M. (2016). Alternative hematological and vascular adaptive responses to high-altitude hypoxia in East African highlanders. *American Journal of Physiology-Lung Cellular and Molecular Physiology*, 312(2), L172–L177. <https://doi.org/10.1152/ajplung.00451.2016>
- Choin, J., Mendoza-Revilla, J., Arauna, L. R., Cuadros-Espinoza, S., Cassar, O., Larena, M., ... Quintana-Murci, L. (2021). Genomic insights into population history and biological adaptation in Oceania. *Nature*, 1–7. <https://doi.org/10.1038/s41586-021-03236-5>
- Clarkson, C., Jacobs, Z., Marwick, B., Fullagar, R., Wallis, L., Smith, M., ... Pardoe, C. (2017). Human occupation of northern Australia by 65,000 years ago. *Nature*, 547(7663), 306–310. <https://doi.org/10.1038/nature22968>
- Cleary, E., Hetzel, M. W., & Clements, A. C. A. (2022). A review of malaria epidemiology and control in Papua New Guinea 1900 to 2021 : Progress made and future directions. *Frontiers in Epidemiology*, 2. Accessed at <https://www.frontiersin.org/articles/10.3389/fepid.2022.980795>
- Coller, M. (2009). SahulTime : Rethinking Archaeological Representation in the Digital Age. *Archaeologies*, 5(1), 110–123. <https://doi.org/10.1007/s11759-009-9096-x>
- Cotes, J. E., Anderson, H. R., & Patrick, J. M. (1974). Lung Function and the Response to Exercise in New Guineans : Role of Genetic and Environmental Factors. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 268(893), 349–361. <https://doi.org/10.1098/rstb.1974.0034>
- Crawford, D. C., Bhangale, T., Li, N., Hellenthal, G., Rieder, M. J., Nickerson, D. A., & Stephens, M. (2004). Evidence for substantial fine-scale variation in recombination rates across the human genome. *Nature Genetics*, 36(7), 700–706. <https://doi.org/10.1038/ng1376>
- Danecek, P., Auton, A., Abecasis, G., Albers, C. A., Banks, E., DePristo, M. A., ... 1000 Genomes Project Analysis Group. (2011). The variant call format and VCFtools. *Bioinformatics*, 27(15), 2156–2158. <https://doi.org/10.1093/bioinformatics/btr330>
- Dannemann, M., Andrés, A. M., & Kelso, J. (2016). Introgression of Neandertal- and Denisovan-like Haplotypes Contributes to Adaptive Variation in Human Toll-like Receptors. *The American Journal of Human Genetics*, 98(1), 22–33. <https://doi.org/10.1016/j.ajhg.2015.11.015>
- Dannemann, M., & Kelso, J. (2017). The Contribution of Neanderthals to Phenotypic Variation in Modern Humans. *American Journal of Human Genetics*, 101(4), 578–589. <https://doi.org/10.1016/j.ajhg.2017.09.010>
- Denham, T. P. (2003). Origins of Agriculture at Kuk Swamp in the Highlands of New Guinea. *Science*, 301(5630), 189–193. <https://doi.org/10.1126/science.1085255>
- Department of Nutrition for Health and Development (NHD). (2011). *Haemoglobin concentrations for the diagnosis of anaemia and assessment of severity*. World Health Organization. Accessed at <http://www.who.int/vmnis/indicators/haemoglobin.pdf>

- Douka, K., Slon, V., Jacobs, Z., Ramsey, C. B., Shunkov, M. V., Derevianko, A. P., ... Higham, T. (2019). Age estimates for hominin fossils and the onset of the Upper Palaeolithic at Denisova Cave. *Nature*, *565*(7741), 640–644. <https://doi.org/10.1038/s41586-018-0870-z>
- Drakeley, C. J., Carneiro, I., Reyburn, H., Malima, R., Lusingu, J. P. A., Cox, J., ... Riley, E. M. (2005). Altitude-dependent and -independent variations in *Plasmodium falciparum* prevalence in northeastern Tanzania. *The Journal of Infectious Diseases*, *191*(10), 1589–1598. <https://doi.org/10.1086/429669>
- Droma, T., McCullough, R. G., McCullough, R. E., Zhuang, J., Cymerman, A., Sun, S., ... Moore, L. G. (1991). Increased vital and total lung capacities in Tibetan compared to Han residents of Lhasa (3,658 m). *American Journal of Physical Anthropology*, *86*(3), 341–351. <https://doi.org/10.1002/ajpa.1330860303>
- Dudbridge, F., & Gusnanto, A. (2008). Estimation of significance thresholds for genome-wide association scans. *Genetic Epidemiology*, *32*(3), 227–234. <https://doi.org/10.1002/gepi.20297>
- Eichstaedt, C. A., Antão, T., Cardona, A., Pagani, L., Kivisild, T., & Mormina, M. (2015). Genetic and phenotypic differentiation of an Andean intermediate altitude population. *Physiological Reports*, *3*(5), e12376. <https://doi.org/10.14814/phy2.12376>
- Enard, D., & Petrov, D. A. (2018). Evidence that RNA Viruses Drove Adaptive Introgression between Neanderthals and Modern Humans. *Cell*, *175*(2), 360-371.e13. Scopus. <https://doi.org/10.1016/j.cell.2018.08.034>
- Fan, S., Hansen, M. E. B., Lo, Y., & Tishkoff, S. A. (2016). Going global by adapting local : A review of recent human adaptation. *Science*, *354*(6308), 54–59. <https://doi.org/10.1126/science.aaf5098>
- Ferrer-Admetlla, A., Liang, M., Korneliussen, T., & Nielsen, R. (2014). On Detecting Incomplete Soft or Hard Selective Sweeps Using Haplotype Structure. *Molecular Biology and Evolution*, *31*(5), 1275–1291. <https://doi.org/10.1093/molbev/msu077>
- Fijarczyk, A., & Babik, W. (2015). Detecting balancing selection in genomes : Limits and prospects. *Molecular Ecology*, *24*(14), 3529–3545. <https://doi.org/10.1111/mec.13226>
- Flint, J., Harding, R. M., Boyce, A. J., & Clegg, J. B. (1998). 1 The population genetics of the haemoglobinopathies. *Baillière's Clinical Haematology*, *11*(1), 1–51. [https://doi.org/10.1016/S0950-3536\(98\)80069-3](https://doi.org/10.1016/S0950-3536(98)80069-3)
- Frisancho, A. R. (2013). Developmental Functional Adaptation to High Altitude : Review. *American Journal of Human Biology*, *25*(2), 151–168. <https://doi.org/10.1002/ajhb.22367>
- Fumagalli, M., Sironi, M., Pozzoli, U., Ferrer-Admetlla, A., Pattini, L., & Nielsen, R. (2011). Signatures of Environmental Genetic Adaptation Pinpoint Pathogens as the Main Selective Pressure through Human Evolution. *PLOS Genetics*, *7*(11), e1002355. <https://doi.org/10.1371/journal.pgen.1002355>
- Gaffney, D., Ford, A., & Summerhayes, G. (2015). Crossing the Pleistocene–Holocene transition in the New Guinea Highlands : Evidence from the lithic assemblage of Kiowa rockshelter. *Journal of Anthropological Archaeology*, *39*, 223–246. <https://doi.org/10.1016/j.jaa.2015.04.006>
- GBD 2013 Mortality and Causes of Death Collaborators. (2015). Global, regional, and national age–sex specific all-cause and cause-specific mortality for 240 causes of death, 1990–2013 : A systematic analysis for the Global Burden of Disease Study 2013. *The Lancet*, *385*(9963), 117–171. [https://doi.org/10.1016/S0140-6736\(14\)61682-2](https://doi.org/10.1016/S0140-6736(14)61682-2)

- Gething, P. W., Elyazar, I. R. F., Moyes, C. L., Smith, D. L., Battle, K. E., Guerra, C. A., ... Hay, S. I. (2012). A Long Neglected World Malaria Map : Plasmodium vivax Endemicity in 2010. *PLoS Neglected Tropical Diseases*, 6(9), e1814. <https://doi.org/10.1371/journal.pntd.0001814>
- Godyna, S., Diaz-Ricart, M., & Argraves, W. (1996). Fibulin-1 mediates platelet adhesion via a bridge of fibrinogen. *Blood*, 88(7), 2569–2577. <https://doi.org/10.1182/blood.V88.7.2569.bloodjournal8872569>
- Golson, J. (Éd.). (2017). *Ten thousand years of cultivation at Kuk Swamp in the highlands of Papua New Guinea*. Acton, ACT, Australia: Australian National University Press.
- Green, R. E., Krause, J., Briggs, A. W., Maricic, T., Stenzel, U., Kircher, M., ... Pääbo, S. (2010). A Draft Sequence of the Neandertal Genome. *Science*, 328(5979), 710–722. <https://doi.org/10.1126/science.1188021>
- Greksa, L. P. (2006). Growth and Development of Andean High Altitude Residents. *High Altitude Medicine & Biology*, 7(2), 116–124. <https://doi.org/10.1089/ham.2006.7.116>
- Griffiths, R. C., & Marjoram, P. (1997). An Ancestral Recombination Graph. In P. Donnelly & S. Tavaré (Éds.), *Progress in Population Genetics and Human Evolution* (p. 257–270). New York, NY: Springer New York. [https://doi.org/10.1007/978-1-4757-2609-1\\_16](https://doi.org/10.1007/978-1-4757-2609-1_16)
- Grove, M., Lamb, H., Roberts, H., Davies, S., Marshall, M., Bates, R., & Huws, D. (2015). Climatic variability, plasticity, and dispersal : A case study from Lake Tana, Ethiopia. *Journal of Human Evolution*, 87, 32–47. <https://doi.org/10.1016/j.jhevol.2015.07.007>
- Gudjonsson, A., Gudmundsdottir, V., Axelsson, G. T., Gudmundsson, E. F., Jonsson, B. G., Launer, L. J., ... Gudnason, V. (2022). A genome-wide association study of serum proteins reveals shared loci with common diseases. *Nature Communications*, 13(1), 480. <https://doi.org/10.1038/s41467-021-27850-z>
- Gupta, S., Gazendam, N., Farina, J. M., Saldarriaga, C., Mendoza, I., López-Santi, R., ... Baranchuk, A. (2021). Malaria and the Heart : JACC State-of-the-Art Review. *Journal of the American College of Cardiology*, 77(8), 1110–1121. <https://doi.org/10.1016/j.jacc.2020.12.042>
- Haas, R., Stefanescu, I. C., Garcia-Putnam, A., Aldenderfer, M. S., Clementz, M. T., Murphy, M. S., ... Watson, J. T. (2017). Humans permanently occupied the Andean highlands by at least 7 ka. *Royal Society Open Science*, 4(6), 170331. <https://doi.org/10.1098/rsos.170331>
- Haldane, J. B. S. (1919). The combination of linkage values, and the calculation of distances between the loci of linked factors. *Journal of Genetics*, 8(4), 299–309.
- Haldar, K., & Mohandas, N. (2009). Malaria, erythrocytic infection, and anemia. *Hematology. American Society of Hematology. Education Program*, 87–93. <https://doi.org/10.1182/asheducation-2009.1.87>
- Hall, J. E., & Guyton, A. C. (2011). *Guyton and Hall textbook of medical physiology*. Philadelphia, PA: Saunders Elsevier. Accessed at <http://www.clinicalkey.com/dura/browse/bookChapter/3-s2.0-C20090602506>
- Hammer, M. F., Woerner, A. E., Mendez, F. L., Watkins, J. C., & Wall, J. D. (2011). Genetic evidence for archaic admixture in Africa. *Proceedings of the National Academy of Sciences*, 108(37), 15123–15128. <https://doi.org/10.1073/pnas.1109300108>
- Harris, K., & Nielsen, R. (2016). The Genetic Cost of Neanderthal Introgression. *Genetics*, 203(2), 881–891. <https://doi.org/10.1534/genetics.116.186890>
- Harrison, G. A., Küchemann, C. F., Moore, M. a. S., Boyce, A. J., Baju, T., Mourant, A. E., ... Clegg, E. J. (1969). The effects of altitudinal variation in Ethiopian populations. *Philosophical Transactions of the Royal Society of London. B, Biological Sciences*, 256(805), 147–182. <https://doi.org/10.1098/rstb.1969.0040>

- Harrison, T., Krigbaum, J., & Manser, J. (2006). Primate Biogeography and Ecology on the Sunda Shelf Islands : A Paleontological and Zooarchaeological Perspective. In S. M. Lehman & J. G. Fleagle (Éds.), *Primate Biogeography : Progress and Prospects* (p. 331–372). Boston, MA: Springer US. [https://doi.org/10.1007/0-387-31710-4\\_12](https://doi.org/10.1007/0-387-31710-4_12)
- Hawkins, S., O'Connor, S., Maloney, T. R., Litster, M., Kealy, S., Fenner, J. N., ... Louys, J. (2017). Oldest human occupation of Wallacea at Laili Cave, Timor-Leste, shows broad-spectrum foraging responses to late Pleistocene environments. *Quaternary Science Reviews*, *171*, 58–72. <https://doi.org/10.1016/j.quascirev.2017.07.008>
- Hein, J., Schierup, M. H., & Wiuf, C. (2005). *Gene genealogies, variation and evolution : A primer in coalescent theory*. Oxford ; New York: Oxford University Press.
- Higham, T., Douka, K., Wood, R., Ramsey, C. B., Brock, F., Basell, L., ... Jacobi, R. (2014). The timing and spatiotemporal patterning of Neanderthal disappearance. *Nature*, *512*(7514), 306–309. <https://doi.org/10.1038/nature13621>
- Hoit, B. D., Dalton, N. D., Gebremedhin, A., Janocha, A., Zimmerman, P. A., Zimmerman, A. M., ... Beall, C. M. (2011). Elevated pulmonary artery pressure among Amhara highlanders in Ethiopia. *American journal of human biology : the official journal of the Human Biology Council*, *23*(2). <https://doi.org/10.1002/ajhb.21130>
- Holm, A., Gomes, L. C., Biering-Soerensen, T., Silvestre, O., & Brainin, P. (2020). Malaria and cardiovascular disease : A systematic review. *European Heart Journal*, *41*(Supplement 2), ehaa946.2817. <https://doi.org/10.1093/ehjci/ehaa946.2817>
- Hu, S., Feng, J., Wang, M., Wufuer, R., Liu, K., Zhang, Z., & Zhang, Y. (2022, juin 14). *Nrf1 is an indispensable redox-determining factor for mitochondrial homeostasis by integrating multi-hierarchical regulatory networks* (p. 2022.05.04.490622). p. 2022.05.04.490622. bioRxiv. <https://doi.org/10.1101/2022.05.04.490622>
- Hubisz, M., & Siepel, A. (2020). Inference of Ancestral Recombination Graphs Using ARGweaver. In J. Y. Duthiel (Éd.), *Statistical Population Genomics* (p. 231–266). New York, NY: Springer US. [https://doi.org/10.1007/978-1-0716-0199-0\\_10](https://doi.org/10.1007/978-1-0716-0199-0_10)
- Hublin, J. J. (2009). The origin of Neandertals. *Proceedings of the National Academy of Sciences*, *106*(38), 16022–16027. <https://doi.org/10.1073/pnas.0904119106>
- Hublin, J.-J., Ben-Ncer, A., Bailey, S. E., Freidline, S. E., Neubauer, S., Skinner, M. M., ... Gunz, P. (2017). New fossils from Jebel Irhoud, Morocco and the pan-African origin of Homo sapiens. *Nature*, *546*(7657), 289–292. <https://doi.org/10.1038/nature22336>
- Hudson, R. R. (1983). Properties of a neutral allele model with intragenic recombination. *Theoretical Population Biology*, *23*(2), 183–201. [https://doi.org/10.1016/0040-5809\(83\)90013-8](https://doi.org/10.1016/0040-5809(83)90013-8)
- Huerta-Sánchez, E., DeGiorgio, M., Pagani, L., Tarekegn, A., Ekong, R., Antao, T., ... Nielsen, R. (2013). Genetic Signatures Reveal High-Altitude Adaptation in a Set of Ethiopian Populations. *Molecular Biology and Evolution*, *30*(8), 1877–1888. <https://doi.org/10.1093/molbev/mst089>
- Huerta-Sánchez, E., Jin, X., Asan, Bianba, Z., Peter, B. M., Vinckenbosch, N., ... Nielsen, R. (2014). Altitude adaptation in Tibetans caused by introgression of Denisovan-like DNA. *Nature*, *512*(7513), 194–197. <https://doi.org/10.1038/nature13408>
- Hung, H., & Carson, M. T. (2014). Foragers, fishers and farmers : Origins of the Taiwanese Neolithic. *Antiquity*, *88*(342), 1115–1131. <https://doi.org/10.1017/S0003598X00115352>
- Itan, Y., Powell, A., Beaumont, M. A., Burger, J., & Thomas, M. G. (2009). The Origins of Lactase Persistence in Europe. *PLoS Computational Biology*, *5*(8), e1000491. <https://doi.org/10.1371/journal.pcbi.1000491>

- Jacobs, G. S., Hudjashov, G., Saag, L., Kusuma, P., Darusallam, C. C., Lawson, D. J., ... Cox, M. P. (2019). Multiple Deeply Divergent Denisovan Ancestries in Papuans. *Cell*, 177(4), 1010-1021.e32. <https://doi.org/10.1016/j.cell.2019.02.035>
- Jobling, M., Hollox, E., Hurler, M., Kivisild, T., & Tyler-Smith, C. (2013). *Human Evolutionary Genetics* (2<sup>e</sup> éd.). Garland Science. <https://doi.org/10.1201/9781317952268>
- Juric, I., Aeschbacher, S., & Coop, G. (2016). The Strength of Selection against Neanderthal Introgression. *PLoS Genetics*, 12(11), e1006340. <https://doi.org/10.1371/journal.pgen.1006340>
- Kaplan, N. L., Hudson, R. R., & Langley, C. H. (1989). The «hitchhiking effect» revisited. *Genetics*, 123(4), 887–899. <https://doi.org/10.1093/genetics/123.4.887>
- Karmin, M., Flores, R., Saag, L., Hudjashov, G., Brucato, N., Crenna-Darusallam, C., ... Cox, M. P. (2022). Episodes of Diversification and Isolation in Island Southeast Asian and Near Oceanian Male Lineages. *Molecular Biology and Evolution*, 39(3), msac045. <https://doi.org/10.1093/molbev/msac045>
- Kayser, M. (2010). The human genetic history of Oceania : Near and remote views of dispersal. *Current Biology: CB*, 20(4), R194–201. <https://doi.org/10.1016/j.cub.2009.12.004>
- Kealy, S., Louys, J., & O'Connor, S. (2018). Least-cost pathway models indicate northern human dispersal from Sunda to Sahul. *Journal of Human Evolution*, 125, 59–70. <https://doi.org/10.1016/j.jhevol.2018.10.003>
- Key, F. M., Fu, Q., Romagne, F., Lachmann, M., & Andres, A. M. (2016). Human adaptation and population differentiation in the light of ancient genomes. *Nature Communications*, 7. ScopuS. <https://doi.org/10.1038/ncomms10775>
- Kho, S., Barber, B. E., Johar, E., Andries, B., Poespoprodjo, J. R., Kenangalem, E., ... McMorran, B. J. (2018). Platelets kill circulating parasites of all major Plasmodium species in human malaria. *Blood*, 132(12), 1332–1344. <https://doi.org/10.1182/blood-2018-05-849307>
- Kingman, J. F. C. (1982). The coalescent. *Stochastic Processes and Their Applications*, 13(3), 235–248. [https://doi.org/10.1016/0304-4149\(82\)90011-4](https://doi.org/10.1016/0304-4149(82)90011-4)
- Kirch, P. V., & Hunt, T. L. (1988). Radiocarbon Dates from the Mussau Islands and the Lapita Colonization of the Southwestern Pacific. *Radiocarbon*, 30(2), 161–169. <https://doi.org/10.1017/S0033822200044106>
- Kirch, Patrick Vinton. (2002). *On the road of the winds : An archaeological history of the Pacific islands before European contact* (1. paperback print). Berkeley, Calif.: Univ. of California Press.
- Kitur, U., Adair, T., Riley, I., & Lopez, A. D. (2019). Estimating the pattern of causes of death in Papua New Guinea. *BMC Public Health*, 19(1), 1322. <https://doi.org/10.1186/s12889-019-7620-5>
- Kong, A., Thorleifsson, G., Gudbjartsson, D. F., Masson, G., Sigurdsson, A., Jonasdottir, A., ... Stefansson, K. (2010). Fine-scale recombination rate differences between sexes, populations and individuals. *Nature*, 467(7319), 1099–1103. <https://doi.org/10.1038/nature09525>
- Krause, J., Fu, Q., Good, J. M., Viola, B., Shunkov, M. V., Derevianko, A. P., & Pääbo, S. (2010). The complete mitochondrial DNA genome of an unknown hominin from southern Siberia. *Nature*, 464(7290), 894–897. <https://doi.org/10.1038/nature08976>
- Kuhlwilm, M., Gronau, I., Hubisz, M. J., de Filippo, C., Prado-Martinez, J., Kircher, M., ... Castellano, S. (2016). Ancient gene flow from early modern humans into Eastern Neanderthals. *Nature*, 530(7591), 429–433. <https://doi.org/10.1038/nature16544>

- Kusuma, P., Brucato, N., Cox, M. P., Letellier, T., Manan, A., Nuraini, C., ... Ricaut, F.-X. (2017). The last sea nomads of the Indonesian archipelago : Genomic origins and dispersal. *European Journal of Human Genetics*, 25(8), 1004–1010. <https://doi.org/10.1038/ejhg.2017.88>
- Kwiatkowski, D. P. (2005). How Malaria Has Affected the Human Genome and What Human Genetics Can Teach Us about Malaria. *The American Journal of Human Genetics*, 77(2), 171–192. <https://doi.org/10.1086/432519>
- Lampah, D. A., Yeo, T. W., Malloy, M., Kenangalem, E., Douglas, N. M., Ronaldo, D., ... Price, R. N. (2015). Severe Malarial Thrombocytopenia : A Risk Factor for Mortality in Papua, Indonesia. *The Journal of Infectious Diseases*, 211(4), 623–634. <https://doi.org/10.1093/infdis/jiu487>
- Larena, M., McKenna, J., Sanchez-Quinto, F., Bernhardsson, C., Ebeo, C., Reyes, R., ... Jakobsson, M. (2021). Philippine Ayta possess the highest level of Denisovan ancestry in the world. *Current Biology: CB*, S0960-9822(21)00977-5. <https://doi.org/10.1016/j.cub.2021.07.022>
- Latinis, D. K., & Stark, K. (2005). Cave Use Variability in Central Maluku, Eastern Indonesia. *Asian Perspectives*, 44(1), 119–136.
- Lawson, D. J., Hellenthal, G., Myers, S., & Falush, D. (2012). Inference of Population Structure using Dense Haplotype Data. *PLOS Genetics*, 8(1), e1002453. <https://doi.org/10.1371/journal.pgen.1002453>
- Lee, P., Chandel, N. S., & Simon, M. C. (2020). Cellular adaptation to hypoxia through hypoxia inducible factors and beyond. *Nature Reviews Molecular Cell Biology*, 21(5), 268–283. <https://doi.org/10.1038/s41580-020-0227-y>
- Leonard, W. R. (1989). Nutritional determinants of high-altitude growth in Nuñoa, Peru. *American Journal of Physical Anthropology*, 80(3), 341–352. <https://doi.org/10.1002/ajpa.1330800308>
- Lewontin, R. C., & Krakauer, J. (1973). Distribution of gene frequency as a test of the theory of the selective neutrality of polymorphisms. *Genetics*, 74(1), 175–195. <https://doi.org/10.1093/genetics/74.1.175>
- Li, N., & Stephens, M. (2003). Modeling Linkage Disequilibrium and Identifying Recombination Hotspots Using Single-Nucleotide Polymorphism Data. *Genetics*, 165(4), 2213–2233. <https://doi.org/10.1093/genetics/165.4.2213>
- Lipson, M., Skoglund, P., Spriggs, M., Valentin, F., Bedford, S., Shing, R., ... Reich, D. (2018). Population Turnover in Remote Oceania Shortly after Initial Settlement. *Current Biology: CB*, 28(7), 1157–1165.e7. <https://doi.org/10.1016/j.cub.2018.02.051>
- Liu, X., Li, Y. I., & Pritchard, J. K. (2019). Trans Effects on Gene Expression Can Drive Omnigenic Inheritance. *Cell*, 177(4), 1022–1034.e6. <https://doi.org/10.1016/j.cell.2019.04.014>
- Lonsdale, J., Thomas, J., Salvatore, M., Phillips, R., Lo, E., Shad, S., ... Moore, H. F. (2013). The Genotype-Tissue Expression (GTEx) project. *Nature Genetics*, 45(6), 580–585. <https://doi.org/10.1038/ng.2653>
- Loos, R. J. F., & Yeo, G. S. H. (2022). The genetics of obesity : From discovery to biology. *Nature Reviews Genetics*, 23(2), 120–133. <https://doi.org/10.1038/s41576-021-00414-z>
- Lopez, M., Choin, J., Sikora, M., Siddle, K., Harmant, C., Costa, H. A., ... Quintana-Murci, L. (2019). Genomic Evidence for Local Adaptation of Hunter-Gatherers to the African Rainforest. *Current Biology*, 29(17), 2926–2935.e4. <https://doi.org/10.1016/j.cub.2019.07.013>

- Lunter, G. (2019). Haplotype matching in large cohorts using the Li and Stephens model. *Bioinformatics*, 35(5), 798–806. <https://doi.org/10.1093/bioinformatics/bty735>
- MacMicking, J. D. (2012). Interferon-inducible effector mechanisms in cell-autonomous immunity. *Nature Reviews Immunology*, 12(5), 367–382. <https://doi.org/10.1038/nri3210>
- Madsen, D. B., Perreault, C., Rhode, D., Sun, Y., Yi, M., Brunson, K., & Brantingham, P. J. (2017). Early foraging settlement of the Tibetan Plateau highlands. *Archaeological Research in Asia*, 11, 15–26. <https://doi.org/10.1016/j.ara.2017.04.003>
- Malaspinas, A.-S., Westaway, M. C., Muller, C., Sousa, V. C., Lao, O., Alves, I., ... Willerslev, E. (2016). A genomic history of Aboriginal Australia. *Nature*, 538(7624), 207–214. <https://doi.org/10.1038/nature18299>
- Mallick, S., Li, H., Lipson, M., Mathieson, I., Gymrek, M., Racimo, F., ... Reich, D. (2016). The Simons Genome Diversity Project : 300 genomes from 142 diverse populations. *Nature*, 538(7624), 201–206. <https://doi.org/10.1038/nature18964>
- Marnetto, D., & Huerta-Sánchez, E. (2017). Haplostrips : Revealing population structure through haplotype visualization. *Methods in Ecology and Evolution*, 8(10), 1389–1392. <https://doi.org/10.1111/2041-210X.12747>
- Mathieson, I. (2021). The omnigenic model and polygenic prediction of complex traits. *The American Journal of Human Genetics*, 108(9), 1558–1563. <https://doi.org/10.1016/j.ajhg.2021.07.003>
- McLaren, W., Gil, L., Hunt, S. E., Riat, H. S., Ritchie, G. R. S., Thormann, A., ... Cunningham, F. (2016). The Ensembl Variant Effect Predictor. *Genome Biology*, 17(1), 122. <https://doi.org/10.1186/s13059-016-0974-4>
- McManus, K. F., Taravella, A. M., Henn, B. M., Bustamante, C. D., Sikora, M., & Cornejo, O. E. (2017). Population genetic analysis of the DARC locus (Duffy) reveals adaptation from standing variation associated with malaria resistance in humans. *PLoS Genetics*, 13(3), e1006560. <https://doi.org/10.1371/journal.pgen.1006560>
- McMichael, T. M., Zhang, L., Chemudupati, M., Hach, J. C., Kenney, A. D., Hang, H. C., & Yount, J. S. (2017). The palmitoyltransferase ZDHHC20 enhances interferon-induced transmembrane protein 3 (IFITM3) palmitoylation and antiviral activity. *Journal of Biological Chemistry*, 292(52), 21517–21526. <https://doi.org/10.1074/jbc.M117.800482>
- McMorran, B. J. (2019). Immune role of platelets in malaria. *ISBT Science Series*, 14(1), 67–76. <https://doi.org/10.1111/voxs.12451>
- McVean, G. A. T., Myers, S. R., Hunt, S., Deloukas, P., Bentley, D. R., & Donnelly, P. (2004). The Fine-Scale Structure of Recombination Rate Variation in the Human Genome. *Science*, 304(5670), 581–584. <https://doi.org/10.1126/science.1092500>
- Mendez, F. L., Watkins, J. C., & Hammer, M. F. (2012). A Haplotype at STAT2 Introgressed from Neanderthals and Serves as a Candidate of Positive Selection in Papua New Guinea. *The American Journal of Human Genetics*, 91(2), 265–274. <https://doi.org/10.1016/j.ajhg.2012.06.015>
- Meyer, M., Arsuaga, J.-L., de Filippo, C., Nagel, S., Aximu-Petri, A., Nickel, B., ... Pääbo, S. (2016). Nuclear DNA sequences from the Middle Pleistocene Sima de los Huesos hominins. *Nature*, 531(7595), 504–507. <https://doi.org/10.1038/nature17405>
- Mondal, M., Bertranpetit, J., & Lao, O. (2019). Approximate Bayesian computation with deep learning supports a third archaic introgression in Asia and Oceania. *Nature Communications*, 10(1), 246. <https://doi.org/10.1038/s41467-018-08089-7>
- Mondal, M., Casals, F., Xu, T., Dall’Olio, G. M., Pybus, M., Netea, M. G., ... Bertranpetit, J. (2016). Genomic analysis of Andamanese provides insights into ancient human migration into Asia and adaptation. *Nature Genetics*, 48(9), 1066–1070. <https://doi.org/10.1038/ng.3621>

- Moore, L. G. (2017). Measuring high-altitude adaptation. *Journal of Applied Physiology*, 123(5), 1371–1385. <https://doi.org/10.1152/jappphysiol.00321.2017>
- Mountain, M.-J. (1991). Bulmer phase I: environmental change and human activity through the late Pleistocene to the Holocene in the highlands of New Guinea: A scenario. In R. Bulmer & A. Pawley (Éds.), *Man and a half: Essays in Pacific anthropology and ethnobiology in honour of Ralph Bulmer*. Auckland: Polynesian Society.
- Mueller, W. H., Schull, V. N., Schull, W. J., Soto, P., & Rothhammer, F. (1978). A multinational Andean Genetic and Health Program: Growth and development in an hypoxic environment. *Annals of Human Biology*, 5(4), 329–352. <https://doi.org/10.1080/03014467800002981>
- Müller, I., Bockarie, M., Alpers, M., & Smith, T. (2003). The epidemiology of malaria in Papua New Guinea. *Trends in Parasitology*, 19(6), 253–259. [https://doi.org/10.1016/S1471-4922\(03\)00091-6](https://doi.org/10.1016/S1471-4922(03)00091-6)
- Myers, S., Bottolo, L., Freeman, C., McVean, G., & Donnelly, P. (2005). A Fine-Scale Map of Recombination Rates and Hotspots Across the Human Genome. *Science*, 310(5746), 321–324. <https://doi.org/10.1126/science.1117196>
- Naraqi, S., Feling, B., & Leeder, S. R. (2003). Disease and death in Papua New Guinea. *Medical Journal of Australia*, 178(1), 7–8. <https://doi.org/10.5694/j.1326-5377.2003.tb05030.x>
- Nordborg, M. (2019). Coalescent Theory. In D. Balding, I. Moltke, & J. Marioni (Éds.), *Handbook of Statistical Genomics* (1<sup>re</sup> éd., p. 145–30). Wiley. <https://doi.org/10.1002/9781119487845.ch5>
- Norman, K., Inglis, J., Clarkson, C., Faith, J. T., Shulmeister, J., & Harris, D. (2018). An early colonisation pathway into northwest Australia 70–60,000 years ago. *Quaternary Science Reviews*, 180, 229–239. <https://doi.org/10.1016/j.quascirev.2017.11.023>
- O’Connell, J. F., & Allen, J. (2015). The process, biotic impact, and global implications of the human colonization of Sahul about 47,000 years ago. *Journal of Archaeological Science*, 56, 73–84. <https://doi.org/10.1016/j.jas.2015.02.020>
- O’Connell, J. F., Allen, J., Williams, M. A. J., Williams, A. N., Turney, C. S. M., Spooner, N. A., ... Cooper, A. (2018). When did Homo sapiens first reach Southeast Asia and Sahul? *Proceedings of the National Academy of Sciences*, 115(34), 8482–8490. <https://doi.org/10.1073/pnas.1808385115>
- O’Connor, S. (2007). New evidence from East Timor contributes to our understanding of earliest modern human colonisation east of the Sunda Shelf. *Antiquity*, 81(313), 523–535. <https://doi.org/10.1017/S0003598X00095569>
- O’Connor, S., Louys, J., Kealy, S., & Samper Carro, S. C. (2017). Hominin Dispersal and Settlement East of Huxley’s Line: The Role of Sea Level Changes, Island Size, and Subsistence Behavior. *Current Anthropology*, 58(S17), S567–S582. <https://doi.org/10.1086/694252>
- Ohta, T. (2017). Nearly Neutral Theory☆. In *Reference Module in Life Sciences*. Elsevier. <https://doi.org/10.1016/B978-0-12-809633-8.06779-0>
- Orlando, L., Allaby, R., Skoglund, P., Der Sarkissian, C., Stockhammer, P. W., Ávila-Arcos, M. C., ... Warinner, C. (2021). Ancient DNA analysis. *Nature Reviews Methods Primers*, 1(1), 1–26. <https://doi.org/10.1038/s43586-020-00011-0>
- Ortiz-Prado, E., & Dunn, J. F. (2011). *High altitude exposure and ischemic stroke: A literature review*. <https://doi.org/10.11575/PRISM/10691>
- O’Sullivan, J. M., Preston, R. J. S., O’Regan, N., & O’Donnell, J. S. (2016). Emerging roles for hemostatic dysfunction in malaria pathogenesis. *Blood*, 127(19), 2281–2288. <https://doi.org/10.1182/blood-2015-11-636464>

- Pagani, L., Lawson, D. J., Jagoda, E., Mörseburg, A., Eriksson, A., Mitt, M., ... Metspalu, M. (2016). Genomic analyses inform on migration events during the peopling of Eurasia. *Nature*, 538(7624), 238–242. <https://doi.org/10.1038/nature19792>
- Pain, A., Ferguson, D. J. P., Kai, O., Urban, B. C., Lowe, B., Marsh, K., & Roberts, D. J. (2001). Platelet-mediated clumping of Plasmodium falciparum-infected erythrocytes is a common adhesive phenotype and is associated with severe malaria. *Proceedings of the National Academy of Sciences*, 98(4), 1805–1810. <https://doi.org/10.1073/pnas.98.4.1805>
- Palmer, B. (2017). *The Languages and Linguistics of the New Guinea Area : A Comprehensive Guide*. Walter de Gruyter GmbH & Co KG.
- Pan-UKB team. (2020). Accessed at <https://pan.ukbb.broadinstitute.org>
- Pareek, C. S., Smoczynski, R., & Tretyn, A. (2011). Sequencing technologies and genome sequencing. *Journal of Applied Genetics*, 52(4), 413–435. <https://doi.org/10.1007/s13353-011-0057-x>
- Patterson, N., Moorjani, P., Luo, Y., Mallick, S., Rohland, N., Zhan, Y., ... Reich, D. (2012). Ancient Admixture in Human History. *Genetics*, 192(3), 1065–1093. <https://doi.org/10.1534/genetics.112.145037>
- Pawley, A., & Australian National University (Éds.). (2005). *Papuan pasts : Cultural, linguistic and biological histories of Papuan-speaking peoples*. Canberra: Pacific Linguistics, Research School of Pacific and Asian Studies, Australian National University.
- Payne, S., Kumar Bc, R., Pomeroy, E., Macintosh, A., & Stock, J. (2018). Thrifty phenotype versus cold adaptation : Trade-offs in upper limb proportions of Himalayan populations of Nepal. *Royal Society Open Science*, 5(6), 172174. <https://doi.org/10.1098/rsos.172174>
- Pedro, N., Brucato, N., Fernandes, V., André, M., Saag, L., Pomat, W., ... Ricaut, F.-X. (2020). Papuan mitochondrial genomes and the settlement of Sahul. *Journal of Human Genetics*, 1–13. <https://doi.org/10.1038/s10038-020-0781-3>
- Peng, C., Chen, H., Li, Y., Yang, H., Qin, P., Ma, B., ... Guo, D. (2021). LRIG3 Suppresses Angiogenesis by Regulating the PI3K/AKT/VEGFA Signaling Pathway in Glioma. *Frontiers in Oncology*, 11. Accessed at <https://www.frontiersin.org/article/10.3389/fonc.2021.621154>
- Peter Bellwood & Eusebio Dizon. (2005). The Batanes Archaeological Project and the Out of Taiwan Hypothesis for Austronesian Dispersal. *南島研究學報*, 1(1). <https://doi.org/10.29884/JAS.200506.0001>
- Petr, M., Pääbo, S., Kelso, J., & Vernet, B. (2019). Limits of long-term selection against Neandertal introgression. *Proceedings of the National Academy of Sciences*, 116(5), 1639–1644. <https://doi.org/10.1073/pnas.1814338116>
- Pickrell, J. K., & Pritchard, J. K. (2012). Inference of population splits and mixtures from genome-wide allele frequency data. *PLoS Genetics*, 8(11), e1002967. <https://doi.org/10.1371/journal.pgen.1002967>
- Piešová, M., & Mach, M. (2020). Impact of Perinatal Hypoxia on the Developing Brain. *Physiological Research*, 69(2), 199–213. <https://doi.org/10.33549/physiolres.934198>
- Prüfer, K., de Filippo, C., Grote, S., Mafessoni, F., Korlević, P., Hajdinjak, M., ... Pääbo, S. (2017). A high-coverage Neandertal genome from Vindija Cave in Croatia. *Science (New York, N.Y.)*, 358(6363), 655–658. <https://doi.org/10.1126/science.aao1887>
- Prüfer, K., Racimo, F., Patterson, N., Jay, F., Sankararaman, S., Sawyer, S., ... Pääbo, S. (2014). The complete genome sequence of a Neandertal from the Altai Mountains. *Nature*, 505(7481), 43–49. <https://doi.org/10.1038/nature12886>

- Prugnolle, F., Manica, A., Charpentier, M., Guégan, J. F., Guernier, V., & Balloux, F. (2005). Pathogen-Driven Selection and Worldwide HLA Class I Diversity. *Current Biology*, *15*(11), 1022–1027. <https://doi.org/10.1016/j.cub.2005.04.050>
- Quach, H., Rotival, M., Pothlichet, J., Loh, Y.-H. E., Dannemann, M., Zidane, N., ... Quintana-Murci, L. (2016). Genetic Adaptation and Neandertal Admixture Shaped the Immune System of Human Populations. *Cell*, *167*(3), 643–656.e17. <https://doi.org/10.1016/j.cell.2016.09.024>
- Racimo, F., Sankararaman, S., Nielsen, R., & Huerta-Sánchez, E. (2015). Evidence for archaic adaptive introgression in humans. *Nature Reviews Genetics*, *16*(6), 359–371. <https://doi.org/10.1038/nrg3936>
- Rahman, A., Tabassum, T., Araf, Y., Al Nahid, A., Ullah, M. A., & Hosen, M. J. (2021). Silent hypoxia in COVID-19 : Pathomechanism and possible management strategy. *Molecular Biology Reports*, *48*(4), 3863–3869. <https://doi.org/10.1007/s11033-021-06358-1>
- Rasmussen, A. H., Rasmussen, H. B., & Silaharoglu, A. (2017). The DLGAP family : Neuronal expression, function and role in brain disorders. *Molecular Brain*, *10*(1), 43. <https://doi.org/10.1186/s13041-017-0324-9>
- Rasmussen, M. D., Hubisz, M. J., Gronau, I., & Siepel, A. (2014). Genome-Wide Inference of Ancestral Recombination Graphs. *PLOS Genetics*, *10*(5), e1004342. <https://doi.org/10.1371/journal.pgen.1004342>
- Rees, J. S., Castellano, S., & Andrés, A. M. (2020). The Genomics of Human Local Adaptation. *Trends in Genetics*, *36*(6), 415–428. <https://doi.org/10.1016/j.tig.2020.03.006>
- Reich, D., Green, R. E., Kircher, M., Krause, J., Patterson, N., Durand, E. Y., ... Pääbo, S. (2010). Genetic history of an archaic hominin group from Denisova Cave in Siberia. *Nature*, *468*(7327), 1053–1060. <https://doi.org/10.1038/nature09710>
- Reilly, P. F., Tjahjadi, A., Miller, S. L., Akey, J. M., & Tucci, S. (2022). The contribution of Neanderthal introgression to modern human traits. *Current Biology*, *32*(18), R970–R983. <https://doi.org/10.1016/j.cub.2022.08.027>
- Riley, I. D. (1983). Population change and distribution in Papua New Guinea : An epidemiological approach. *Journal of Human Evolution*, *12*(1), 125–132. [https://doi.org/10.1016/S0047-2484\(83\)80017-7](https://doi.org/10.1016/S0047-2484(83)80017-7)
- Rupert, J. L., & Hochachka, P. W. (2001). The Evidence for Hereditary Factors Contributing to High Altitude Adaptation in Andean Natives : A Review. *High Altitude Medicine & Biology*, *2*(2), 235–256. <https://doi.org/10.1089/152702901750265332>
- Sabeti, P. C. (2006). Positive Natural Selection in the Human Lineage. *Science*, *312*(5780), 1614–1620. <https://doi.org/10.1126/science.1124309>
- Sabeti, P. C., Varilly, P., Fry, B., Lohmueller, J., Hostetter, E., Cotsapas, C., ... Lander, E. S. (2007). Genome-wide detection and characterization of positive selection in human populations. *Nature*, *449*(7164), 913–918. <https://doi.org/10.1038/nature06250>
- Sams, A. J., Dumaine, A., Nédélec, Y., Yotova, V., Alfieri, C., Tanner, J. E., ... Barreiro, L. B. (2016). Adaptively introgressed Neandertal haplotype at the OAS locus functionally impacts innate immune responses in humans. *Genome Biology*, *17*(1), 246. <https://doi.org/10.1186/s13059-016-1098-6>
- Sankararaman, S., Mallick, S., Dannemann, M., Prüfer, K., Kelso, J., Pääbo, S., ... Reich, D. (2014). The genomic landscape of Neanderthal ancestry in present-day humans. *Nature*, *507*(7492), 354–357. <https://doi.org/10.1038/nature12961>

- Sankararaman, S., Mallick, S., Patterson, N., & Reich, D. (2016). The Combined Landscape of Denisovan and Neanderthal Ancestry in Present-Day Humans. *Current Biology*, 26(9), 1241–1247. <https://doi.org/10.1016/j.cub.2016.03.037>
- Scheinfeldt, L. B., Soi, S., Thompson, S., Ranciaro, A., Woldemeskel, D., Beggs, W., ... Tishkoff, S. A. (2012). Genetic adaptation to high altitude in the Ethiopian highlands. *Genome Biology*, 13(1), R1. <https://doi.org/10.1186/gb-2012-13-1-r1>
- Schiffels, S., & Durbin, R. (2014). Inferring human population size and separation history from multiple genome sequences. *Nature genetics*, 46(8), 919–925. <https://doi.org/10.1038/ng.3015>
- Ségurel, L., & Bon, C. (2017). On the Evolution of Lactase Persistence in Humans. *Annual Review of Genomics and Human Genetics*, 18(1), 297–319. <https://doi.org/10.1146/annurev-genom-091416-035340>
- Sella, G., & Barton, N. H. (2019). Thinking About the Evolution of Complex Traits in the Era of Genome-Wide Association Studies. *Annual Review of Genomics and Human Genetics*, 20(1), 461–493. <https://doi.org/10.1146/annurev-genom-083115-022316>
- Senn, N., Maraga, S., Sie, A., Rogerson, S. J., Reeder, J. C., Siba, P., & Mueller, I. (2010). Population Hemoglobin Mean and Anemia Prevalence in Papua New Guinea : New Metrics for Defining Malaria Endemicity? *PLOS ONE*, 5(2), e9375. <https://doi.org/10.1371/journal.pone.0009375>
- Shaffer, J. P. (1995). Multiple Hypothesis Testing. *Annual Review of Psychology*, 46(1), 561–584. <https://doi.org/10.1146/annurev.ps.46.020195.003021>
- Siddle, K. J., & Quintana-Murci, L. (2014). The Red Queen’s long race: Human adaptation to pathogen pressure. *Current Opinion in Genetics & Development*, 29, 31–38. <https://doi.org/10.1016/j.gde.2014.07.004>
- Silubonde, T. M., Baumgartner, J., Ware, L. J., Malan, L., Smuts, C. M., & Norris, S. (2020). Adjusting Haemoglobin Values for Altitude Maximizes Combined Sensitivity and Specificity to Detect Iron Deficiency among Women of Reproductive Age in Johannesburg, South Africa. *Nutrients*, 12(3), 633. <https://doi.org/10.3390/nu12030633>
- Simonson, T. S., Yang, Y., Huff, C. D., Yun, H., Qin, G., Witherspoon, D. J., ... Ge, R. (2010). Genetic Evidence for High-Altitude Adaptation in Tibet. *Science*, 329(5987), 72–75. <https://doi.org/10.1126/science.1189406>
- Skoglund, P., Posth, C., Sirak, K., Spriggs, M., Valentin, F., Bedford, S., ... Reich, D. (2016). Genomic insights into the peopling of the Southwest Pacific. *Nature*, 538(7626), 510–513. <https://doi.org/10.1038/nature19844>
- Skov, L., Hui, R., Shchur, V., Hobolth, A., Scally, A., Schierup, M. H., & Durbin, R. (2018). Detecting archaic introgression using an unadmixed outgroup. *PLOS Genetics*, 14(9), e1007641. <https://doi.org/10.1371/journal.pgen.1007641>
- Smith, J. M. B. (2001). Did early hominids cross sea gaps on natural rafts. In *Faunal and Floral Migration and Evolution in SE Asia-Australasia* (p. 409–416). CRC Press.
- Souilmi, Y., Lauterbur, M. E., Tobler, R., Huber, C. D., Johar, A. S., Moradi, S. V., ... Enard, D. (2021). An ancient viral epidemic involving host coronavirus interacting genes more than 20,000 years ago in East Asia. *Current Biology*, 31(16), 3504–3514.e9. <https://doi.org/10.1016/j.cub.2021.05.067>
- Speidel, L., Forest, M., Shi, S., & Myers, S. R. (2019). A method for genome-wide genealogy estimation for thousands of samples. *Nature Genetics*, 51(9), 1321–1329. <https://doi.org/10.1038/s41588-019-0484-x>
- Stearns, F. W. (2010). One Hundred Years of Pleiotropy : A Retrospective. *Genetics*, 186(3), 767–773. <https://doi.org/10.1534/genetics.110.122549>

- Stembridge, M., Williams, A. M., Gasho, C., Dawkins, T. G., Drane, A., Villafuerte, F. C., ... Ainslie, P. N. (2019). The overlooked significance of plasma volume for successful adaptation to high altitude in Sherpa and Andean natives. *Proceedings of the National Academy of Sciences of the United States of America*, *116*(33), 16177–16179. <https://doi.org/10.1073/pnas.1909002116>
- Stern, A. J., & Nielsen, R. (2019). Detecting Natural Selection. In *Handbook of Statistical Genomics* (p. 397–40). John Wiley & Sons, Ltd. <https://doi.org/10.1002/9781119487845.ch14>
- Stern, A. J., Wilton, P. R., & Nielsen, R. (2019). An approximate full-likelihood method for inferring selection and allele frequency trajectories from DNA sequence data. *PLOS Genetics*, *15*(9), e1008384. <https://doi.org/10.1371/journal.pgen.1008384>
- Summerhayes, G. R., Field, J. H., Shaw, B., & Gaffney, D. (2017). The archaeology of forest exploitation and change in the tropics during the Pleistocene: The case of Northern Sahul (Pleistocene New Guinea). *Quaternary International*, *448*, 14–30. <https://doi.org/10.1016/j.quaint.2016.04.023>
- Summerhayes, G. R., Leavesley, M., Fairbairn, A., Mandui, H., Field, J., Ford, A., & Fullagar, R. (2010). Human Adaptation and Plant Use in Highland New Guinea 49,000 to 44,000 Years Ago. *Science*, *330*(6000), 78–81. <https://doi.org/10.1126/science.1193130>
- Sutikna, T., Tocheri, M. W., Faith, J. T., Jatmiko, Due Awe, R., Meijer, H. J. M., ... Roberts, R. G. (2018). The spatio-temporal distribution of archaeological and faunal finds at Liang Bua (Flores, Indonesia) in light of the revised chronology for *Homo floresiensis*. *Journal of Human Evolution*, *124*, 52–74. <https://doi.org/10.1016/j.jhevol.2018.07.001>
- Szpiech, Z. A., & Hernandez, R. D. (2014). selscan: An efficient multithreaded program to perform EHH-based scans for positive selection. *Molecular Biology and Evolution*, *31*(10), 2824–2827. <https://doi.org/10.1093/molbev/msu211>
- Szpiech, Z. A., Novak, T. E., Bailey, N. P., & Stevison, L. S. (2021). Application of a novel haplotype-based scan for local adaptation to study high-altitude adaptation in rhesus macaques. *Evolution Letters*, *5*(4), 408–421. <https://doi.org/10.1002/evl3.232>
- Terrell, J. E. (2004). The « sleeping giant » hypothesis and New Guinea's place in the prehistory of Greater Near Oceania. *World Archaeology*, *36*(4), 601–609. <https://doi.org/10.1080/0043824042000303782>
- The 1000 Genomes Project Consortium, Gibbs, R. A., Boerwinkle, E., Doddapaneni, H., Han, Y., Korchina, V., ... Rasheed, A. (2015). A global reference for human genetic variation. *Nature*, *526*(7571), 68–74. <https://doi.org/10.1038/nature15393>
- The Nobel Prize in Physiology or Medicine 2022. (2022). Consulté 8 mars 2023, à l'adresse NobelPrize.org website: <https://www.nobelprize.org/prizes/medicine/2022/summary/>
- Tobler, R., Rohrlach, A., Soubrier, J., Bover, P., Llamas, B., Tuke, J., ... Cooper, A. (2017). Aboriginal mitogenomes reveal 50,000 years of regionalism in Australia. *Nature*, *544*(7649), 180–184. <https://doi.org/10.1038/nature21416>
- Totino, P. R. R., Daniel-Ribeiro, C. T., & Ferreira-da-Cruz, M. de F. (2016). Evidencing the Role of Erythrocytic Apoptosis in Malarial Anemia. *Frontiers in Cellular and Infection Microbiology*, *6*. <https://doi.org/10.3389/fcimb.2016.00176>
- Trájer, A. J. (2022). Late Quaternary changes in malaria-free areas in Papua New Guinea and the future perspectives. *Quaternary International*. <https://doi.org/10.1016/j.quaint.2022.04.003>

- Trájer, A. J., Sebestyén, V., & Domokos, E. (2020). The potential impacts of climate factors and malaria on the Middle Palaeolithic population patterns of ancient humans. *Quaternary International*, 565, 94–108. <https://doi.org/10.1016/j.quaint.2020.10.056>
- Tran, H., Tanaka, A., Litvinovich, S. V., Medved, L. V., Haudenschild, C. C., & Argraves, W. S. (1995). The Interaction of Fibulin-1 with Fibrinogen : A POTENTIAL ROLE IN HEMOSTASIS AND THROMBOSIS (\*). *Journal of Biological Chemistry*, 270(33), 19458–19464. <https://doi.org/10.1074/jbc.270.33.19458>
- Tretina, K., Park, E.-S., Maminska, A., & MacMicking, J. D. (2019). Interferon-induced guanylate-binding proteins : Guardians of host defense in health and disease. *The Journal of Experimental Medicine*, 216(3), 482–500. <https://doi.org/10.1084/jem.20182031>
- van den Bergh, G. D., Li, B., Brumm, A., Grün, R., Yurnaldi, D., Moore, M. W., ... Morwood, M. J. (2016). Earliest hominin occupation of Sulawesi, Indonesia. *Nature*, 529(7585), 208–211. <https://doi.org/10.1038/nature16448>
- Van Valen, L. (1973). A New Evolutionary Law. *Evolutionary Theory*, 1, 1–30.
- Vattathil, S., & Akey, J. M. (2015). Small Amounts of Archaic Admixture Provide Big Insights into Human History. *Cell*, 163(2), 281–284. <https://doi.org/10.1016/j.cell.2015.09.042>
- Vernot, B., & Akey, J. M. (2014). Resurrecting Surviving Neandertal Lineages from Modern Human Genomes. *Science*, 343(6174), 1017–1021. <https://doi.org/10.1126/science.1245938>
- Vernot, B., Tucci, S., Kelso, J., Schraiber, J. G., Wolf, A. B., Gittelman, R. M., ... Akey, J. M. (2016). Excavating Neandertal and Denisovan DNA from the genomes of Melanesian individuals. *Science (New York, N.Y.)*, 352(6282), 235–239. <https://doi.org/10.1126/science.aad9416>
- Vespasiani, D. M., Jacobs, G. S., Cook, L. E., Brucato, N., Leavesley, M., Kinipi, C., ... Romero, I. G. (2022). Denisovan introgression has shaped the immune system of present-day Papuans. *PLOS Genetics*, 18(12), e1010470. <https://doi.org/10.1371/journal.pgen.1010470>
- Villafuerte, F. C., & Corante, N. (2016). Chronic Mountain Sickness : Clinical Aspects, Etiology, Management, and Treatment. *High Altitude Medicine & Biology*, 17(2), 61–69. <https://doi.org/10.1089/ham.2016.0031>
- Voight, B. F., Kudaravalli, S., Wen, X., & Pritchard, J. K. (2006). A Map of Recent Positive Selection in the Human Genome. *PLOS Biology*, 4(3), e72. <https://doi.org/10.1371/journal.pbio.0040072>
- Weitz, C. A., Garruto, R. M., Chin, C. T., & Liu, J. C. (2004). Morphological growth and thorax dimensions among Tibetan compared to Han children, adolescents and young adults born and raised at high altitude. *Annals of Human Biology*, 31(3), 292–310. <https://doi.org/10.1080/0301446042000196316>
- White, N. J. (2018). Anaemia and malaria. *Malaria Journal*, 17(1), 371. <https://doi.org/10.1186/s12936-018-2509-9>
- Woolcock, A. J., Colman, M. H., & Blackburn, C. R. B. (1972). Factors Affecting Normal Values for Ventilatory Lung Function<sup>1,2</sup>. *American Review of Respiratory Disease*. (world). <https://doi.org/10.1164/arrd.1972.106.5.692>
- World Health Organization. (2022a). *World malaria report 2022*. Geneva: World Health Organization. Accessed at <https://www.who.int/teams/global-malaria-programme/reports/world-malaria-report-2022>

- World Health Organization. (2022b, janvier 28). Child mortality (under 5 years). Consulté 27 février 2023, à l'adresse <https://www.who.int/news-room/fact-sheets/detail/levels-and-trends-in-child-under-5-mortality-in-2020>
- Yermakovich, D., Pankratov, V., Võsa, U., Yunusbayev, B., Estonian Biobank Research Team, & Dannemann, M. (2022). Long-range regulatory effects of Neandertal DNA in modern humans. *Genetics*, iyac188. <https://doi.org/10.1093/genetics/iyac188>
- Yi, X., Liang, Y., Huerta-Sanchez, E., Jin, X., Cuo, Z. X. P., Pool, J. E., ... Wang, J. (2010). Sequencing of 50 Human Exomes Reveals Adaptation to High Altitude. *Science*, 329(5987), 75–78. <https://doi.org/10.1126/science.1190371>
- Yuan, K., Ni, X., Liu, C., Pan, Y., Deng, L., Zhang, R., ... Xu, S. (2021). Refining models of archaic admixture in Eurasia with ArchaicSeeker 2.0. *Nature Communications*, 12(1), 6232. <https://doi.org/10.1038/s41467-021-26503-5>
- Zhai, W., Nielsen, R., & Slatkin, M. (2009). An Investigation of the Statistical Power of Neutrality Tests Based on Comparative and Population Genetic Data. *Molecular Biology and Evolution*, 26(2), 273–283. <https://doi.org/10.1093/molbev/msn231>
- Zhang, X., Witt, K. E., Bañuelos, M. M., Ko, A., Yuan, K., Xu, S., ... Huerta-Sanchez, E. (2021). The history and evolution of the Denisovan-EPAS1 haplotype in Tibetans. *Proceedings of the National Academy of Sciences*, 118(22), e2020803118. <https://doi.org/10.1073/pnas.2020803118>
- Zhang, Y., & Xiang, Y. (2016). Molecular and cellular basis for the unique functioning of Nrf1, an indispensable transcription factor for maintaining cell homeostasis and organ integrity. *Biochemical Journal*, 473(8), 961–1000. <https://doi.org/10.1042/BJ20151182>
- Zhou, H., Cao, J., Yang, F., Fan, D., Li, H., Fan, T., & Sun, P. (2021). Member Domain 3 (LRIG3) Activates Hypoxia-Inducible Factor-1  $\alpha$  /Vascular Endothelial Growth Factor (HIF-1  $\alpha$  /VEGF) Pathway to Inhibit the Growth of Bone Marrow Mesenchymal Stem Cells in Glioma. *Journal of Biomaterials and Tissue Engineering*, 11(5), 1022–1027. <https://doi.org/10.1166/jbt.2021.2629>
- Zhou, X., & Stephens, M. (2012). Genome-wide efficient mixed-model analysis for association studies. *Nature Genetics*, 44(7), 821–824. <https://doi.org/10.1038/ng.2310>

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## **PUBLICATIONS**

## CURRICULUM VITAE

**Name:** Mathilde Frédérique E. André  
**Date of birth:** November 9, 1995  
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**Address:** University of Tartu, Institute of Genomics, Riia 23b, 51010  
Tartu, Estonia  
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### Education:

2019–2023 Doctoral studies, University of Tartu, Faculty of Science and Technology, Institute of Genomics, Chair of Evolutionary Biology  
2017–2019 MSc, Evolutionary Anthropology (“Master Humanités Médicales”), University of Aix-Marseille (AMU)  
2013–2017 BSc, Major in Biological Sciences with Minor in History of Art and Archaeology, Catholic University of Louvain (UCLouvain)

### Professional employment:

2021–2023 Junior Research Fellow of Modern Population

### Teaching:

2020–2021 Teaching assistant, University of Tartu, Bachelor in Science and Technology, Chair of Evolutionary Biology, Evolution and the Natural World course

### International Courses and Conferences:

- 2023** Society for Molecular Biology and Evolution meeting (SMBE), Ferrara, Italy  
Poster: Genomic Signatures of Local Adaptation to Extreme Environments in Papua New Guinean Highlanders and Lowlanders
- 2023** The Biology of Genomes, Cold Spring Laboratory (CSHL), USA  
Poster: Positive selection in the genomes of two Papua New Guinean populations at distinct altitude levels
- 2022** Congress of the Indo-Pacific Prehistory Association (IPPA), Chiang Mai, Thailand  
Oral presentation: Phenotype Adaptation and Positive Selection to Altitude in Papua New Guinean Highlanders
- 2022** EMBO | EMBL Symposium: Reconstructing the human past: using ancient and modern genomics, Heidelberg, Germany  
Flask talk and poster: Papuan New Guineans blood composition is affected by positive selection
- 2021** Australian Archeological Association conference, online  
Oral presentation: Papua New Guineans Show Signs of Biological Adaptations to Altitude

- 2021** Wellcome Connecting Science conference– Human Evolution– From Fossils to Ancient and Modern Genomes, online  
Poster: Natural Selection and phenotypic associations in Papuan Highlanders
- 2021** ASHG virtual meeting 2021, online  
Poster: Positive selection to altitude and phenotypic associations in Papua New Guinea highlanders
- 2021** EMBO Course: Population genomics: Background, tools, and programming, online
- 2021** 1st Virtual Conference for Women Archaeologists and Paleontologists, online  
Oral presentation: Papua New Guineans show unique phenotypic traits at altitude
- 2021** Introduction to the Statistical Analysis of Genome-wide Association Studies, held by the University of Surrey, online

#### Awards:

- 2022** Dora Pluss Scholarship

#### Publications:

- 2023** **M André**, N Brucato, G Hudjasov, V Pankratov, D Yermakovich, R Kreevan, J Kariwiga, J Muke, A Boland, J-F Deleuze, V Meyer, N Evans, M P Cox, M Leavesley, M Dannemann, T Org, M Metspalu, M Mondal, F-X Ricaut, *Positive selection in the genomes of two Papua New Guinean populations at distinct altitude levels*, bioRxiv, [preprint], 2022 Dec, <https://doi.org/10.1101/2022.12.15.520226>
- 2022** N Brucato, **M André**, G Hudjashov, M Mondal, M P Cox, M Leavesley, F-X Ricaut, *Chronology of natural selection in Oceanian genomes*, IScience, 2022 Jun, 25(7), <https://doi.org/10.1016/j.isci.2022.104583>
- 2021** Y Chen, **M André**, K Adhikari, M Blin, B Bonfante, J Mendoza-Revilla, M Fuentes-Guajardo, S Palmal, J C Chacón-Duque, M Hurtado, V Villegas, V Granja, C Jaramillo, W Arias, R B Lozano, P Everardo-Martínez, J Gómez-Valdés, H Villamil-Ramírez, C C S de Cerqueira, T Hünemeier, V Ramallo, R Gonzalez-José, L Schüler-Faccini, M-C Bortolini, V Acuña-Alonzo, S Canizales-Quinteros, C Gallo, G Poletti, G Bedoya, F Rothhammer, D Balding, D J Tobin, S Wang, P Faux 2, A Ruiz-Linares; *A genome-wide association study identifies novel gene associations with facial skin wrinkling and mole count in Latin Americans*; British Journal of Dermatology; 2021 Nov; 18(5):988-998; <https://doi.org/10.1111/bjd.20436>
- 2021** N Brucato, **M André**, R Tsang, L Saag, J Kariwiga, K Sesuki, T Beni, W Pomat, J Muke, V Meyer, A Boland, J-F Deleuze, H Sudoyo, M Mondal, L Pagani, I Gallego Romero, M Metspalu, M P Cox, M Leavesley, F-X Ricaut; *Papua New Guinean Genomes Reveal the Complex Settlement of North Sahul*; Molecular Biology and Evolution; 2021 Oct; 38(11):5107–5121; doi:10.1093/molbev/msab238

- 2021** M André, N Brucato, S Plutniak, J Kariwiga, J Muke, A Morez, M Leavesley, M Mondal, F-X Ricaut; *Phenotypic differences between highlanders and lowlanders in Papua New Guinea*; PLOS ONE; 2021 Jul; 16(7): e0253921; doi:10.1371/journal.pone.0253921
- 2020** N Pedro, N Brucato, V Fernandes, M André, L Saag, W Pomat, C Besse, A Boland, J-F Deleuze, C Clarkson, H Sudoyo, M Metspalu, M Stoneking, M P Cox, M Leavesley, L Pereira, F-X Ricaut; *Papuan mitochondrial genomes and the settlement of Sahul*; Journal of Human Genetics, 2020 Jun; 65:875:887; doi: 10.1038/s10038-020-0781-3

## ELULOOKIRJELDUS

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### Haridustee:

2019–2023 Doktoriõpingud, Genoomika instituut, Loodus- ja täppisteaduste valdkond, Tartu Ülikool  
2017–2019 Magistrikraad (*MSc*), evolutsiooniline antropoloogia (“Master Humanités Médicales”), Aix-Marseille Ülikool (AMU), Prantsusmaa  
2013–2017 Bakalaureusekraad (*BSc*), bioloogia, kõrvaleriala kunstiajalugu ja arheoloogia, Louvaini Katoliiklik Ülikool (UCLouvain), Belgia

### Erialane tööhõive:

2021–2023 Kaasaegse populatsioonigeneetika nooremteadur

### Õpetamine:

2020–2021 Õppeassistent, Evolutsioonilise bioloogia õppetool, Molekulaar- ja Rakubioloogia instituut, Loodus- ja täppisteaduste valdkond, Tartu Ülikool. Bakalaureuseaine “Evolutsioon ja loodus”

### Rahvusvahelised kursused ja konverentsid:

- 2023** Molekulaarbioloogia ja evolutsiooniühingu konverents, Ferrara, Itaalia.  
Poster: Genoomsed mustrid ja piirkondlik kohastumus ekstreemsetele keskkonnatingimustele Paapua Uus-Guinea mäestiku- ja tasandikuelanikel.
- 2023** Genoomide bioloogia, Cold Spring Harbour Laboratory (CSHL), USA  
Poster: Paapua Uus-Guinea eri kõrgustel elavatele populatsioonidele mõjuv positiivne valik .
- 2022** India ja Vaikse ookeani piirkonna esiajaloo ühingu kongress (IPPA), Chiang Mai, Taimaa  
Suuline ettekanne: Kõrgusest tingitud fenotüübi kohastumine ja positiivne valik Paapua Uus-Guinea mäestikuelanikel.
- 2022** EMBO | EMBL Sümpoosium: Inimese mineviku rekonstrueerimine vana-DNA abil. Heidelberg, Saksamaa  
Välkettekannet ja poster: Positiivse valiku mõju Paapua Uus-Guinea elanike verekoostisele.
- 2021** Austraalia arheoloogide ühingu konverents, veebipõhine.  
Suuline ettekanne: Paapua-uusguinealased on kohanenud kõrgel elamiseks.
- 2021** Wellcome Connecting Science konverents: Inimese evolutsioon – fossiilidest vanade ja kaasaegsete genoomideni, veebipõhine.  
Poster: Looduslik valik ja fenotüüpide seosed Paapua mäestikuelanikel.

- 2021** ASHG virtuaalkonverents 2021, veebipõhine  
Poster: Positiivne valik mäestikes elamiseks ja fenotüübi seosed Paapua Uus-Guinea mäestikuelanikel.
- 2021** EMBO kursus: Populatsioonigenoomika: taust, tööriistad ja programmid.  
Veebipõhine.
- 2021** Esimene virtuaalkonverents naisarheoloogidele ja -paleontoloogidele.  
Veebipõhine.  
Suuline ettekanne: Paapua Uus-Guinea mäestikuelanikeisloomulikud tunnused
- 2021** Sissejuhatus ülegenoomsete assotsiatsiooniuuringute statistikameetoditesse.  
Surrey Ülikool, veebipõhine.

**Teaduspreemiad ja tunnustused:**

**2022** Dora Pluss stipendium

**Publikatsioonid:** Loetletud inglisekeelse CV rubriigis publikatsioonid ('Publications')

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1. **Toivo Maimets.** Studies of human oncoprotein p53. Tartu, 1991, 96 p.
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