

JAANUS SUUMANN

Gastric biomarkers and their dynamics  
as a less invasive method to  
evaluate stomach health  
in bariatric surgery patients



DISSERTATIONES MEDICINAE UNIVERSITATIS TARTUENSIS

**353**

**JAANUS SUUMANN**

Gastric biomarkers and their dynamics  
as a less invasive method to  
evaluate stomach health  
in bariatric surgery patients



UNIVERSITY OF TARTU

Press

Department of Surgery, Institute of Clinical Medicine, University of Tartu, Tartu, Estonia

Dissertation is accepted for the commencement of the degree of Doctor of Philosophy (Medicine) on September 1<sup>st</sup> 2023 by the Council of the Faculty of Medicine, University of Tartu, Estonia

Supervisors: Associate Professor Toomas Sillakivi, MD, PhD  
Department of Surgery, Institute of Clinical Medicine,  
Faculty of Medicine, University of Tartu, Tartu, Estonia  
Head of the Division of Abdominal Surgery, Tartu University  
Hospital, Estonia

Professor Ants Peetsalu, MD, PhD (*In memoriam*)  
Institute of Clinical Medicine, University of Tartu, Tartu, Estonia

Reviewers: Professor Jaak Kals, MD, PhD  
Department of Surgery, Institute of Clinical Medicine,  
Faculty of Medicine;  
Department of Biochemistry, Institute of Biomedicine and  
Translational Medicine, University of Tartu, Tartu, Estonia  
Head of the Division of Vascular Surgery, Tartu University  
Hospital, Estonia

Professor Aare Märtson, MD, PhD  
Department of Traumatology and Orthopedics, Institute of Clinical  
Medicine, Faculty of Medicine, University of Tartu, Estonia  
Head of 1 Clinical Area, Tartu University Hospital, Estonia

Opponent: Professor Almantas Maleckas, MD, PhD  
Department of Surgery, Faculty of Medicine, Lithuanian  
University of Health Sciences, Kaunas, Lithuania  
Department of Gastrosurgical Research and Education,  
Institute of Clinical Sciences, Sahlgrenska Academy,  
University of Gothenburg, Gothenburg, Sweden  
Abdominal surgeon (bariatric surgeon), Sahlgrenska University  
Hospital, Gothenburg, Sweden

Commencement: 24<sup>st</sup> of November 2023

Publication of this dissertation is granted by the University of Tartu.

ISSN 1024-395X (print)

ISSN 2806-240X (pdf)

ISBN 978-9916-27-360-9 (print)

ISBN 978-9916-27-361-6 (pdf)

Copyright: Jaanus Suumann, 2023

University of Tartu Press  
www.tyk.ee

# TABLE OF CONTENTS

|   |    |
|---|----|
| LIST OF ORIGINAL PUBLICATIONS .....   | 7  |
| ABBREVIATIONS .....   | 8  |
| 1. INTRODUCTION .....   | 9  |
| 2. REVIEW OF THE LITERATURE .....   | 11 |
| 2.1. Definition of overweight and obesity .....   | 11 |
| 2.2. Prevalence of overweight and obesity .....   | 11 |
| 2.3. The burden of obesity .....  | 11 |
| 2.4. Possible causes of obesity .....   | 12 |
| 2.5. Treatment of obesity .....   | 13 |
| 2.5.1. Lifestyle interventions .....  | 13 |
| 2.5.2. Medical therapy .....  | 14 |
| 2.5.3. Surgery for obesity .....  | 15 |
| 2.5.3.1. Indications for bariatric surgery .....  | 15 |
| 2.5.3.2. Bariatric surgical procedures .....  | 15 |
| 2.5.3.3. Mechanisms of weight loss in bariatric operations ...  | 20 |
| 2.5.3.4. Selection of the operation method .....  | 20 |
| 2.5.3.5. Effectiveness of bariatric surgery .....   | 21 |
| 2.5.3.6. Cost-effectiveness of surgical versus non-surgical<br>treatment for obesity .....              | 23 |
| 2.6. Complications of bariatric surgery .....   | 23 |
| 2.6.1. Surgical complications .....   | 23 |
| 2.6.2. Nutrient deficiencies after bariatric surgery and<br>postoperative replacement prophylaxis ..... | 24 |
| 2.7. Anatomy and physiology of the stomach .....  | 24 |
| 2.7.1. Anatomy of the stomach .....   | 24 |
| 2.7.2. Physiology of the stomach .....  | 24 |
| 2.8. Gastric biomarkers .....   | 26 |
| 2.9. Interpretation of the biomarker profile .....  | 28 |
| 3. SUMMARY OF THE LITERATURE REVIEW .....   | 31 |
| 4. AIMS OF THE STUDY .....  | 32 |
| 5. MATERIAL AND METHODS .....   | 33 |
| 5.1. Patients .....   | 33 |
| 5.2. Inclusion and exclusion criteria .....   | 33 |
| 5.3. Patient questionnaire .....  | 33 |
| 5.4. Biomarker test examination .....   | 34 |
| 5.5. Interpretation of GastroPanel .....  | 34 |
| 5.6. Esophagogastroduodenoscopy .....   | 35 |
| 5.7. Histology and H. pylori .....  | 35 |
| 5.8. Surgical procedures .....  | 35 |

|  |     |
|--|-----|
| 5.9. Statistics .....  | 36  |
| 5.10. Study groups .....   | 36  |
| 6. RESULTS .....   | 38  |
| 6.1. Correlation between gastric biomarkers and gastrobiopsies<br>(Publication II) .....   | 38  |
| 6.2. Changes in the biomarker profile in different operation groups<br>(LRYGB and LSG) during 2- year follow-up after bariatric surgery<br>(Publications III, I) ..... | 40  |
| 7. DISCUSSION .....  | 46  |
| 8. CONCLUSIONS .....   | 54  |
| 9. SUMMARY IN ESTONIAN .....   | 55  |
| REFERENCES .....   | 59  |
| ACKNOWLEDGMENTS .....  | 69  |
| PUBLICATIONS .....   | 71  |
| CURRICULUM VITAE .....   | 100 |
| ELULOOKIRJELDUS .....  | 102 |

## LIST OF ORIGINAL PUBLICATIONS

This thesis is based on the following original publications, which are referred to in the text by their Roman numerals (I–III):

- I Sillakivi T, Suumann J, Kirsimägi U, Peetsalu A. Plasma levels of gastric biomarkers in patients after bariatric surgery: biomarker after bariatric surgery. *Hepatogastroenterology*. 2013 Nov-Dec;60(128):2129–32.
- II Suumann J, Sillakivi T, Riispere Ž, Syrjänen K, Sipponen P, Kirsimägi Ü, Peetsalu A. Serological biomarker testing helps avoiding unnecessary endoscopies in obese patients before bariatric surgery. *BMC Obes*. 2018 Feb 20;5:9.
- III Suumann J, Sillakivi T, Riispere Ž, Syrjänen K, Sipponen P, Kirsimägi Ü, Peetsalu A. Serum gastric biomarker levels after sleeve gastrectomy and Roux-en-Y gastric bypass operations: a prospective study with 1-year follow-up. *Surgery Gastroenterology and Oncology*. 2019 Apr 24(2):86

### **Author's contribution:**

Involvement in the study design, in patients' recruitment and follow-up, in collecting clinical data, in participating in majority of the operations, and in writing of the text of the manuscripts.

## ABBREVIATIONS

|         |  |
|---------|--|
| AG      | atrophic gastritis (mucosal atrophy)                 |
| AGA     | atrophic gastritis of the antrum                     |
| AGC     | atrophic gastritis of the corpus                     |
| AGP     | atrophic pan-gastritis (AG of the antrum and corpus) |
| BMI     | body mass index                                      |
| EGDS    | esophagogastroduodenoscopy                           |
| %EWL    | percentage of excess weight loss                     |
| GERD    | gastroesophageal reflux disease                      |
| G17b(f) | basal (fasting) Gastrin 17                           |
| G17s    | stimulated amidated Gastrin 17                       |
| GP      | GastroPanel  |
| HPAB    | Helicobacter pylori antibodies                       |
| HP      | Helicobacter pylori                                  |
| HS      | healthy stomach                                      |
| LA      | Los Angeles classification of esophagitis            |
| LRYGB   | laparoscopic Roux-en-Y gastric bypass                |
| LSG     | laparoscopic Sleeve Gastrectomy                      |
| NHS     | non-healthy stomach                                  |
| PgI     | Pepsinogen I   |
| PgII    | Pepsinogen II  |
| RYGB    | Roux-en-Y gastric bypass                             |
| SG      | Sleeve Gastrectomy                                   |
| USS     | updated Sydney System                                |

# 1. INTRODUCTION

The prevalence of overweight and obesity has increased over several decades becoming an increased health risk in modern society both in developed and developing countries. Nowadays it has become priority in healthcare all over the world [Ogden et al., 2006; Benotti et al., 1995; Williams et al., 2015; Kelly et al., 2013]. According to the World Health Organization, in 2016 more than 1.9 billion adults 18 years of age and older were overweight. Of these over 650 million were obese [WHO, 2021].

Overweight and obesity are risk factors for gastroesophageal reflux disease, erosive esophagitis, hiatal hernia, Barrett's esophagus, esophageal adenocarcinoma, *Helicobacter pylori* infection. Most of these conditions are observed up to 2–3 times more frequently in obese patients than in normal weight persons [Gerson LB., 2009].

In parallel to the growth of obese people, the number of bariatric operations has also increased, since it is the the only effective way to treat obesity, resulting in sustainable and significant weight loss in the long term (along with the resolution of metabolic comorbidities in up to 80% of cases) [Flum et al., 2007 ; Azagury et al., 2011].

Operations on the stomach alter more or less the gastric mucosa and gastric secretory functions in gastric remnants, as was shown earlier after gastric vagotomies or partial gastric resections in peptic ulcer cases [Peetsalu et al., 1990]. However, information about these changes and association with postoperative morbidity in the bariatric cohort is limited.

Upper digestive endoscopy with gastrobiopsies has been the gold standard in pre- and postoperative investigation of bariatric patients to evaluate the state of the stomach and to detect possible abnormalities. Nevertheless, the value of and the need for routine endoscopy in asymptomatic patients remains controversial [Parikh et al., 2016; Csendes et al., 2007; D'Hondt et al., 2013; Almazeedi et al., 2013; Gerson LB., 2009].

During past decades, the use of serological biomarker testing (“serological biopsy”) has gained increasing popularity as a non-invasive diagnostic tool for dyspeptic patients and asymptomatic subjects to diagnose both functional disorders and gastric diseases, including *Helicobacter pylori* (HP) infection and mucosal atrophy (AG) [Agreus et al., 2012]. The latest innovation in this technology represents a panel of 4 stomach-specific biomarkers (Pepsinogen I and II, Gastrin-17 and HP antibody) (Pgl, PglI, G-17, HPAP) known as Gastro-Panel® (GP) test, which distinguishes between 8 diagnostic marker profiles [Agreus et al., 2012; Syrjänen et al. 2019].

The knowledge of alterations in stomach physiology after bariatric surgery is an important clinical issue. The sequelae caused by resections or bypasses are persistent and will remain and act over many postoperative years or even decades. Since the number of bariatric operations is growing worldwide, the possibility to evaluate the “health” of gastric mucosa pre- and postoperatively,

using noninvasive methods, would be very attractive, especially in patients after (mini) gastric bypass operation.

In our studies we aimed to find out if serological biomarker testing could avoid invasive endoscopies in obese patients undergoing for bariatric surgery and to characterize the gastric biomarker profile and its dynamics after sleeve gastrectomy (SG) and Roux-en-Y gastric bypass (RYGB). Identifying changes in the gastric biomarker profile could provide important new information about possible complications and complaints after bariatric surgery, as well as could help to institute individualised therapy, including replacement therapy, after specific types of bariatric operation.

## **2. REVIEW OF THE LITERATURE**

### **2.1. Definition of overweight and obesity**

The World Health Organization (WHO) defines overweight and obesity as abnormal or excessive fat accumulation that presents a risk to health [WHO, 2021]. Body mass index (BMI), calculated by dividing body weight in kilograms by the square of height in meters, is a simple metric used to indicate overall body fatness [WHO, 2021]. For adults, current guidelines from the US Centers for Disease Control and Prevention (CDC) and the WHO define a normal BMI range as 18.5 to 24.9, whereas a BMI of  $\geq 25$  kg/m<sup>2</sup> is considered to be overweight, and a BMI of  $\geq 30$  kg/m<sup>2</sup> is classified as obese, with severe obesity defined as a BMI of  $\geq 40$  kg/m<sup>2</sup>. For children, age needs to be considered when defining overweight and obesity [WHO, 2021; Chooi et al., 2019].

### **2.2. Prevalence of overweight and obesity**

According to WHO, in 2016, 39% (1.9 billion) adults aged 18 years and older (39% of men and 40% of women) were overweight and about 13% (650 million) of the world's adult population (11% of men and 15% of women) were obese. Overweight and obesity are serious problems among children: in 2016, over 340 million children and adolescents aged 5–19 were overweight or obese. In 2019, an estimated 38.2 million children under the age of 5 years were overweight or obese [WHO, 2021]. The data published in 2015 by Ogden and colleagues showed that more than one-third of adults and 17% of adolescents in the United States were obese in 2011–2015 [Ogden et al., 2015]. According to the data from the National Institute for Health Development, 32.4% of adults (aged 16–64) were overweight and 20% obese in Estonia in 2020 [TAI, 2021]. Of the adolescent population (aged 11–15), 17.1% were overweight/obese in 2018 [TAI, 2019].

### **2.3. The burden of obesity**

Obesity represents a major health challenge because it substantially increases the risk of the diseases such as type 2 diabetes mellitus (T2DM), fatty liver disease, hypertension, myocardial infarction, stroke, dementia, osteoarthritis, obstructive sleep apnoea and several cancers, thereby contributing to a decline in both quality of life and life expectancy. Obesity is also associated with unemployment, social disadvantages and reduced socio-economic productivity, thus increasingly creating an economic burden [Blüher et al., 2019].

In 2016 the total cost of chronic diseases due to obesity and overweight in the U.S. was \$1.72 trillion—equivalent to 9.3 percent of the U.S. gross domestic product (GDP) [Waters et al 2018].

Importantly, the World Obesity Federation and other organizations, including the American and Canadian Medical Associations, have declared

obesity a chronic progressive disease clearly distinct from being just a risk factor for other diseases [Bray et al., 2017].

Reducing the obesity-related burden to health and societies as well as reversing the increase in obesity prevalence is a high priority for the WHO, which included the target to halt obesity prevalence at the level it was in 2010 as one of the main targets of the ‘Global Action Plan for the Prevention and Control of Noncommunicable Diseases 2013–2020’ [WHO, 2021].

## 2.4. Possible causes of obesity

The fundamental cause of obesity is a long-term energy imbalance between too many calories consumed and too few calories expended. Global estimates, based on food balance sheets, indicate that the available per capita food in developed countries was 3138 kcal/person/day in 1969/1971 and 3360 kcal/person/day between 2005 and 2007. In developing countries, the per capita food available increased from 2055 to 2619 kcal/person/day during the same three and a half decades [Alexandratos et al., 2012].

Although obesity prevalence increased in every single country in the world, regional differences exist in both obesity prevalence and its trends. Changes in the global food system together with increased sedentary behaviour seem to be the main drivers for the obesity pandemic [Blüher, 2019].

Most areas of the world have shown economic development that has resulted in increased purchasing power which is an important factor in explaining the increasing caloric intake [Gerbens-Leenes et al., 2010].

A study by Vandevijvere et al. found that a higher energy intake was sufficient to explain the increasing population body weight, especially in high-income countries [Vandevijvere et al., 2015].

Biological factors play also a role in the pathogenesis of obesity. The key role of certain brain regions in the regulation of body weight became evident from observations that animals with lesions and humans with tumours, affecting the hypothalamus, develop abnormal food-seeking behaviour and obesity [Farooqi, 2014].

A mutation in the *ob* gene (which encodes the adipose tissue hormone leptin) causes severe obesity in *ob/ob* mice, therefore it became apparent that the central neural circuits that control energy homeostasis integrate signals from peripheral tissues such as the adipose tissue [Zhang et al. 1994; Coleman et al., 1969; Farooqi et al., 2014]. Mutations in the genes coding for leptin, leptin receptor, melanocortin 4 receptor, pro-opiomelanocortin and others might cause severe obesity in humans, which underlines the importance of biological factors in the pathogenesis of obesity [Montague et al., 1997; Clément et al., 1998]. As monogenetic mutations are rare, changes in population genetics cannot explain the rise in obesity prevalence in just 40 years.

Individual factors (such as the genetic background or the gut–brain–hormone axis) influence susceptibility to obesity, which may develop in an obesogenic environment (for example, influenced by eating culture, transportation and

computerization). Obesity is not caused by personal choice or by society, but rather by the relationship between an individual and their environment [Blüher, 2019]. Obesity is the result of the interplay between heterogenic factors, deriving from a person's eating behaviour, physical activity and individual energy expenditure determinants [Butland et al., 2007].

Several studies have demonstrated differences in the microbiome composition of obese, overweight, and lean individuals [Turnbaugh et al. 2009]. Although the exact mechanisms of the action of gut microbes on obesity are not fully known, the diversity of microorganisms in the human gastrointestinal tract is very important. Gut microbes (microbiota) may influence human/host body weight in several ways: promoting increased fat storage in the adipose tissue, processing indigestible luminal polysaccharides into short chain fatty acids, influencing the absorption of nutrients and production of hormones (ghrelin, leptin, glycagon-like peptide-1 (GLP-1)) [Mulla et al. 2018]. In an experiment scientists collected the fecal microbiota from identical adult female twin pairs discordant for obesity and transplanted it into a germ-free mice gastrointestinal tract. The mice harboring the obese twin's microbiota became/turned obese, but the mice harboring the lean co-twin's microbiota did not [Ridaura et al. 2013].

Bile acids also play an important role in body weight regulation being mediators of intestinal absorption of lipophilic nutrients. They are also considered mediators of systemic metabolism, serving as ligands for farnesoid X receptor (FXR) and Takeda G-protein-coupled receptor 5 (TGR5). In the gut lumen they can regulate the production of incretin hormones, such as GLP-1. Bile acids also influence the microbiome through the antimicrobial properties of conjugated bile species. Bile acids can increase the circulating levels of fibroblast growth factor-19 (FGF-19) in the postprandial state, which in turn regulates glucose and lipid metabolism, as well as bile acid synthesis. The plasma levels of both primary and secondary bile acids are increased after laparoscopic gastric bypass (LRYGB) and laparoscopic sleeve gastrectomy (LSG) [Mulla et al. 2018].

## **2.5. Treatment of obesity**

### **2.5.1. Lifestyle interventions**

Lifestyle and behavioural interventions aimed at reducing calorie intake and increasing energy expenditure have limited effectiveness, because complex and persistent hormonal, metabolic and neurochemical adaptations defend against weight loss and promote weight regain [Blüher, 2019]. Speakman et al. demonstrated that the energetic balance is regulated in a way to maintain the target value („set point“). Discrepancies between the level of the signal and the target are translated into effects on energy expenditure and energy intake to equalise the discrepancy and maintain homeostasis [Speakman et al., 2011].

Studies on lifestyle interventions in the treatment of obesity usually have a short observation period (1–2 years) and show a modest effect on weight loss. Weight regain is common after the conclusion of a lifestyle intervention, as

observed in a study by Foster et al. [Foster et al., 2010]. Wadden et al. demonstrated that with no further treatment patients typically regain one third of lost weight in the first follow-up year, with continuing weight gain thereafter, and return to their baseline weight within 4–5 years. About half of patients receive additional weight compared to baseline weight [Wadden et al., 1989].

### 2.5.2. Medical therapy

The panoply of available medications for weight loss has evolved significantly in the last half century. The Food and Drug Administration (FDA) has approved weight loss medications dating back to 1959 for short-term (<12 weeks) use. With increased recognition of obesity as a chronic, complex medical disease, newer agents have been approved as long-term therapy, and the cornerstone of treatment is chronic behavior and lifestyle change [Velazquez et al., 2018].

As indicated in the 2015 Endocrine Society Clinical Practice Guidelines on Pharmacologic Management of Obesity, all patients with a body mass index (BMI) >25 kg/m<sup>2</sup> warrant intervention with diet, exercise, and behavior modification. Weight loss medications should be introduced as adjuncts to diet, exercise, and behavioral modification for patients with a BMI >30 kg/m<sup>2</sup> or patients with a BMI >27 kg/m<sup>2</sup>, if they have at least one obesity related comorbid condition, such as T2DM, dyslipidaemia, hypertension, sleep apnea. The objective of using pharmacotherapy in management of obesity is to amplify patient adherence to lifestyle changes and to overcome the biological adaptations that occur with weight loss. Most anti-obesity drugs have an efficacy of 3–7% (estimated net weight loss). The usage of these medications is limited because of their adverse effects [Apovian et al. 2015; Srivastava et al., 2018].

Major FDA-approved anti-obesity medications are: Phentermine, Orlistat, Phentermine/topiramate extended release (ER), Naltrexone sustained release (SR)/bupropion sustained release (SR), Liraglutide and Semaglutide. Phentermine is a noradrenergic drug that acts on the sympathetic nervous system, causing an increase in norepinephrine release. This neurotransmitter release leads to appetite suppression in addition to increased resting energy expenditure [Rothman et al. 2001]. Orlistat inhibits the activity of pancreatic and gastric lipase, thereby decreasing fat absorption by 30%. Orlistat is indicated for weight loss in conjunction with a reduced-calorie diet [Tak et al., 2021; Apovian et al., 2016]. Phentermine/topiramate ER is the first combination medication approved for chronic management of obesity. Topiramate ER acts on GABA receptors, leading to appetite suppression. The combination drug has shown greater potential weightloss effects than monotherapy alone for each, while also reducing adverse effects [Tak et al., 2021; Apovian et al., 2016]. Naltrexone SR/bupropion SR– the mechanism of action for bupropion SR is inhibition of dopamine and norepinephrine reuptake. Naltrexone acts to antagonize the feedback loop that limits bupropion's anorexic effects; hence, this drug combination works synergistically [Greenway et al., 2009]. Liraglutide and semaglutide are analogs of human glucagon-like peptide-1 (GLP-1)

with a much longer half-life of 13 h compared with the human endogenous GLP-1 that lasts only a few minutes. These drugs mimic the actions of endogenous GLP-1, a hormone released from the small intestine, which plays a role in the peripheral regulation of appetite through anorexigenic effects in addition to effects of increasing release of insulin from the pancreas in the presence of glucose [van Can *et al.* 2014].

The latest studies with semaglutide (analog of human GLP-1) have shown promising results in weight loss. In a systematic literature review with meta-analysis Smith *et al.*, demonstrated that semaglutide 2.4 mg weekly is associated with a greater weight change from baseline (>5%) compared to pharmacological comparators for weight management in overweight or obesity [Smith *et al.*, 2022].

Comparative studies of medical treatment versus surgery are still in progress and long-term results are not known yet.

### 2.5.3. Surgery for obesity

#### 2.5.3.1. Indications for bariatric surgery

Bariatric surgery is indicated to patients older than 18 years having the following characteristics:

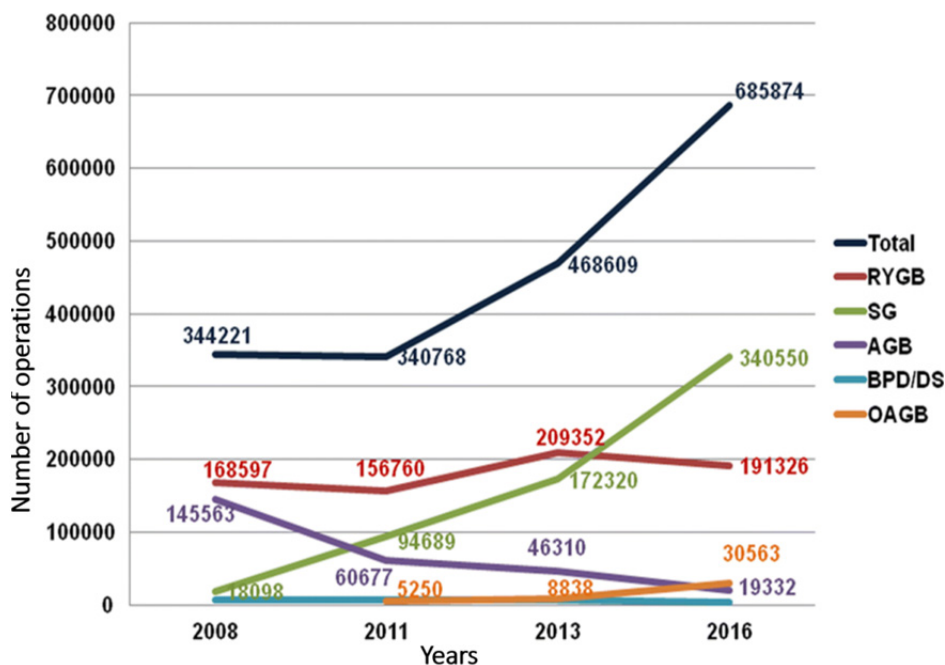
- BMI  $\geq 40$  kg/m<sup>2</sup>
- BMI 35–40 kg/m<sup>2</sup> with co-morbidities in which surgically induced weight loss is expected to improve the disorder (such as metabolic disorders, cardiovascular diseases (myocardial infarction, stroke), respiratory diseases (chronic obstructive pulmonary disease, sleep apnea), severe osteoarthritis, obesity related severe psychological problems, polycystic ovarian syndrome).
- Patients with BMI  $>30 < 35$  kg/m<sup>2</sup> with T2DM may be considered for bariatric surgery on an individual basis, as there is evidence-based data supporting bariatric surgery benefits in regard to T2DM remission or improvement [Fried *et al.*, 2014; Di Lorenzo *et al.* 2020].

The above mentioned indications, which have been used for almost 30 years (with minor changes), are currently under adjustment, as the new guidelines of the American Society for Metabolic and Bariatric Surgery (ASMBS) and the International Federation for the Surgery of Obesity and Metabolic Disorders (IFSO) 2022 were released. These new guidelines will increase the proportion of people eligible for bariatric surgery [Eisenberg *et al.*, 2022].

#### 2.5.3.2. Bariatric surgical procedures

In parallel to the growth of obese people, the number of bariatric operations has also increased, since it is the the only effective way to treat obesity in the long term. A variety of surgical options are currently offered for treatment of obesity. The beginning of the third millennium has seen constant evolution of invasive techniques, such as laparoscopy and flexible transoral endoscopy [Lee, 2017].

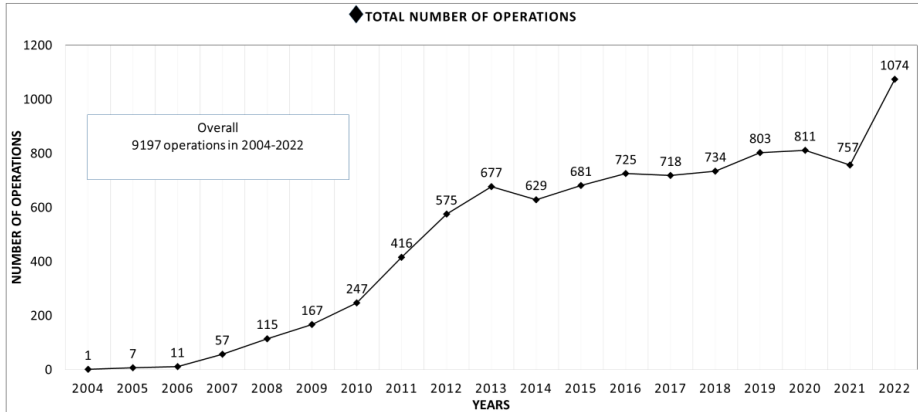
According to the IFSO survey, the total number of primary bariatric operations performed in 2016 was 609,897 worldwide. The most frequently performed surgical bariatric/metabolic procedure was sleeve gastrectomy (SG) (N=340,550), followed by Roux-en-Y gastric bypass (RYGB) (N=191,326), One-Anastomosis Gastric Bypass (OAGB) (N=30,563), Adjustable Gastric Banding (AGB) (N=19,332) and Biliopancreatic Diversion/Duodenal Switch (BPD/DS) (N=3346) [Angrisani et al., 2018] (**Figure 1**).



**Figure 1.** Number of main primary bariatric/metabolic surgical procedures from 2008 to 2016 worldwide. AGB adjustable gastric banding, RYGB Roux-en-Y gastric bypass, SG sleeve gastrectomy, BPD-DS biliopancreatic diversion-duodenal switch, OAGB one-anastomosis gastric bypass [Angrisani et al., 2018] (Reproduced with the permission of Springer Nature)

The first bariatric operation in Estonia (adjustable gastric banding) was performed at the North Estonia Medical Center in 2004, since then their number has increased (**Figure 2**). In 2021 there were 757 bariatric operations (569 operations per one million inhabitants), the two most favored type operations being RYGB (N=323) and SG (N=219). In 2022 the total number was 1,074 [EBMKS, 2023] (**Figure 2, 3**). The total number of bariatric/metabolic procedures performed in Estonia between 2004 and 2022 was 9,197 (**Figure 2**).

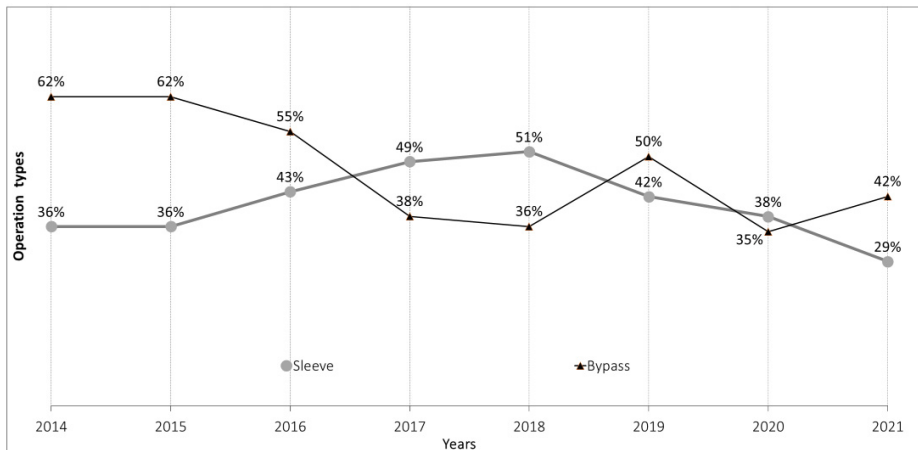
## Bariatric operations in Estonia



**Figure 2.** Number of bariatric operations in Estonia in 2004–2022 [EBMKS, 2023]

## Estonian trends

Sleeve vs Bypass



**Figure 3.** Bariatric/metabolic surgical procedures in Estonia in 2004–2021. Sleeve – Sleeve gastrectomy; Bypass – Roux-en-Y gastric bypass [EBMKS, 2023]

### Sleeve gastrectomy (SG).

SG was performed for the first time in 1988 as part of the biliopancreatic diversion with duodenal switch (BPD-DS). SG is a restrictive type bariatric operation where most of the corpus and antrum is resected and removed to create a long, tubular gastric conduit along the lesser curve of the stomach

**(Figure 4A).** It was first used in a two-staged approach in an attempt to reduce surgical risk in superobese patients with SG as the first stage. Since 2007 SG has been used as a primary bariatric procedure. The main strengths of SG are the low rate of complications, short operative time, absence of foreign material, lack of gastrointestinal anastomosis and malabsorption, patient's acceptance, and feasibility to be converted to multiple other bariatric procedures [Angrisani et al., 2018].

#### **Roux-en-Y gastric bypass (RYGB).**

Laparotomic Roux-en-Y gastric bypass (RYGB) was described for the treatment of morbid obesity in 1966 by Mason and Ito, who proposed it for treating morbid obesity. It is a combined (restrictive and malabsorptive) method where by separating the upper part of the stomach a new smaller stomach (pouch) is created. The small intestine is divided and one end is anastomosed to the pouch. A second anastomosis is made to connect the disconnected stomach and duodenum to the small bowel. This connection enables the digestive fluids to meet ingested food enabling nutrient breakdown and absorption. The distance between the two connections can vary, but is generally 50 to 150 cm. Laparoscopic RYGB was first described by Wittgrove et al. in 1994. It was the most popular type bariatric operation for decades and is still considered by many as the gold standard [Angrisani et al., 2018] **(Figure 4B).**

#### **Biliopancreatic Diversion-Duodenal Switch (BPD-DS).**

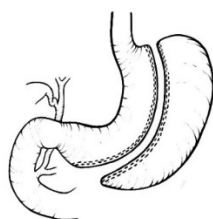
Biliopancreatic diversion was described by Scopinaro in 1976. This procedure consists in a horizontal distal gastrectomy with a proximal gastric pouch with closure of the duodenal stump, a gastroileostomy, with a 250-cm limb of the distal ileum, and a biliopancreatic limb anastomosed to the distal ileum, creating a 50-cm common channel. In 1988 Hess modified BPD to BPD-DS where the most important changes involved use of vertical gastrectomy and division of the duodenum between the pyloric valve and the sphincter of Oddi **(Figure 4C).** Deriving from experience with BPD, BPD-DS has a longer common channel to reduce the likelihood of vitamin, mineral, and protein deficiencies. The first laparoscopic BPD-DS was performed in 1999 [Angrisani et al., 2018].

#### **One-Anastomosis Gastric Bypass (OAGB).**

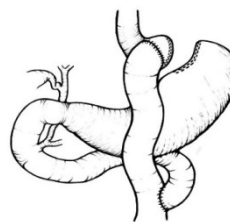
OAGB was first reported in 2001 by Rutledge. It is a combined (restrictive and malabsorptive) method performed with the laparoscopic technique. During the OAGB procedure, the surgeon first reduces the size of the "working" stomach by separating a tube-like pouch of the stomach from the rest of the stomach. This tubular gastric pouch is then anastomosed to the intestine, bypassing up to 200cm of the upper part of the intestine. This technique differs from the traditional RYGB which requires two anastomoses [Angrisani et al., 2018] **(Figure 4D).**

### Adjustable gastric banding (AGB).

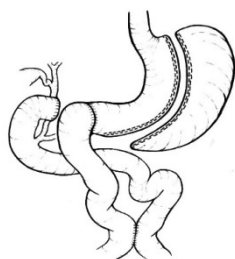
AGB is a restrictive type bariatric procedure first described by Kuzmak et al. in 1986. During the AGB procedure a silicone band (equipped with a firmly attached inflatable balloon) is placed around the upper part of the stomach, creating a 15- 30ml pouch. The size of the pouch outlet is adjusted by adjusting the volume of fluid in the balloon through the access port situated under the skin usually in the upper abdomen. The first band was placed laparoscopically in a human by Belachew and colleagues in September 1993 [Angrisani et al., 2018] (Figure 4E).



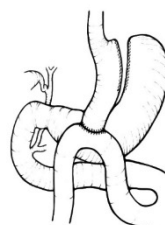
A Sleeve gastrectomy (SG)



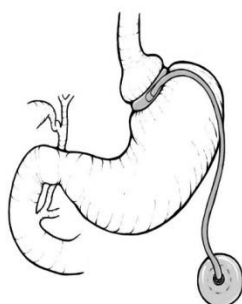
B Roux-en-Y gastric bypass (RYGB)



C Biliopancreatic Diversion-Duodenal Switch (BPD-DS)



D One Anastomosis Gastric Bypass (OAGB)



E Adjustable gastric banding (AGB)

**Figure 4.** Graphic depiction of bariatric surgery types (Figure by T. Veršinina)

### 2.5.3.3. Mechanisms of weight loss in bariatric operations

The mechanisms of bariatric operations contributing to weight loss can vary in the different type procedures, being often combined. These mechanisms include effects on regulation of energy balance, both central and peripheral nervous systems` regulation of appetite and metabolism, but also changes in gut hormones and microbioma [Mulla et al., 2018]. Bariatric surgery changes the secretion of gut hormones (incretines) which play a key role in regulating appetite, satiety, food intake, systemic metabolism, and insulin secretion [Campbell et al., 2013].

The most important gut hormones are:

- GLP-1 (glucagon-like peptide-1). This L-cell derived hormone increases glucose-dependent insulin secretion, inhibits clycagon secretion, confers glucose sensitivity to glucose-resistant  $\beta$  cells, stimulates  $\beta$  cell proliferation and neogenesis, and inhibits  $\beta$  cell apoptosis. GLP-1 analogues are effective therapeutics for human T2D and obesity. GLP-1 is increased after RYGP (in the fasting and postprandial states`) and LSG (in the postprandial state).
- PYY (Peptide YY). Secreted from L-cells of the distal small intestine and the colon. PYY levels are increased in the postprandial state after RYGB and LSG, and they act on reducing food intake.
- FGF-19 (fibroblast growth factor-19) – peptide hormone that regulates bile acid synthesis, as well as glucose and lipid metabolism.
- GIP (Glucose-dependent insulinotropic polypeptide) – peptid hormone secreted by K cells in the proximal small intestine. GIP signalling increases glucose dependent insulin secretion, postprandial glucagon secretion, and intestinal glucose absorbtion. GIP action in the adipose tissue promotes storage, with increased glucose uptake, conversion of clucose to fatty acids, and activation of lipoprotein lipase. Postprandial levels of GIP are reduced after RYGB.
- Ghreline – orexigenic hormone, produced in oxyntic glands in the gastric fundus. Its concentrations increase in the fasted state and decreases in the postprandial state. Ghrelin levels are suppressed following bariatric surgery in some studies.
- Oxyntomodulin – anorexigenic hormone that is increased after a glucose load. Its levels are high after RYGP [Mulla et al.,2018].

### 2.5.3.4. Selection of the operation method

Opting for the appropriate operation type for weight loss continues to be a challenging and subjective process. In experienced hands, most operations can be successful in providing a given patient with successful weight loss, amelioration of comorbidities and improvement in quality in life [Needleman et al. 2008]. Currently, laparoscopic Roux-en-Y gastric bypass (LRYGB) and laparoscopic sleeve gastrectomy (LSG) are the most commonly performed bariatric procedures worldwide to treat obesity [Angrisani et al., 2021].

In a systematic review (5 RCT) of Colquitt et al., LRYGB had no superiority to LSG in the percentage of excess weight loss (%EWL) [Colquitt et al., 2014]. According to the study of Peterli et al. (SM-BOSS), there was no significant difference in excess BMI loss between LSG and LRYGB at 5-year follow-up (for LSG, 61.1%, vs LRYGB, 68.3% [Peterli et al. 2018]. Similar results were obtained in the SLEEVEPASS trial (the estimated mean %EWL at 10 years was 43.5% after sleeve gastrectomy and 50.7% after gastric bypass) [Salminen et al., 2022]. For superobese patients (BMI>50), biliopancreatic diversion with duodenal switch resulted in greater weight loss than RYGB (2RCT;  $P<0.001$ ) [Colquitt et al., 2014].

Gastroesophageal reflux disease (GERD) can certainly be one of the key points in the decision making process. Although several studies [Pallati et al. 2014; Sucandy et al., 2015] have demonstrated improvement in GERD after LSG, some studies report the opposite [Melissas et al., 2015; Sheppard et al. 2015]. A recent study of Csendes et al. demonstrates „de novo“ GERD in 58.5% of patients (without reflux symptoms and normal endoscopy preoperatively), and worsening endoscopic parameters in patients with preoperative GERD symptoms 10 years after LSG [Csendes et al. 2019]. This is also supported by a paper of Genco et al., where the Barrett mucosa is present in up to 17% of asymptomatic patients 5 years after LSG [Genco et al., 2017]. In a SM-BOSS study Peterli et al. demonstrate that GERD symptoms worsened more often after LSG than after LRYGB (31.8% vs 6.3%;  $P=0.006$ ). In the same study de novo reflux symptoms occurred in 31.6% of the LSG patients compared to 10.7% in the LRYGB group ( $P=0.01$ ) [Peterli et al., 2018]. In a SLEEVE-PASS study the authors reported that at 10 years, esophagitis was more prevalent after LSG than after LRYGB (31% vs 7%;  $P<0.001$ ) while the Barrett esophagus was described in 4% of patients (in both groups) [Salminen et al., 2022].

According to the current conditional recommendations of the Interdisciplinary European Guidelines on Metabolic and Bariatric Surgery, LSG should not be performed in patients with severe GERD symptoms and/or severe oesophagitis [Di Lorenzo et al., 2020].

#### 2.5.3.5. Effectiveness of bariatric surgery

Effectiveness and durability are considered the key attributes of bariatric surgery when compared with the non-surgical approaches to achieving weight loss. During the past three decades, bariatric surgery has gained an increasingly important role in the management of the most severe cases of obesity, since it is practically the only effective and way to treat obesity, resulting in sustainable and significant weight loss in the long term (along with the resolution of metabolic comorbidities in up to 80% of cases), as well as in improving quality of life [O'Brien PE et al. 2019; Flum et al. 2007].

The effectiveness of bariatric surgery is often described by weight loss and/or with resolution of or improvement in obesity related diseases. Weight

loss is usually reported as the mean percentage of excess weight loss (%EWL), which is the standard in the bariatric surgery nomenclature. This calculation is derived from the formula: percentage of excess weight loss = (weight loss/excess weight) × 100, where excess weight = total preoperative weight – ideal weight. Changes in absolute weight (kilograms), BMI, and percentage of initial weight are also reported when appropriate.

In a meta-analysis of Buchwald et al., the effectiveness of bariatric surgery on weight loss and resolution of obesity related comorbidities was demonstrated in 22094 patients. The mean %EWL at the outcome time point for which the comorbidities were assessed was 47.5% (95% CI, 40.7%-54.2%) for gastric banding, 61.6% (56.7%-66.5%) for gastric bypass, 68.2% (61.5%-74.8%) for gastroplasty, and 70.1% (66.3%-73.9%) for biliopancreatic diversion or duodenal switch. Type 2 diabetes was completely resolved in 76.8% of patients and resolved or improved in 86.0%. Hyperlipidemia improved in 70% or more patients, hypertension was resolved in 61.7% of patients and resolved or improved in 78.5%. Obstructive sleep apnea was resolved in 85.7% of patients [Buchwald et al. 2004]. The results of a Swedish Obesity Subjects (SOS) study of Sjöström L., demonstrate good long term effects of weight loss surgery: mean changes in body weight after 2, 10, 15 and 20 years were -23%, -17%, -16% and -18% in the surgery group and 0%, 1%, -1% and -1% in the control group respectively. Compared with usual care, bariatric surgery was associated with long-term reduction in overall mortality and decreased incidences of diabetes, myocardial infarction, stroke and cancer [Sjöström, 2013]. In a systematic review (6 RCT) Colquitt et al. demonstrated that bariatric operations, regardless of the surgical intervention used, had better results in weight loss compared with no surgery at 1–10 years of follow-up (p<0.001). Resolution of comorbidities was also higher after surgery compared with conventional therapy [Colquitt et al., 2009]. In a systematic review O'Brien et al. published long-term outcomes of weight loss after 10 or more years following different bariatric operations. There were 18 reports on gastric bypass, 16 of which were for the RYGB variant and two were for the OAGB variant. All gastric bypasses combined showed a weighted mean % EWL of 56.7% at 10 or more years with a mean of 55.4% EWL for RYGB and 80.9% EWL for OAGB. The mean EWL for the 17 reports on laparoscopic adjustable gastric banding (LAGB) was 45.9%. For two RCTs with the use of LAGB, the mean weight loss was 55.9% EWL. Eleven reports on biliopancreatic diversion-duodenal switch (BPD ± DS) presented a mean of 74.1% EWL. In studies of BPD (N = 4), the weighted mean was 71.5% EWL, while for DS (N = 7), it was 75.2% EWL. Two reports on sleeve gastrectomy with a total of 79 patients presented a mean of 57.0% EWL [O'Brien et al. 2019].

### 2.5.3.6. Cost-effectiveness of surgical versus non-surgical treatment for obesity

The cost-effectiveness of bariatric surgery compared to non-surgical (NST) treatment has been discussed by many authors. Studies of Borisenko et al. demonstrated that among German and Danish bariatric patients, bariatric surgery, compared to NST, was cost-effective at 10 years and over the lifetime. At 10 years the incremental cost-effectiveness ratio (ICER) per quality-adjusted life years (QALYs) was EUR2457 in Germany and 17,818DKK in Denmark. Over the lifetime, surgery led to the saving EUR8522 and generated an increment of 0.7 years of life or 3.2 QALYs in Germany [Borisenko et al., 2017a,b]. The authors also showed that surgery may produce significant reduction in healthcare costs and a delay in surgery for up to 3 years may result in reduction in life years and QALYs gained [Borisenko et al., 2017a,b]. According to An et al., bariatric surgery was cost-effective (ICER of 674USD per QALY) compared to NST in Korean people with morbid obesity [An et al., 2020]. A study conducted by Gulliford et al, with obese 250, 000 patients demonstrated better outcomes of bariatric surgery compared to NST. In that study, bariatric surgery was associated with an increase in total life years of 6,097 per 1000 participants (accumulated over the lifetime of the participants), an increase in a number of life years, lived free from chronic comorbidities, of 10,297 per 1000 participants compared to NST. Although according to the study, bariatric surgery was not cost-saving, the increased healthcare costs were exceeded by the health benefits to obese individuals [Gulliford et al 2017].

## 2.6. Complications of bariatric surgery

### 2.6.1. Surgical complications

Bariatric surgery procedures are safe and carry a low risk of complications, especially in the setting of specialized centers. The average rate of complications in bariatric surgery is less than 10% [Sjöström et al., 2004] and overall mortality rate is <1% in most published series, with lower-risk patients showing a mortality rate of <0.35% (similar to that for cholecystectomy) [Dumon et al., 2011]. Buchwald and colleagues reported a 0.28% of early (less than 30 day) and 0.35% of late (30 days to 2 years) mortality rate after bariatric surgery in 85,048 patients [Buchwald et al., 2007]. The Swedish Obesity Study reported a mortality of 0.25% within 90 days of surgery [Sjöström et al., 2007].

Some complications can be procedure-specific. The most frequently reported perioperative complications associated with LRYGB are wound infection (2.98%), anastomotic leak (2.05%), gastrointestinal tract hemorrhage (1.93%), bowel obstruction (3.6%), and pulmonary embolus (0.41%), while late complications are stomal stenosis (4.73%), bowel obstruction (3.15%), incisional hernia 0.47%, marginal ulcer (0.3–16%) [Podnos et al., 2003; Farrell et al., 2009; Christou et al., 2004; Martin et al., 2011; Azagury DE., 2011; Sacks et al., 2011].

The most common complications associated with LSG include staple line leak (1.17%), post-operative hemorrhage (3.57%), and stenosis of the resected stomach (up to 4%) [Dumon et al., 2011; Parik et al., 2012].

### **2.6.2. Nutrient deficiencies after bariatric surgery and postoperative replacement prophylaxis**

Nutrient deficiencies are common after bariatric surgery. They are related to reduction in food Intake, as well as to the physiological impact of surgically induced anatomical changes in the gastrointestinal tract [Davies et al. 2007]. In a paper of Gehrler et al, the authors point out that prior to bariatric surgery, 57% of the patients had at least one deficiency [Gehrler et al., 2010]. In a review paper, Parrot and colleagues give an overview about the prevalence of (possible) micronutrient deficiencies before weight loss surgery (WLS) and the results are the following: thiamine deficiency as high as 29%; B12 deficiency 2–18%; folate deficiency as high as 54%; iron deficiency as high as 45%; vitamin D deficiency as high as 90%. The same paper describes also the prevalence of micronutrient deficiencies after WLS: B12 deficiency at 2–5years <20% for RYGB and 4–20% for SG; folate deficiency up to 65%; iron deficiency from 3 months to 10 years <18% for SG and 20–55% for RYGB; vitamin D deficiency up to 100% [Parrot et al., 2017]. In a (prospective) study with 99 bariatric patients, Kikkas et al., demonstrated that the risk for cumulative iron and vitamin B12 deficiency 5 years after LSG was 20% and 48%, respectively [Kikkas et al., 2019]. Recent guidelines recommend that all patients pursuing weight loss surgery should undergo a preoperative clinical nutrition evaluation by a registered dietician and all patients after bariatric surgery should use nutritional supplements for life [Mechanick et al., 2020; Parrott et al., 2017].

## **2.7. Anatomy and physiology of the stomach**

### **2.7.1. Anatomy of the stomach**

Topographically, the stomach can be divided into five regions: the cardia and gastroesophageal (GE) junction, the fundus, the corpus, the antrum, and the pylorus. The fundus and the corpus harbor acid-secreting glands, and the antrum harbors the alkaline secreting surface epithelium and the endocrine, gastrin-secreting G-cells [Soybel D., 2005].

### **2.7.2. Physiology of the stomach**

#### *Gastric mucosa*

Functionally, the gastric mucosa is divided into acid-secreting and nonacid secreting regions. The acid- and pepsinogen-secreting mucosa is found in the corpus and fundus. The acid secreting unit of the mucosa is the gastric gland. At the base of the gastric gland lie the pepsinogen-secreting chief cells. The middle of the gastric gland is largely populated with the HCl-secreting parietal cells.

Toward the lumen, at the neck, parietal cells are still present, but give way to mucus neck cells and then, near the opening, the mucosa is largely populated with surface epithelial cells. Intercalated between parietal cells and smaller immature cells are enterochromaffin-like (ECL) cells expressing histidine decarboxylase, an enzyme that is essential in the production of the paracrine agonist, histamine [Shubert, 2015; Soybel D., 2005].

#### *Neuroendocrine regulation of acid secretion*

Parietal cells are responsible for gastric acid secretion, which aids digestion of food, absorption of minerals and control of harmful bacteria. However, a fine balance of activators and inhibitors of parietal cell-mediated acid secretion is required to ensure proper digestion of food, while preventing damage to the gastric and duodenal mucosa. As a result, parietal cell secretion is highly regulated through numerous mechanisms including the vagus nerve, gastrin, histamine, ghrelin, somatostatin, glucagon-like peptide 1, and other agonists and antagonists. The tight regulation of parietal cells ensures the proper secretion of HCl. The  $H^+K^+ATPase$  enzyme expressed in parietal cells regulates the exchange of cytoplasmic  $H^+$  for extracellular  $K^+$ . The  $H^+$  secreted into the gastric lumen by the  $H^+K^+ATPase$  combines with luminal  $Cl^-$  to form gastric acid, HCl. Inhibition of the  $H^+K^+ATPase$  is the most efficacious method of preventing harmful gastric acid secretion. Proton pump inhibitors and potassium competitive acid blockers are widely used to therapeutically inhibit acid secretion. Stimulated delivery of the  $H^+K^+ATPase$  to the parietal cell apical surface requires the fusion of intracellular tubulovesicles with the overlying secretory canaliculus, a process that represents the most prominent example of apical membrane recycling. In addition to their unique ability to secrete gastric acid, parietal cells also play an important role in gastric mucosal homeostasis through the secretion of multiple growth factor molecules. The gastric parietal cell therefore plays multiple roles in gastric secretion and protection, as well as in coordination of physiological repair [Engevik et al., 2019; Ramsay et al., 2011].

#### *Alkaline secretion by the gastric mucosa*

The non-acid secreting mucosa of the gastric antrum and pylorus is characterized by the presence of relatively simple glands populated with the mucus- and  $HCO_3^-$  – secreting surface epithelium. The surface epithelium, both in the antrum and corpus/fundus regions, is the basis of the “mucosal barrier” [Soybel D., 2005].

#### *Gastric digestion and contributions to downstream absorption*

The stomach contributes to digestion of solid food by mixing chyme with acid and pepsin (pepsinogen autoactivated in the presence of luminal acid), which helps break down protein to simple peptides that will be absorbed or broken down further by intestinal peptidases. Subpopulations of parietal cells also secrete intrinsic factor, an essential cofactor in the absorption of vitamin B12 downstream in the terminal ileum. Gastric acid itself enables absorption of

specific metals and nonmetal cations, including  $\text{Ca}^{2+}$ ,  $\text{Fe}^{3+}$  and other trace metals. At low pH,  $\text{Ca}^{2+}$  is more fully released from binding bases, and is thus more available for absorption in the duodenum. Similarly,  $\text{Fe}^{2+}$  auto-oxidizes in the presence of luminal acid, placing it in a form more easily absorbed in the small intestine [Soybel D., 2005].

### *Gastric motility*

The stomach has three layers of muscularis: an inner circular layer, a middle longitudinal layer, and an outer but incomplete oblique layer. Motor functions in the stomach are segregated by the region. The fundus relaxes as fluids and solids enter the esophagus, a response known as receptive relaxation, and further as food actually enters the funds, a process known as adaptive relaxation. This response allows the liquid to pool in the fundic pouch while the solid components of the meal remain in the mainstream of flow toward the pylorus. On the greater curvature, in the muscularis of the upper corpus, lies the primary electrical pacemaker of the stomach. Superimposed on the basic electrical rhythm of the pacemaker, the corpus and antrum engage in a coordinated propulsion of the luminal contents toward the pylorus. The pylorus itself acts as a sieve, remaining open in anticipation of the wave of peristalsis. As the wave advances, small particles pass through the pyloric sphincter; when the wave hits, the pylorus closes, thereby acting as a barricade. The chyme, propelled with increasing velocity against the pyloric sphincter, is thus broken up by enzymatic digestion in combination with mechanical disruption [Soybel D., 2005].

### *Satiety*

The role of the stomach in regulating food intake has become an increasingly important issue, especially with the growing number of procedures for bariatric surgery. In this regard, the recently described hormone ghrelin has assumed central importance. Ghrelin is an appetite stimulating hormone that is released by the gastric mucosa into the portal circulation when the stomach is empty, passing into the central circulation to stimulate appetite centers in the hypothalamus; the circulating levels of ghrelin fall precipitously as soon as the stomach begins to fill. In bariatric surgical procedures with creation of small pouches that distend rapidly, the baseline and premeal peaks of ghrelin are suppressed, suggesting that blunting of ghrelin responses may contribute to suppression of appetite after bariatric surgery. Although ghrelin is by no means the dominating signal for control of satiety, its effects must be understood in the context of other neural and hormonal inputs to satiety centers in the pituitary [Soybel D., 2005].

## **2.8. Gastric biomarkers**

Stomach-specific biomarker testing (“serological biopsy”), being an alternative method to traditional EGDS, can provide a new non-invasive option to evaluate the stomach mucosa and gastric physiology in order to diagnose both functional

disorders and gastric diseases, including HP infection and AG [Agreus et al. 2012; Storskrubb et al. 2008; Syrjänen et al. 2019].

The latest innovation in this technology represents a panel of 4 stomach-specific biomarkers (Pgl, PglI, G17 and HPAB) known as GastroPanel® (GP) test (Biohit, Oyj, Finland), which distinguishes between 8 diagnostic biomarker profiles. Initially, GastroPanel had 5 diagnostic categories, but the new report includes 3 additional categories (**Figure 5**) [Agreus et al. 2012; Syrjänen et al. 2019].

**Pepsinogen I (Pgl)** is a precursor enzyme of pepsin, synthesized by the chief cells and the neck cells of the gastric corpus. As a pepsin precursor, the major part of Pgl is excreted into the gastric lumen, but a minor fraction is released into the blood circulation. Concentration of circulating Pgl is strongly correlated with quantity of chief cells in the corpus, and any loss of these cells due to mucosal atrophy results in a linear decrease in blood Pgl levels [Syrjänen et al., 2019].

**Pepsinogen II (PglI)** is produced by the chief cells and the mucous neck cells of the gastric corpus, in the pyloric glands of the antrum, as well as in Brunner's glands of the proximal duodenum. Elevated PglI level as a stand-alone marker reflects mucosal inflammation, the highest values usually being detected in HP-associated active gastritis [Syrjänen et al., 2019].

**Gastrin-17 (G17).** Gastrins are linear peptide hormones produced by the G-cells in the duodenum, in the pyloric part of the antrum, and in the pancreas. The main function of gastrins is to stimulate the secretion of gastric acid by the parietal cells in the corpus, as well as to increase the motility of the antrum. In addition, gastrins are known to stimulate gastric chief cells to secrete PGs, and also to induce the contraction of the lower esophageal sphincter. The dominating forms of gastrin in plasma/serum are amidated G-34 and amidated G-17, the latter, however, being specifically of antral G-cell origin. In the healthy antrum, G-17 is the most potent form of all gastrins. Plasma G-17 levels within the normal range indicate a normal structure and function of the antrum, whereas low or high values of G-17 reflect abnormalities in acid output by the corpus. Gastrin-17 concentration in blood during fasting (G17 basal (G17b)) falls as the acidity of the gastric contents rises (pH below 2.5) (less than 1 pmol/l indicates an increased secretion of hydrochloric acid). The gastrin-17 (basal) value also falls in the case of antrum mucosa atrophy, where G-cells disappear. Low gastrin-17 levels in blood during fasting may therefore indicate either an increased secretion of hydrochloric acid or antrum mucosal atrophy. Antrum mucosa atrophy, however, is dependent upon infection with HP. If gastrin-17 concentration is above 7 pmol/l, it is usually due to decreased secretion of hydrochloric acid. Distinguishing between antrum mucosa atrophy caused by HP infection and an increased secretion of hydrochloric acid (gastrin17 basal <1 pmol/l) can be performed by measuring the response of

gastrin-17 after protein stimulation (G17 stimulated (G17s)). If a person has been infected by HP and his/her gastrin-17 value is still low after protein stimulation (less than 3 pmol/l), it may indicate atrophic gastritis of the antrum. However, if HP antibody levels are not elevated, the results indicate increased secretion of hydrochloric acid [Syrjänen et al., 2019]. Gastrin-acid feedback mechanism is shown in **Figure 6**.

**Helicobacter pylori antibody (HPAB).** HP infection is the most important cause of chronic gastritis resulting in atrophic gastritis (AG) over time. In the stomach, HP is found within the mucous layer overlying the gastric epithelium, and within the mucosal glands, but it does not appear to invade the epithelial cells. The mucosa underneath and surrounding the areas of HP colonization is invariably inflamed. This condition is referred to as chronic superficial or non-atrophic gastritis [Syrjänen et al., 2019].

## 2.9. Interpretation of the biomarker profile

The manufacturer-validated reference values of the four biomarkers are the following: Pgl 30–160µg/l; PglII 3–15µg/l; Pgl/PglII ratio 3–20, G17b 1–7pmol/l; G17s 3–30 pmol/l; HPAb<30EIU [Storskrubb et al. 2008].

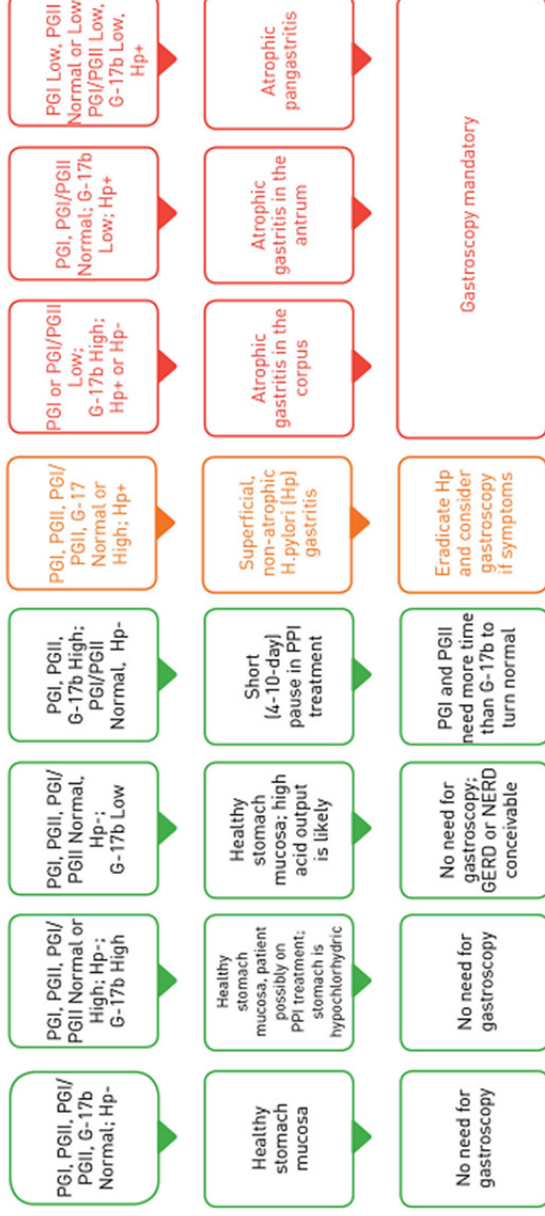
Based on the blood plasma levels of these biomarkers after fasting or after food (protein rich drink) stimulation, a GastroPanel® report is obtained from which valuable conclusions can be drawn about the structure and function of the gastric antral and corpus (oxyntic) mucosa, as well as about acid secretion or stomach acidity [Agreus et al. 2012].

The results are interpreted, using the special GastroSoft® software, by classifying them according to the biomarker levels, under one of the eight diagnostic categories (**Figure 5**):

- 1) normal profile
- 2) superficial (HP) gastritis – (Pgl, PglII, Pgl/PglII, G-17 normal or high; HPAb >30EIU)
- 3) atrophic gastritis of the antrum (AGA) – (Pgl, Pgl/PglII normal; G-17b low; HPAb>30EIU)
- 4) atrophic gastritis of the corpus (AGC) – (Pgl, Pgl/PglII low; G-17b high; HPAb <30EIU or >30EIU)
- 5) atrophic pan-gastritis (AG of the antrum and corpus) – (AGP) (Pgl low, PglII normal or low; Pgl/PglII low, G-17b low; HPAb >30EIU),
- 6) normal (healthy) stomach, high acid output – Pgl, PglII, Pgl/PglII, G17(s) normal; G17(b) low; HP-negative
- 7) normal (healthy) stomach, low acid output – Pgl, PglII normal or high; Pgl/PglII normal; G17 (b) high; HP-negative: eg. patients receiving PPI treatment
- 8) mucosal „state“ caused by a pause after continuous PPI treatment – Pgl, PglII, G17(b) high; Pgl/PglII norm; HP-negative. It is usually followed by „Rebound acid hypersecretion“ (G17b levels extremely low) [Agreus et al. 2012; Sipponen et al., 2007; Syrjänen et al., 2019].

# GastroPanel® – interpretation guide snapshot

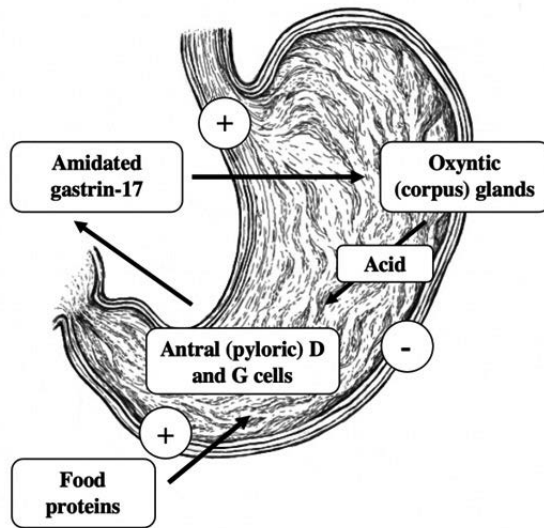
Structural and functional causes of dyspeptic symptoms diagnosed by GastroPanel test  
(PGI, PGII, PGI/PGII, G-17, Hp-Ab)



Normal structure   
  Referral to General Practitioner   
  Referral to Specialist; gastroscopy/biopsy

PGI – Pepsinogen I; PGII – Pepsinogen II; G-17b – basal (fasting) Gastrin 17; GERD – gastroesophageal reflux disease; NERD – Non-erosive reflux disease; PPI – proton pump inhibitor

**Figure 5.** Interpretation of GastroPanel biomarker test [Biohit Oyj., 2023]



+ – positive feedback; – – negative feedback

**Figure 6.** Mechanism of gastrin-acid feedback control (Figure by T. Veršinina)

### 3. SUMMARY OF THE LITERATURE REVIEW

Obesity, defined as abnormal or excessive fat accumulation, is a major health issue affecting about 650 million of the world's adult population. Obesity has been declared a chronic progressive disease that substantially increases the risk of diseases such as T2DM, fatty liver disease, hypertension, myocardial infarction, stroke, dementia, osteoarthritis, obstructive sleep apnoea. Obesity carries a major economic burden.

Obesity is the result of the interplay between heterogenic factors, deriving from a person's eating behaviour, physical activity and individual energy expenditure determinants. Multiple options have been described for treating overweight and obesity. However, since lifestyle interventions and medical therapy have limited effectiveness in reducing weight, then bariatric surgery has gained popularity, as it is the the only effective way to treat obesity in the long term. Nowadays, a variety of surgical options are currently offered for treatment of obesity, RYGB and SG being the two most common bariatric procedures performed in the world. The mechanisms of bariatric operations contributing to weight loss can vary in the different type procedures, being often combined. These mechanisms include effects on regulation of energy balance, both the central and peripheral nervous systems` regulation of appetite and metabolism, but also changes in gut hormones and microbioma.

As most bariatric procedures show similarly good results in excess weight loss, then multiple aspects (GERD, hiatal hernia, nutrient deficiencies) have to be considered when choosing the operation type. Bariatric surgery procedures are safe and carry a low risk of complications, especially when performed by experienced bariatric surgeons, in the setting of specialized centers.

Stomach-specific biomarker testing ("serological biopsy"), being an alternative method to traditional EGDS, can provide a new non-invasive option to evaluate the stomach mucosa and gastric physiology in order to diagnose both functional disorders and gastric diseases, including HP infection and AG. The latest innovation in this technology represents a panel of 4 stomach-specific biomarkers (Pgl, PglI, G-17 and HPAB) known as GastroPanel® test, which distinguishes between 8 diagnostic biomarker profiles. Based on the profiles, a report is obtained from which valuable conclusions can be drawn about the structure and function of the gastric antral and corpus (oxyntic) mucosa, as well as about acid secretion or stomach acidity.

## 4. AIMS OF THE STUDY

- 1) To find out if less invasive serological biomarker testing could avoid invasive preoperative endoscopies in obese patients referred to bariatric surgery.
- 2) To study the function of the gastric mucosa and acid secretion after bariatric surgery: changes in the gastric biomarker profile up to 2-year follow-up after bariatric surgery.
- 3) To evaluate whether postoperative gastric biomarker changes are related to operation and anatomical changes alone, or whether there could occur also some pathohistological alterations of the gastric mucosa; also how soon changes in biomarker levels develop postoperatively.
- 4) To evaluate whether preoperative, postoperative or preoperative/postoperative rate of stomach-specific biomarkers is correlated with excess weight loss at 1-year follow-up and whether a relevant test could be used as an additional tool to select the most appropriate operation type (LSG, LRYGB) for each bariatric patient.

## 5. MATERIAL AND METHODS

### 5.1. Patients

The whole study is based on the data of 105 patients (33 male, 72 female) who underwent bariatric operation at the Surgery Clinic of Tartu University Hospital between 2008 and 2016 (**Table 1**). Some patients were involved in several studies. Approval was obtained from the Research Ethics Committee of the University of Tartu, license no. 218/T-6.

**Table 1. Characteristics of publications**

| Study type and period of data collection | No of patients (male/female) | Goals of the study   | Journal (No)                                 |
|--|------------------------------|--|--|
| Prospective 2008–2011                    | 40<br>12/28                  | Gastric biomarker profile after bariatric surgery                              | Hepatogastroenterology (I)                   |
| Prospective 2013–2015                    | 65<br>21/44                  | Preoperative correlation of gastric biomarkers with gastrobiopsies             | BMC Obesity (II)                             |
| Prospective 2013–2016                    | 57<br>16/41                  | Dynamics of gastric biomarkers during 1-year follow-up after bariatric surgery | Surgery, Gastroenterology and Oncology (III) |

### 5.2. Inclusion and exclusion criteria

The eligible patients were adults aged 18 years and older with a BMI of >40 or >35, with certain obesity-related comorbidities (hypertension, diabetes, dyslipidaemia, sleep apnea, osteoarthritis, polycystic ovarian syndrome), providing informed written consent for participation in the study. Patients who were younger than 18 years or did not meet the criteria for bariatric operation or who were unwilling to participate in the study were excluded.

Patients who participated in the study were interviewed, tested by GP, and subjected to esophagogastroduodenoscopy examination with directed biopsies.

### 5.3. Patient questionnaire

In all patients, eligible for bariatric operation and willing to participate in the study, medical history (co-morbidities, medications, prior operations, allergy) and physical parameters (height, weight, BMI, waist circumference) were recorded. Also the patients were interviewed using a standardized questionnaire about the specific history of upper abdominal complaints (heartburn, reflux,

dyspepsia). The interview was conducted prior to biomarker test, EGDS and surgery (Publication II), during the follow-up visit at 1, 3, 12 months (Publication III) and 2 years (Publication I) after bariatric surgery.

#### **5.4. Biomarker test examination**

Blood samples were collected for GP testing preoperatively and were repeated at postoperative days 1 and 3, and subsequently at 1-, 3, 12 months and 2 years postoperatively (in different study groups). GP biomarker test was conducted according to the Biohit standard rules: the patient had to be starved for at least 10 hours before the study (no food, no liquids), smoking was prohibited 4–10 hours before the study. The patient was not allowed to use antacides, PPI-s and histamine blockers for at least 1 week.

Blood was collected and processed according to the instructions given by the manufacturer of GP biomarker test (GastroPanel; Biohit, Plc, Helsinki, Finland): 10 mL of fasting blood were collected, plasma was separated, a specific gastrin stabilizer (Biohit Plc, Helsinki, Finland) was added, and the sample was stored at -70°C until analysis. After drawing the basal (fasting) sample, the patient drank a protein rich drink; after 20 minutes a second blood sample (stimulation sample) was collected (10 mL) and the above described procedure for sample preservation was carried out. All samples were properly stored and transported to the service laboratory of Biohit Oyj for analysis. The plasma levels of the following biomarkers were measured with the ELISA assay: fasting (basal) Pgl and PglI, G17b and G17s and HPAB. All assays were carried out in duplicate at the service laboratory of Biohit Plc (Helsinki, Finland) [Syrjänen et al., 2019; Sillakivi et al., 2013].

#### **5.5. Interpretation of GastroPanel**

GP is an automated ELISA test that measures the plasma levels of the following biomarkers: Pgl and PglI, G17b and G17s, HPAB. The manufacturer-validated reference values of the four biomarkers were used: Pgl 30–160µg/l; PglI 3–15µg/l; Pgl/PglI ratio 3–20, G17b 1–7pmol/l; G17s 3–30 pmol/l; HPAB<30EIU [Storskrubb et al. 2008; Dixon et al., 1996].

The results are interpreted, using the special GastroSoft® software, by classifying the results, according to the biomarker levels, under one of the five diagnostic categories:

- 1) normal profile
- 2) superficial (HP) gastritis – (Pgl, PglI, Pgl/PglI, G-17 Normal or High; HPAB >30EIU)
- 3) atrophic gastritis of the antrum (AGA) – (Pgl, Pgl/PglI Normal; G-17b Low; HPAB>30EIU)
- 4) atrophic gastritis of the corpus (AGC) – (Pgl, Pgl/PglI Low; G-17b High; HPAB <30EIU or >30EIU)

5) atrophic pan-gastritis (AG of the antrum and corpus) (AGP) – (Pgl Low, PglI Normal or Low; PglI/PglI Low, G-17b Low; HPAB >30EIU), as detailed elsewhere [Agreus et al., 2012; Sipponen et al., 2007].

At the time we conducted our study, GP had 5 different marker profiles, while the new (updated) version includes 8 profiles [Syrjänen et al., 2019].

## 5.6. Esophagogastroduodenoscopy

Routine EGDS examination with directed biopsies was performed preoperatively (Publications II and III) and 12 months after bariatric operation (in SG patients) (Publication III)

All endoscopic evaluations were carried out using pharyngeal anesthesia with 4% Lidocaine gel on the scope. Biopsy specimens were obtained using standard biopsy forceps.

During EGDS, the esophageal, gastric and duodenal mucosa was visually inspected and abnormalities were detected. The degree of esophagitis was evaluated according to the Los Angeles classification (LA) [Armstrong et al. 1996]. In every patient, two biopsies from the antrum (2cm from the pyloric ring) and two biopsies from the corpus were collected. Additional biopsies were taken only when necessary.

All endoscopies were performed at the Surgery Clinic of Tartu University Hospital, Department of Abdominal Surgery by, two endoscopists (JS, TS).

## 5.7. Histology and *H. pylori*

The histology of the biopsies was evaluated separately for the gastric antrum and corpus, according to the updated Sidney System (USS) classification [Dixon et al. 1996], by 3 independent pathologists (ZR, KS, PS). In the case of discrepant results, the biopsies were re-evaluated by a pathologists' panel, and the consensus diagnosis was used as the final one. The biopsy specimens were fixed overnight in neutral buffered formalin and stained for morphological and HP examinations using the hematoxylin-eosin method and the Giemsa method. At histopathological evaluation, chronic gastritis, inflammation activity, mucosal atrophy and intestinal metaplasia were classified as follows: none, mild, moderate, severe. HP-colonization and its abundance were semi-quantitatively estimated, separately in the antral and corpus mucosa, by microscopic counting, as absent, mild, moderate and severe, as described earlier [Stolte et al. 1989; Peetsalu et al., 2005].

## 5.8. Surgical procedures

Laparoscopic Sleeve Gastrectomy and Laparoscopic Roux-en-Y gastric bypass operations (according to the standardised laparoscopic technique) were used for the study patients (**Figure 4**).

The SG provides reduction in the gastric volume through vertical stapler resection along the greater gastric curvature. A major part of the corpus and a small part of the antrum were resected, leaving a remnant (tubular gastric „sleeve“) of 10–15% of the initial stomach comprising a certain amount of both the antral and corpus mucosa. In the case of SG, a bougie size of 35 Fr was used and resection of the stomach was started at 4 cm from the pylorus, the volume of the created sleeve was about 100–120ml. The weight of the resected stomach was measured using an electronic scale and the volume of the resected stomach was measured using a tap water test (a small hole was made in the corner of the specimen and was filled with tap water until the water poured out of the created defect) as described earlier [Weiner et al., 2007].

RYGB is a combined gold standard method where, by horizontal division separating the upper part of the stomach with the use of staplers, a new, smaller stomach is formed, while more than 90% of it is bypassed and excluded from the food passage. In RYGB, the volume of the created pouch is about 30–50 ml, the length of the biliopancreatic limb is 50–60 cm and the length of the alimentary limb is 120 cm. Food is directed from this small pouch into the middle part of the small intestine through a gastro-jejunal anastomosis. No organs are removed, nor are any resections carried out.

## 5.9. Statistics

The data were analysed using Statistica® version 10.0 and the 13 software package. Continuous variables were presented as mean (M) values with standard deviation (SD), and nominal variables were presented as relative frequencies. Comparison between the groups was performed using the two-sample *t*-test or the Mann-Whitney *U* test for continuous variables, where appropriate, and Fisher's exact test for nominal variables. All statistical tests were two-sided and  $p \leq 0.05$  was considered significant.

## 5.10. Study groups

### Publication II

In a prospective study, conducted between January 2013 and September 2015, 65 patients referred to bariatric surgery at the Surgery Clinic of Tartu University Hospital, were included and data was collected according to a standardised protocol (Methods).

According to the Gastropanel results, all patients were divided into 2 groups: HS (Healthy stomach) group and NHS (Non-healthy stomach) group. In the HS stomach group all biomarkers were normal. In the NHS group one or more biomarker levels were abnormal, describing the following pathological conditions: 1) Non-atrophic *H. Pylori* (HP+) gastritis – only *H. Pylori* antibody test was positive; 2) atrophic antrum gastritis – Pgl, Pgl/PgII normal; G17(b) low; HP+; 3) atrophic corpus gastritis – Pgl and/or Pgl/PgII low; G17(b) high; HP+ or HP-; 4) atrophic pangastritis (antrum and corpus) – Pgl low, PgII normal or

low; PgI/PgII low; G17(b) low; HP+. Gastropanel test results in the HS group and the NHS group were evaluated against the gold standard, EGDS with biopsies.

The aim of the study was to find out if the data collected from endoscopic findings and gastrobiopsies correlates with the results of GP. Another aim was to ascertain if GP test as a less invasive method can be used instead of EGDS in selected patients undergoing elective bariatric surgery.

### **Publication III**

This prospective study of 57 bariatric patients is a continuation of a study conducted between 2013 and 2015 at Tartu University Hospital. Altogether 29 LSG and 28 LRYGB patients, matched according to preoperative BMI, sex and age, were included in the study population. The data was collected according to a standardised protocol pre- and postoperatively (Methods). The study describes changes in the biomarker profile in different operation groups (LRYGB and LSG) during 1- year follow-up after bariatric surgery and compares the biomarker profile with the data of histopathological evaluation of the samples collected on EGDS preoperatively and 12 months postoperatively (only the LSG group).

The main aim of the study was to evaluate whether postoperative changes in gastric biomarkers are related to operation and anatomical changes alone, or whether there could occur also some patohistological alterations of the gastric mucosa; also how soon the changes in biomarker levels develop postoperatively. Another aim was to to evaluate whether preoperative, postoperative or preoperative/postoperative rate of stomach-specific biomarkers is correlated with excess weight loss at 1-year follow-up.

### **Publication I**

Publication I is based on the investigation of the biomarker profile of 40 patients 2 years (average 22 months) after bariatric surgery (LSG and LRYGB operations). There were 12 male and 28 female patients in the study group who had undergone either SG or RYGB operation. Twenty SG and 20 RYGB patients, previously matched according to preoperative BMI, postoperative time, sex and age, were selected and included in the study.

All patients were interviewed using a standardised questionnaire and blood samples were collected for biomarker testing (Methods).

The aim of the study was to compare the stomach specific biomarker profile between the SG and RYGB groups.

## 6. RESULTS

### 6.1. Correlation between gastric biomarkers and gastrobiopsies (Publication II)

The key characteristics of the study patients are presented in **Table 2**.

**Table 2.** Key characteristics of the patients (Publication II)

| <b>Patient characteristics</b>                            |              |
|---|--------------|
| Mean age, years (SD)                                      | 43.1 (9.08)  |
| Female patients, n (%)                                    | 44 (67.7)    |
| Mean preoperative weight, kg (SD)                         | 128.3 (21.5) |
| Mean preoperative body mass index, kg/m <sup>2</sup> (SD) | 44.3 (5.12)  |
| Concomitant diseases, n (%)                               | 57/65 (87.7) |
| Type II diabetes, n (%)                                   | 9 (13.8)     |
| Hypertension, other cardiac diseases, n (%)               | 39 (60)      |
| Obstructive sleep apnea, n (%)                            | 20 (30.7)    |
| Degenerative joint disease, n (%)                         | 13 (20)      |
| Hypercholesterolemia, n (%)                               | 42 (64.6)    |
| Smokers, n (%)  | 17 (26.1)    |

SD – standard deviation

According to the Gastropanel test results, 22 (34%) patients were classified into the HS group and 43 (66%) into the NHS group. There was a statistically significant difference in the mean Pgl, PglII, Pgl/PglII and IgG values between the HS and NHS groups ( $p < 0.001$ ) (**Table 3**).

**Table 3.** Biomarker levels for the two categories (HS/NHS) of patients (Publication II)

| <b>Biomarkers</b>                            | <b>Whole Series</b>             | <b>HS Group<br/>n=22</b>        | <b>NHS Group<br/>n=43</b>       | <b>p-value*</b>   |
|--|---------------------------------|---------------------------------|---------------------------------|-------------------|
| Mean G17b $\pm$ SD, pmol/l (ranges)*         | 3.9 $\pm$ 5.0<br>(0–28.1)       | 2.05 $\pm$ 2.2<br>(0–9.9)       | 4.9 $\pm$ 5.7<br>(0–28.1)       | <b>0.028</b>      |
| Mean G17s $\pm$ SD, pmol/l (ranges)**        | 12.3 $\pm$ 10.3<br>(0–40.9)     | 8.8 $\pm$ 8.4<br>(0.2–36.1)     | 14.1 $\pm$ 10.8<br>(0–40.9)     | <b>0.048</b>      |
| Mean PG I $\pm$ SD, $\mu$ g/l (ranges)       | 81.8 $\pm$ 38.6<br>(22.7–197.5) | 58.2 $\pm$ 27.9<br>(33.0–151.1) | 93.9 $\pm$ 38.0<br>(22.7–197.5) | <b>&lt;0.0001</b> |
| Mean PG II $\pm$ SD, $\mu$ g/l (ranges)      | 7.2 $\pm$ 5.9<br>(0.3–29.3)     | 3.37 $\pm$ 1.7<br>(0.3–8.4)     | 9.2 $\pm$ 6.4<br>(1.4–29.3)     | <b>&lt;0.0001</b> |
| Mean PG I/PG II $\pm$ SD, $\mu$ g/l (ranges) | 15.5 $\pm$ 12.3<br>(3.3–103.3)  | 21.6 $\pm$ 18.9<br>(10.7–103.3) | 12.4 $\pm$ 4.6<br>(3.2–25.2)    | <b>&lt;0.0001</b> |
| Mean IgG $\pm$ SD, EIU (ranges)              | 64.2 $\pm$ 40.8<br>(2.1–121.1)  | 13.12 $\pm$ 7.7<br>(2.1–27.3)   | 90.4 $\pm$ 21.1<br>(38.1–121.1) | <b>&lt;0.0001</b> |

HS – Healthy stomach; NHS – Non-healthy stomach; G17b – basal gastrin-17; G17s – stimulated gastrin-17; \*Mann-Whitney U-Test

The concordance between GastroPanel test and USS histology (5 categories in both) is shown in **Table 4**. Agreement between the two methods was significant: Kappa=0.68 (95% CI 0.504–0.854), with an overall agreement of 55/65 (84.6%; 95% CI 73.9–91.4%) across the five diagnostic categories.

The prevalence of HP infection in the entire cohort was 43/65 (66%) in GP test and 41 (63%) in the histological evaluation of the biopsies. In the HS group 6/22 (27%) of patients had mild chronic HP-negative gastritis of the antrum and corpus without a significant difference in their biomarker level compared with the other patients in that group.

In the NHS group, of all patients with histologically confirmed chronic gastritis, 4 had moderate degree AGA. The biomarker test also confirmed those 2 AGA cases (G17b and G17s were low in the first case and G17s response was below a threshold of 3 pmol). One patient, who was classified as a case of panatrophy by GastroPanel, had moderate degree chronic gastritis of the antrum and corpus with mild degree HP infection, but no atrophy.

Four patients in the HS group and 10 in the NHS group had endoscopically confirmed esophagitis, while 2 and 8 of them respectively, had also reflux complaints (**Table 5**).

According to the results of our study, preoperative EGDS could have been avoided in 31% of the patients (20/65) in the study group.

We did not find any correlation between G17b and esophagitis at EGDS, or between G17b levels and GERD complaints.

**Table 4.** Concordance between biomarker test results and histological diagnosis (Publication II)

| Diagnostic categories of the GastroPanel test | Gastric mucosa histology (USS classification*) |                            |                      |                      |                  | Total     |
|---|--|----------------------------|----------------------|----------------------|------------------|-----------|
|   | Normal   | Superficial (HP) Gastritis | Antral atrophy (AGA) | Corpus atrophy (AGC) | Panatrophy (AGP) |           |
| <b>Normal profile</b>                         | 16   | 6                          | 0                    | 0                    | 0                | <b>22</b> |
| <b>Superficial (HP) Gastritis</b>             | 0  | 38                         | 3                    | 0                    | 0                | <b>41</b> |
| <b>Antral atrophy</b>                         | 0  | 0                          | 1                    | 0                    | 0                | <b>1</b>  |
| <b>Corpus atrophy</b>                         | 0  | 0                          | 0                    | 0                    | 0                | <b>0</b>  |
| <b>Panatrophy</b>                             | 0  | 1                          | 0                    | 0                    | 0                | <b>1</b>  |
| <b>Total</b>                                  | <b>16</b>                                      | <b>45</b>                  | <b>4</b>             | <b>0</b>             | <b>0</b>         | <b>65</b> |

\*The Updated Sydney System (USS) classification of gastritis

**Table 5.** Endoscopic findings and clinical symptoms in pre-operative assessment of the HS and NHS groups (Publication II)

| Endoscopic findings        | Healthy stomach<br>(HS Group)<br>n=22 | Non-healthy stomach<br>(NHS Group)<br>n=43 |
|----------------------------|---------------------------------------|--|
| Hiatal hernia              | 4                                     | 15   |
| Esophagitis                | 4                                     | 10   |
| Erosions/erosive gastritis | 3                                     | 14   |
| Duodenal polyp             | -                                     | 1  |
| GERD*                      | 2                                     | 8  |

GERD – gastroesophageal reflux disease; \*clinical symptom

## **6.2. Changes in the biomarker profile in different operation groups (LRYGB and LSG) during 2- year follow-up after bariatric surgery (Publications III, I)**

The key clinical characteristics of the study groups are presented in **Table 6** (Publication III) and **Table 7** (Publication I).

**Tables 8 and 9** show the gastric biomarker levels (in dynamics) of LSG and LRYGB patients from 2 different studies (Publications III and I).

In the LSG group PgI level declined rapidly after surgery and at 12 months follow-up mean PgI level was significantly lower (20.5 µg/L), compared to its preoperative level (75 µg/L), ( $p < 0.0001$ ) (Publication III). Data from Publication I showed that 2 years after surgery, PgI values were almost identical to those described at 1-year follow-up (Publication I). Abnormally low PgI level ( $< 30$  µg/L) in the LSG group was seen in 82% of the cases at 12-month follow-up (Publication III) and in 80% of cases 2 years after surgery (Publication I). PgI values in the LRYGB group increased from 1POP day to 3POP day and started to decrease at 1-month follow-up, being significantly lower (32.3 µg/L) at 12-month follow-up compared to their preoperative values ( $p < 0.0001$ ) (**Table 9**, Publication III). Two years after LRYGB operation, the mean value of PgI in the study group was 32.2 µg/L (**Table 9**, Publication I). PgII level and the PgI/PgII ratio declined after surgery, being significantly lower at 12 months compared to their preoperative values (PgII  $p < 0.0006$ ; PgI/PgII  $p < 0.0001$ ) (**Table 9**, Publication III). Low levels of PgII and a low PgI/PgII ratio was seen 2 years after surgery in the LSG group (**Table 8**, Publication I).

In the LRYGB groups, PgI and PgII levels rose markedly during the first 3 postoperative days, thereafter their levels and the PgI/PgII ratio declined significantly (**Table 9**, Publication III).

**Table 8** presents a 83% increase in the mean G17f values to 7.9pmol/L in the LSG group by the end of one POP year (Publication III). The results from Publication I showed that in the LSG group mean G17f level was high 2 years after surgery (13.9 pmol/L), and the proportion of cases with G17f exceeding the upper normal cut-off limit was 40% (**Table 8**, Publication I). In the LRYGB

group, the mean values of G17f increased slightly immediately after operation and declined to relatively low levels, i.e., 2.5pmol/L at year 1 (**Table 9**, Publication III). Two years after LRYGB operation, the mean values of G17f in the study group was 3.1pmol/L (**Table 9**, Publication I).

In the LSG groups, G17s was normal (response >3 pmol/l) in 40% of the cases at year 1 (**Table 8**, Publication III) and in 100% of the cases at 2 years after the operation (**Table 8**, Publication I).

In the LRYGB groups, 7.4% of the cases showed a normal stimulation response of G17 at one-year follow-up and 25% of the cases 2 years after the operation (**Table 9**, Publication III and Publication I).

Preoperative histopathological findings confirmed chronic gastritis in 79.3% of the LSG patients and in 67.9% of the LRYGP patients (**Table 10**, Publication III). The prevalence of HP colonization was high in both groups, at 62.1% and 61%, respectively. Postoperative histopathological findings revealed no substantial changes in chronic gastritis scores for LSG patients (in one case moderate degree antral mucosa atrophy had resolved). HP colonization reduced from 62% to 44% (**Table 10**, Publication III).

According to the Spearman test, the preoperative- and one-year follow-up levels of the gastric biomarkers (Pgl, PglI G17f and G17s, HPAB) and the preoperative/one-year ratio did not correlate with %EWL at 1 year either in the LSG or the LRYGB group (all p=NS).

**Table 6.** Key characteristics of the LSG and LRYGP patients (Publication III)

|  | LSG group<br>n=29 | LRYGP group<br>n=28 | p-value |
|--|-------------------|---------------------|---------|
| <b>Preoperatively</b>                            |                   |                     |         |
| Mean age, years (SD)                             | 42±10.7           | 43±7.9              | 0.776   |
| Females  | 21                | 20                  | 0.999   |
| Mean weight, kg (SD)                             | 127±24.0          | 128±18.4            | 0.909   |
| Mean body mass index, kg/m <sup>2</sup> (SD)     | 44±5.6            | 44±4.9              | 0.959   |
| Mean weight of removed gastric specimen, g (SD)  | 159.1±37.1        | -                   |         |
| Mean volume of removed gastric specimen, ml (SD) | 692.2±167.2       | -                   |         |
| <b>Postoperatively (1-year follow-up)</b>        |                   |                     |         |
| Mean weight, kg (SD)                             | 86.8±21.8         | 92.7±15.9           | 0.253   |
| Mean body mass index, kg/m <sup>2</sup> (SD)     | 30.0±5.7          | 31.6±4.6            | 0.238   |
| Mean excess weight loss % (SD)                   | 77.3±25.4         | 65.1±21.3           | 0.054   |

*LSG* – laparoscopic sleeve gastrectomy; *LRYGP* – laparoscopic Roux-en-Y gastric bypass; *SD* – standard deviation

**Table 7.** Demographic data of the LRYGB and LSG patient groups (Publication I)

| <b>Time period</b> | <b>Patient data</b>                  | <b>LRYGB<br/>n=20</b> | <b>LSG<br/>n=20</b> | <b>p-value</b> |
|--------------------|--------------------------------------|-----------------------|---------------------|----------------|
| Preoperative       | Male/female ratio                    | 6/14                  | 6/14                | NS             |
|                    | Mean age, years                      | 44.1                  | 47.2                | NS             |
|                    | Mean BMI, kg/m <sup>2</sup>          | 49.0                  | 47.8                | NS             |
| Postoperative      | Follow-up (months<br>from operation) | 21.6                  | 22.7                | NS             |
|                    | Mean BMI, kg/m <sup>2</sup>          | 33.45                 | 32.2                | NS             |
|                    | Mean excess weight<br>loss %         | 62.9                  | 67.2                | NS             |

*LSG* – laparoscopic sleeve gastrectomy; *LRYGP* – laparoscopic Roux-en-Y gastric bypass

**Table 8.** Gastric biomarkers (in dynamics) for the LSG patients (Publications III and I)

| Biomarkers  | Publication III         |                          |                          |                        |                         |                         | Publication I                                     |  |                         |
|---|-------------------------|--------------------------|--------------------------|------------------------|-------------------------|-------------------------|---|--|-------------------------|
|   | Preoperative            | 1 day POP                | 3 day POP                | 1 month POP            | 3 months POP            | 12 months POP           | Change of mean values, 1 year vs preoperative (%) | P value (mean preoperative vs 1year POP) | 22 months POP           |
| Mean PG I±SD (range)  | 75±30.1<br>(34.4–154.6) | 32.1±21.6<br>(5.3–109.9) | 16.6±11.3<br>(2.8–47.6)  | 13.2±7.5<br>(2–39.4)   | 18.3±13.9<br>(0.8–53.3) | 20.5±9.6<br>(7.9–46.6)  | -73%  | <0.0001                                  | 20.0±11.5<br>(4.3–43.8) |
| PG I is abnormally low (<30 µg/L) (percentage)              | 0%                      | 54%                      | 82%                      | 96%                    | 82%                     | 82%                     |   | <0.0001                                  | 80%                     |
| Mean PG II±SD (range)                                       | 6.1±4.1<br>(1.1–16)     | 3.8±2.5<br>(1.1–12.9)    | 1.5±1.0<br>(0–3.9)       | 2.1±1.5<br>(0.4–6.8)   | 2.9±2.4<br>(0.6–10.7)   | 4.3±2.9<br>(1.3–12.8)   | -30%  | 0.0006                                   | 4.3±1.8<br>(1.9–8.2)    |
| PG II is abnormally low (<3 µg/L) (percentage)              | 10%                     | 46%                      | 89%                      | 79%                    | 64%                     | 39%                     |   |  | 30%                     |
| Mean PGI/PGII±SD (range)                                    | 14.7±5.6<br>(3.2–31.1)  | 8.9±3.6<br>(2.1–17)      | 13.2±9.6<br>(3.2–51.5)   | 8.1±5.5<br>(2.5–28.2)  | 7.3±3.7<br>(1.3–16.1)   | 5.5±2.3<br>(1.7–11.8)   | -63%  | <0.0001                                  | 4.8±2.3<br>(1.6–9.9)    |
| PGI/PGII is abnormally low (<3) (percentage)                | 3.6%                    | 3.6%                     | 0%                       | 10.7%                  | 3.6%                    | 7.1%                    |   |  | 25%                     |
| Mean G17f±SD (range) pmol/L                                 | 4.3±6.4<br>(0.1–28.1)   | 11.8±9.5                 | 10.1±11.8<br>(0.01–42.3) | 6.5±7.5<br>(0.01–26.3) | 7.5±9.1<br>(0–42)       | 7.9±11.2<br>(0.1–46.5)  | +83%  | 0.0356                                   | 13.9±17.2<br>(2–58.7)   |
| G17f is abnormally high (>7pmol/L) (percentage)             | 13.8%                   | 60.7%                    | 42.9%                    | 32.1%                  | 35.7%                   | 32%                     |   |  | 40%                     |
| Mean G17s±SD (range) pmol/L                                 | 9.9±9.1<br>(0–40)       | 13.2±12.0<br>(0.3–49.9)  | 13.2±12.0<br>(0.3–49.9)  | 13.1±9.1<br>(0–41.3)   | 12.7±8.2<br>(0–30.6)    | 14.6±12.8<br>(0.1–45.8) | +48%  | 0.0109                                   | 21.3±16.6<br>(3.8–60.8) |
| Stimulation response of G17 normal (>3 pmol/L) (percentage) | 58.6%                   | 14.3%                    | 14.3%                    | 39.3%                  | 33.3                    | 40%                     |   |  | 100%                    |
| <i>Helicobacter pylori</i> positive (percentage)            | 65.5%                   |                          |                          |                        |                         | 53.6%                   |   | 0.364                                    | 45%                     |

LSG – laparoscopic sleeve gastrectomy; POP – postoperatively; PGI – Pepsinogen I; SD – standard deviation; PG II – Pepsinogen II; G17f – fasting (basal) Gastrin 17; G17s – stimulated amidated Gastrin 17

**Table 9.** Gastric biomarkers (in dynamics) for the LRYGP patients (Publications III and I)

| Publication III   |                           |                            |                          | Publication I           |                           |                       |   |   |                                  |
|---|---------------------------|----------------------------|--------------------------|-------------------------|---------------------------|-----------------------|---|---|----------------------------------|
| Biomarkers  | Preoperative              | 1 day POP                  | 3 day POP                | 1 month POP             | 3 months POP              | 12 months POP         | Change of mean values, 1 year vs preoperative (%) | P value (mean preoperative vs 1 year POP) | 22 months POP                    |
| Mean PG I±SD (range)<br>µg/L                                | 84.3±44.0<br>(22.7–186.8) | 169.8±68.4<br>(71.2–356.4) | 200.5±61.7<br>(70.7–400) | 33.2±21.5<br>(1.8–91.6) | 33.9±25.6<br>(10.8–125.3) | 32.3±20.4<br>(11–88)  | -62%  | <0.0001                                   | 32.2 ±12.2<br>(13.5–55.0)<br>40% |
| PG I is abnormally low (<30 µg/L) (percentage)              | 3.6%                      | 0%                         | 0%                       | 58.3%                   | 65.2%                     | 59.3%                 |   |   |                                  |
| Mean PG II±SD (range)<br>µg/L                               | 7.9±6.9<br>(0.3–29.3)     | 17.3±11.7<br>(3–50)        | 29.7±22.4<br>(1.7–73.3)  | 3.5±3.3<br>(0.4–16.2)   | 4.4±4.2<br>(1.1–20.6)     | 6.8±5.2<br>(1.1–20.9) | -14%  | 0.5219                                    | 5.5 ±2.3<br>(1.6–11.2)<br>10%    |
| PG II is abnormally low (<3 µg/L) (percentage)              | 25%                       | 0%                         | 8%                       | 66.7%                   | 60.9%                     | 18.5%                 |   |   |                                  |
| Mean PGI/PGII±SD (range)                                    | 16.9±17.7<br>(3.9–103.4)  | 12.8±6.6<br>(4–34)         | 12.5±12.1<br>(2.7–61.8)  | 12.7±9.8<br>(0.5–49.7)  | 9.9±5.4<br>(2.6–22.7)     | 5.0±3.5<br>(1.4–13.8) | -70%  |   | 6.6 ±2.9<br>(1.7–11.7)<br>10%    |
| PGI/PGII is abnormally low (<3) (percentage)                | 0%                        | 0%                         | 12%                      | 4.2%                    | 4.4%                      | 18.5%                 |   | 0.0016                                    |                                  |
| Mean G17±SD (range)<br>pmol/L                               | 3.55±3.38<br>(0–12.6)     | 4.11±4.83<br>(0–23.5)      | 4.49±5.48<br>(0–19.8)    | 4.19±9.66<br>(0–46.6)   | 2.68±3.60<br>(0–11.1)     | 2.56±3.67<br>(0–12.3) | -28%  | 0.294                                     | 3.1 ±4.3<br>(0–15.0)<br>15%      |
| G17f is abnormally high (>7pmol/L) (percentage)             | 14.3%                     | 24%                        | 20%                      | 12.5%                   | 17.4%                     | 11.1%                 |   |   |                                  |
| Mean G17s±SD (range)<br>pmol/L                              | 14.4±10.9<br>(1.5–40)     | 14.4±10.9<br>(1.5–40)      | 4.7±5.6<br>(0.2–19.2)    | 5.1±11.5<br>(0–54.3)    | 2.4±3.7<br>(0–14.2)       | 2.9±4.6<br>(0–20.7)   | -80%  | <0.0001                                   | 2.6 ±3.8<br>(0–14.2)<br>25%      |
| Stimulation response of G17 normal (>3 pmol/L) (percentage) | 75%                       | 75%                        | 16.7%                    | 0%                      | 0%                        | 7.4%                  |   |   |                                  |
| <i>Helicobacter pylori</i> positive (percentage)            | 64.3%                     |                            |                          |                         |                           | 48.2%                 |   | 0.233                                     | 50%                              |

LRYGP – laparoscopic Roux-en-Y gastric bypass; POP – postoperatively; PGI – Pepsinogen I; SD – standard deviation; PG II – Pepsinogen II; G17f – fasting (basal) Gastrin 17; G17s – stimulated amidated Gastrin 17

**Table 10.** Stomach histology of the LSG and LRYGP patients (Publication III)

| Histological findings                       | Preoperatively | Postoperatively | Preoperatively  |
|---|----------------|-----------------|-----------------|
|   | LSG<br>n=29    | LSG*<br>n=25    | LRYGP**<br>n=28 |
| Normal, n (%)                               | 6 (20.7)       | 2 (8)           | 9 (32.1)        |
| Chronic gastritis                           | 23 (79.3)      | 23 (92.0)       | 19 (67.9)       |
| - with moderate or severe<br>corpus atrophy | 0 (0)          | 0 (0)           | 0 (0)           |
| - with moderate or severe<br>antrum atrophy | 1 (4.3)        | 0 (0)           | 3 (15.8)        |
| <i>Helicobacter pylori</i> positivity       | 18 (62.1)      | 11 (44)         | 17 (60.7)       |

\* postoperatively 4 patients in the LSG group refused to undergo EGDS; \*\* in LRYGP group, postoperative EGDS was not performed; *LSG* – laparoscopic sleeve gastrectomy; *LRYGP* – laparoscopic Roux-en-Y gastric bypass; *EGDS* – esophagogastroduodenoscopy

## 7. DISCUSSION

The knowledge of alterations in stomach physiology after bariatric surgery is an important clinical issue. The sequelae caused by resections or bypasses are persistent and will remain and act over many postoperative years or even decades. Since the number of bariatric operations is growing worldwide, the possibility to evaluate the “health” of the gastric mucosa pre- and postoperatively, using noninvasive methods, would be very attractive, especially in patients after (mini) gastric bypass operation when evaluation of the gastric remnant with routine gastroendoscopy is impossible.

The present study is the first where the utility of biomarker test in the pre-operative management of bariatric surgery patients was evaluated in a 100% biopsy-confirmed clinical setting.

Using gastroscopic biopsies as the gold standard, the concordance between biomarker testing and histology was substantial, with a kappa value of 0.68 and an overall agreement (across 5 categories) of 84.6%. These values favorably compete with those reported in previous validation studies [Agreus et al., 2012; Storskrubb et al., 2008]. This is not unexpected because the biomarker test used (GastroPanel) is based on four biomarkers reflecting the function and structural integrity of the stomach mucosa [Agreus et al., 2012; Sillakivi et al., 2013; Storskrubb et al., 2008; Sipponen et al., 2007; Telerenta-Keerie et al., 2010]. Accordingly, Pepsinogen levels and their ratio are decreased in corpus atrophy, accompanied by elevated G-17. G-17 level also gives indication of gastric acid secretion, being low with high acid output and high when the stomach is acid-free (due to PPI treatment or AG). In antrum atrophy, G-17 is low and does not respond to protein stimulation (lack of G-cells).

In our series, using GP testing, the HS and NHS groups were distinguished with high accuracy. According to the test results, the serum levels of all 4 biomarkers were significantly different between the two groups, remarkably, the marker of inflammation (PgII) was almost three times higher in the NHS group.

As described by many authors, HP is the key causative factor of severe gastric pathology, including peptic ulcer disease and gastric cancer [Agreus et al., 2012; Shrestha et al., 2014; Nomura et al., 1991; Peleteiro et al., 2014]. In the study cohort (Publication II), the prevalence of HP (66%) in candidates for bariatric operation was significantly higher than that reported for bariatric patients in many previous studies from Belgium, Finland and the USA (3.4–17%) [D’Hondt et al., 2013; Peromaa-Haavisto et al., 2013; Gomez et al., 2014], but similar to that reported from Greece and Brazil (53–66%) [Papavramidis et al., 1996; Assef et al., 2015; Dietz et al., 2012]. In the Estonian population, HP prevalence is closely associated with the birth cohort [Oona et al., 2004; Vorobjova et al., 2000]: HP has become more rare among younger generations. In our bariatric cohort, the high HP prevalence is in accordance with the data of earlier studies (56–69%) involving the same birth cohorts of general population in Estonia [Oona et al., 2004; Vorobjova et al., 2000].

Because HP is associated with severe clinical sequelae [Agreus et al., 2012; Shrestha et al., 2014; Nomura et al., 1991; Peleteiro et al., 2014], its eradication is indicated [D'Hondt et al., 2013] and leads to regression of the inflammatory process in the gastric mucosa and significantly reduces the risk for its known complications at the population's level. Indeed, the reported preoperative endoscopic findings (hiatal hernia, 16–25%; esophagitis, 13–30%) from geographic regions with low HP prevalence [D'Hondt et al., 2013, Peromaa-Haavisto et al., 2013, Gomez et al., 2014], as well as from the the high prevalence regions [Papavramidis et al., 1996; Assef et al., 2015; Dietz et al., 2012], are consistent with similar morbidity in our cohort (hiatal hernia, 29%; esophagitis, 21%). Chronic inflammation of the stomach mucosa was detected in 75% (49/65) and atrophy in 6.2% (4/65) of the patients. As expected, gastric diseases (gastritis, 65.1%; AG, 16.7%) are more frequent in regions [Papavramidis et al., 1996, Dietz et al., 2012] with high HP prevalence, like Estonia, as compared with the low-prevalence regions (gastritis, 9.1–28%; AG 0.9%) [D'Hondt et al., 2013; Peromaa-Haavisto et al., 2013; Gomez et al., 2014].

Most of the patients in the NHS group had HP-related gastritis without atrophy. In such cases, gastroscopy is optional if the patient requests it [Agreus et al., 2012]. Gastroscopy is mandatory only in cases of suspected AG or in patients with sustained symptoms. In the NHS group, only four patients had moderate AGA which requires regular monitoring by endoscopy to disclose the eventual progression and increased risk of gastric cancer [Nomura et al., 1991; Benotti et al., 1995]. Of these four AGA cases, only one was clearly confirmed and another one was suspected on the basis of biomarker testing. In the other two cases, AGA was only confirmed with biopsy. In GastroPanel, the G -17 values were within normal limits, implicating that abundant G-cells were still present to sustain the normal G-17 output. Most likely, these cases represent patchy mucosal atrophy instead of a diffuse disease. It is not well established how such patchy atrophy behaves in the long run, and whether regular endoscopic monitoring is indicated or whether biomarker testing is sufficient. It is likely that the gastric mucosa in these patients can significantly recover after HP eradication, while inflammation symptoms diminish or disappear and the process of mucosal atrophy can be arrested, as reported earlier by many authors [Benotti et al., 1995; Lehmann et al., 2000; Storskrubb et al., 2008; Dixon et al., 1996].

In our series, AGA detected by biomarkers was rare, which has been shown earlier [Graham et al., 2006]. A recent meta-analysis of the published GastroPanel literature confirmed that the test works better for detection of AGC (Pgl, Pgl/PgII ratio) compared to AGA (G-17). A simple explanation is that low G-17 levels can result from two distinct causes: AGA and high acid output [Agreus et al., 2012; Syrjänen K., 2016; Sipponen et al., 2005]. No one biomarker that is regulated by more than one trigger can be a highly specific indicator among the others [Syrjänen K., 2016]. To make distinction between these two (AGA, high acid output), it is mandatory to test G-17 after protein

stimulation (G-17s). Failure to increase G17s output implicates lack of G-cells and presence of AGA [Syrjänen K., 2016].

Another explanation for the rarity of AGA in our series could be the relatively young age of the patients. In fact, GastroPanel was not primarily designed for testing bariatric surgery patients, but rather for diagnosis and screening of elderly patients with AG and for screening for the increased risk of gastric cancer [Teleranta-Keerie et al., 2010]. However, bariatric surgery can be safely performed also in patients aged 60 years or older [Giordano et al., 2015; Eisenberg et al., 2022]. In this sub-group, the potential role of the gastric biomarker test can be particularly important, as the incidence of atrophy and gastric cancer increases with age. Furthermore, using the biomarker test, we could easily diagnose almost all HP-infections and administer timely treatment to diminish the risk of AG and gastric cancer.

There was also one false positive “panatrophy” (according to GP) in our series, while histologically only superficial HP related gastritis was confirmed. Rather, this fact could be related to technical issues.

In patients with AG, follow-up EGDS is still needed. Thus the sleeve gastrectomy method (SG) would be preferable, because routine EGDS after bypass operation is unfeasible. In large series of operated patients, however, practically no post-operative problems have been reported for the bypass group [Safatle-Ribeiro et al., 2007; Csendes et al., 2012; Csendes et al., 2006]. Only a few case reports are available on postoperative cancer [Escalona et al., 2005; Khitin et al., 2003; Ribeiro et al., 2013].

Regarding the use of the normal GP profile as a surrogate for the healthy stomach (HS), 22/65 subjects were categorised into this group according to its criteria. Clinically, 20 of them were asymptomatic, had no history of abdominal complaints, and only 2 had reflux symptoms. On EGDS, only minor abnormalities were detected that were considered clinically insignificant: non-HP gastritis, mild or moderate degree esophagitis, or gastric mucosa erosions. It is clear that management of these disorders does not require a delay in elective surgery, nor is it a contraindication for operation [Parikh et al., 2016]. In 4 cases, esophagitis (LA grade A/B) was found to be associated with hiatal hernia, and 2 of these subjects reported reflux complaints. According to international consensus [Anderson et al., 2008; Parkin DM., 2004], for patients with upper abdominal complaints, endoscopic investigation is indicated. In patients with symptomatic esophagitis, the recommended surgical procedure could be gastric bypass rather than sleeve gastrectomy. Although the opinions on the use of sleeve gastrectomy in esophagitis are controversial [Hawasli et al., 2016], the probability of complicated esophagitis has been shown to increase postoperatively [Melissas et al., 2015; Graham et al., 2006]. Such cases respond poorly to medical treatment [Stenard et al., 2015], despite the fact that a major portion of the gastric corpus is resected, which results in significant reduction in parietal cell mass and a decline in acid output. Current conditional recommendation from the Interdisciplinary European Guidelines on Metabolic and Bariatric

Surgery suggest not to perform LSG in patients with severe GERD symptoms and/or severe esophagitis [Di Lorenzo et al. 2020].

In a study of Sipponen et al. the authors demonstrated that low G17b levels in the general population are a marker of high basal acid output, which in turn predisposes to gastric acid reflux and esophagitis [Sipponen et al., 2005]. In this series, however, we failed to find correlation between esophagitis and low G17b levels, as only one out of the 4 patients in the HS group and 2/10 in the NHS group showed G17b levels below the cut-off value. Although some studies have obtained results similar to ours [Monkemuller et al., 2008; Peitz et al., 2011], there are also reports on such correlation between G-17 and esophagitis [Goni et al., 2015].

In the light of the above data, it is evident that in symptomatic esophagitis, endoscopy plays a role also in guiding the selection of the surgical method (i.e., preferring gastric bypass over gastric sleeve), which is crucial to ensure optimal treatment outcome. In our series, 3 asymptomatic patients in the HS group had, despite the normal marker profile, erosions in the stomach (antrum), with a Lanza score of less than 4 (i.e. not severe). Recently, some authors have reported gastric erosions in bariatric surgery patients [Fernandes et al., 2016] and others have reported them also in asymptomatic volunteers in population studies, more frequently in HP-negative than HP-positive subjects [Lehmann et al., 2000]. Although the cause of such erosions may be multifactorial, all 3 patients in our study took several medications known to damage the gastric mucosa. Yet the erosions seen in the HS group can be considered clinically insignificant: the patients were asymptomatic, and no complications like hemorrhage were found on EGDS. Accordingly, we cannot consider minor erosions in patients with the normal biomarker profile as an indication for changing treatment practises in these bariatric surgery patients.

Our data demonstrate that gastric biomarker test can definitely help select asymptomatic patients (20/22 in this series) with minor but clinically non-significant gastric mucosa alterations for whom preoperative endoscopic investigation can be safely replaced by non-invasive biomarker test. Endoscopy should only be reserved for symptomatic patients to confirm the diagnosis and to opt for the surgical method, as has been pointed out earlier [Anderson et al., 2008; Parkin DM., 2004].

Regarding NHS patients, the rationale should be the same as for HS patients: those with reflux complaints should undergo endoscopic investigation to confirm the diagnosis and to plan possible preoperative treatment.

Endoscopic findings in bariatric surgery patients can be highly variable [Fernandes et al., 2016]. To avoid postoperative complications, including ulcer [Fernandes et al., 2016], it is important to evaluate the patients pre-operatively to detect (by using GP) [Agreus et al., 2012] and eradicate HP infection. Although there were no cases of ulcer disease in our material, these steps are always important in this special group of patients. In the case of suspected peptic ulcer, EGDS is essential; the same applies to patients with a family history of gastric cancer.

The results of our study confirm that the normal biomarker profile in the test is an excellent surrogate for the healthy stomach, and this non-invasive test could replace EGDS in the pre-operative management of bariatric surgery patients. Indeed, using biomarker test, it could have been possible to avoid EGDS in 20/22 patients in the HS group, i.e, in 31% (20/65) of all bariatric patients in our cohort. These asymptomatic patients with the normal biomarker profile are at a very low risk to develop a clinically significant disease in the gastric mucosa, including peptic ulcer and gastric cancer [Agreus et al., 2012; Sipponen et al., 1993].

As expected, both operative methods (LSG, 73% and LRYGB, 65%) yielded satisfactory 1-year %EWL results, with the data being comparable to relevant literature data (Perrone et al., 2017; McNickle AG et al., 2017). Similar results (LSG, 67% and LRYGB, 63%) of EWL% at 2 years postoperatively can be found in our other study (Publication I).

Little is known about the status of the gastric mucosa and possible changes in gastric biomarkers after bariatric operations. Our studies demonstrate that marked dynamic changes in the biomarker profile can be detected in both operated groups (LRYGB and LSG) during 2- year follow-up after bariatric surgery (Publications III, I).

A major decrease in PgI and PgII values was noted in the LSG patients already during the first postoperative days; the values stabilised after 3 months and remained low for at least one year postoperatively. In 82% of the patients, PgI was below the reference value (30 µg/L), normally indicating severe atrophy of the corpus mucosa in a non-operated stomach. Similar results (reduction in PgI levels) have been shown in an earlier study in peptic ulcer patients 1 year after vagotomy and after Billroth I gastric resection (the antrum and a segment of the gastric corpus being removed). In that study none of the gastric ulcer patients developed atrophy of the gastric corpus mucosa although they showed a pronounced decrease of Pg I in comparison with its preoperative level [Peetsalu et al., 1990].

In the LRYGB patients, the increase of PgI and PgII level during the first postoperative days was of short duration and can be attributed to operation related trauma. Starting from the first postoperative month until the end of the first postoperative year, the PgI values for about 60% of the patients were below the reference range. Similar results can be found in a study by Sundbom et al. [Sundbom et al., 2007].

The dynamics of G17f differed between the two study groups. In the case of LSG, its values were higher than the reference values in 61% of the patients at postoperative day 1, however, by the end of the first postoperative year, the proportion of such patients had stabilised at 32%. In the LRYGB patients, the levels of G17b were low. As no protein stimulant reached the gastric antrum, there was no response of G17s to any stimulant and the levels of G17s were as low as the levels of G17b.

Our study demonstrates dynamic changes in the serum levels of the studied gastric biomarkers after operative treatment during a 1-year follow-up period. The changes are primarily caused by alterations induced by the operation and were more pronounced in the LSG than in the LRYGB patients. As confirmed by biopsies, these changes in biomarker levels did not coincide with any morphological changes in the gastric mucosa during one year of follow-up. Similar results in the mean values of the gastric biomarkers can be seen at 2 years postoperatively in both the LSG and LRYGB groups (Publication I). Given that both studies were conducted on the same (bariatric) population, one can anticipate that changes in the gastric biomarker levels will be stable at least during a period of 1–2 years postoperatively in case there is no weight regain.

In the absence of a detectable mucosal pathology, the obtained biomarker findings suggest „reduction“ and „down-regulation“ in the function of the gastric corpus in particular; one could also implicate the status of hypoacidity in fasting gastric remnants after both operations. Accordingly, in the LSG patients, the PgI and PgII levels decreased and the G17b levels increased. Such a rapid change can be explained by the fact that most of the gastric corpus in the LSG patients was resected and hence the number of PgI producing cells decreased significantly, as suggested in our Publication I [Sillakivi et al., 2013]. By resecting a substantial proportion of the stomach corpus and fundus, also a huge parietal cell mass is removed from the corpus, resulting in a marked reduction in gastric acid output. The antral G cells respond quickly to the decrease in acid output and boost their G17 output. A similar effect on G17 values is seen also in patients during PPI treatment: hypochlorhydria leads to the stimulation of G17 production [Agreus et al., 2009]. Increased G17 production is the normal function of the atrophy-free gastric antral mucosa. Acid secretion is reduced, although the histology of the corpus biopsies remains unaffected. Because of hypoacidity, there may arise the risk for malabsorption of micronutrients (iron, calcium, etc) and vitamin-B12 [Schubert ML., 2014; Schubert ML., 2015; Sipponen et al., 2003].

Unlike in LSG, nothing is removed from the abdominal cavity during LRYGB operation, where a small pouch created from the proximal part of the stomach is anastomosed to the small bowel leaving 90% of the primary stomach out of food passage. In our studies, changes in the gastric biomarker levels are less pronounced in this group as compared to the LSG patients. The increase of PgI and PgII in the first few postoperative days is most likely caused by irritation (inflammation) due to surgical manipulation. At the late follow-up of the LRYGB patients, the decrease of PgI and PgII may be explained by the resting status of the stomach, as proposed also earlier [Sundbom et al., 2003]. Also, fast food passage from the small pouch to the small intestine can even further reduce secretion in the bypassed stomach.

We noted a decrease of PgI from month 1 onwards, followed by stable, abnormally low values in 60% of the patients after one year. At the 1-year follow-up visit, the G17 values among 11% of the LRYGB patients were above the upper normal limit. It can be speculated that this is the result of a reaction

due to vagotomy which decreases gastric acid secretion in the bypassed stomach.

The small pouch cannot produce PgI in adequate amounts. The reason is not atrophy of the pouch or remnant itself but the insufficient mucosal mass involved in PgI production in the pouch. At the same time, acid secretion in the bypassed stomach may still take place, since G17b is mostly within normal limits and not increased (as should be the case with the acid-free stomach). In the bypassed stomach, however, acid secretion takes place in a blind loop and does not counteract the risk of malabsorption. In fact, the risk of postoperative malabsorption must be even higher after LRYGB than after LSG.

The mean PgII values in both operation groups changed at first parallel with the mean values of PgI, being lower at the end of the first postoperative month and increasing thereafter. By the end of year 1, the initial reduction had diminished to only 30% and 14% of the mean preoperative values in the LSG and LRYGB groups, respectively. It should be noted that PgII is produced not only in the corpus but also in the other parts of the stomach (antrum, fundus, duodenum). The above changes can be explained by mucous neck cell hyperplasia, which has been demonstrated in animal experiments [Arapis et al., 2015].

In the present setting, the gastric mucosal pathology in the LSG group, both preoperatively and at 1-year follow-up, was mainly related to HP-associated chronic (non-atrophic) gastritis.

Alterations were more pronounced in the antrum than in the corpus, which is consistent with earlier data [Onzi et al., 2014]. Atrophic gastritis, which invariably affects the gastric biomarker profile, was diagnosed in only one patient preoperatively but was no more detectable at 1-year follow-up. This resolution can be attributed to successful HP eradication therapy, which is well documented in the literature [Zhou et al., 2003; Benotti et al., 1995; Lehmann et al., 2000]. Another option, which is hard to exclude, is that atrophy in the preoperative biopsies was patchy and the right focus was missed in the follow-up biopsies.

Although we do not have postoperative gastrobiopsies from the LRYGB group, then according to the literature, no substantial changes in the mucosa have been found either in the gastric pouch or in the gastric remnant (Csendes et al., 2006; Safatle-Ribeiro et al., 2007; Csendes et al., 2012).

According to our study, the preoperative gastric biomarker levels are not associated with %EWL at 1-year follow-up, which precludes the prospect to predict weight loss, or to select an appropriate operation type preoperatively on the basis of the GP biomarker profile. The GP was designed for screening the atrophy of gastric mucosa in asymptomatic subjects and should not be used as a screening test for gastric cancer. Similarly, as suggested by Marchesi et al., GP test is of limited value in predicting morphological changes in the stomach mucosa after bariatric surgery, as is shown also in this study, due to the simple reason that any such morphological changes are practically non-existent [Marchesi et al., 2017]. In patients who underwent bariatric surgery, the value

of the gastric biomarkers lies in their applicability in the follow-up period. GP test gives an opportunity to follow up the gastric status after surgery, and in LSG patients, to estimate the severity of functional disturbance in acid output.

Monitoring stomach physiology after bariatric surgery is clinically important, because the sequelae caused by resection or bypass may interfere with the welfare and health of the operated subjects in the long run. A hypoacidic stomach remnant, which is common after LSG, and bypassing of food passage in LRYGB, may impair digestion, as well as absorption of vitamins (eg. vitamin-B12), micronutrients and some pharmaceuticals [Schubert ML., 2014; Schubert ML., 2015].

Further studies will provide more knowledge and indices for specifying whether changes in these functions are clinically relevant or not and whether they are related, for example, to malabsorption of vitamins, micronutrients and medicines in the long term.

Our study also has some weaknesses. The number of patients is too small for using sensitivity and specificity in order to compare histology findings and biomarkers preoperatively. A limitation of our study is that in one study group the knowledge of the preoperative plasma biomarker levels and stomach histology is lacking. This study does not report surgical complications. Another weakness of our study is the lack of postoperative gastric biopsies in the LRYGB group. According to other studies, however, there are no major histopathological changes either in the pouch or in the bypassed stomach during the first postoperative years [Csendes et al., 2006; Safatle-Ribeiro et al., 2007; Csendes et al., 2012). Also, it should be taken into account that the population is limited, as the study was conducted at a single center and most of the patients were residents of only the southern and eastern parts of Estonia. The full benefits of the non-invasive biomarker screening of bariatric surgery patients pre- and postoperatively can only be established in larger cohorts, with participants from different populations and with different prevalences of HP-infection.

### **Future perspectives**

- The studied biomarkers could hypothetically enable to abandon invasive preoperative EGDS among some patients. It must be considered that in recent years the indications for endoscopy have broadened, mostly due to the problems caused by postoperative GERD.
- At present, the biomarkers that we used in our work did not enable to predict the operative success nor aid the selection of operation method. Further studies are needed to better select patients for optimal treatment.
- Nowadays there are no good guidelines for choosing between different treatment options (medical vs surgical treatment).
- Currently surgical treatment is superior, but presumably proportion of medical treatment will increase in the future.
- New medications are promising, but studies are still in progress. Long term outcomes and persistency of results over time have major importance.

## 8. CONCLUSIONS

1. The results of our study confirm that the normal biomarker profile in GP test is an excellent surrogate for the healthy stomach. In asymptomatic obese patients with the normal biomarker profile, this non-invasive test could replace preoperative EGDS. According to the results of our study, preoperative EGDS could have been avoided in 31% of the patients of the study group (20/65).
2. The changes in the biomarker levels are stable for at least 2 years post-operatively. Biomarker profile in SG and RYGB group are quite similar one and two years after surgery.
3. Our study demonstrates that significant (dynamic) changes occur in the biomarker profile after bariatric surgery. The changes are primarily caused by operation and do not indicate pathological changes in gastric mucosa. The changes are more pronounced in the LSG patient group.
4. Preoperative gastric biomarker levels do not correlate with %EWL at 1-year follow up, indicating that the operative success in %EWL cannot be predicted on the basis of the preoperative GP marker profile.

## 9. SUMMARY IN ESTONIAN

### Mao biomarkerid ja nende dünaamika kui väheinvasiivne meetod mao seisundi hindamiseks bariaatrilise kirurgia patsientidel

#### 9.1. Sissejuhatus

Viimastel aastakümnetel on ülekaalulisuse ja rasvumise levimus oluliselt suurenenud ning sellest on saanud tõsine terviserisk nii arengu- kui arenenud riikides. Tänapäeval on rasvumine muutunud tervishoiu prioriteediks üle kogu maailma [Ogden et al., 2006; Benotti et al., 1995; Williams et al. 2015; Kelly et al. 2015]. Maailma Tervishoiu Organisatsiooni (WHO) andmetel oli 2016. aastal rohkem kui 1,9 miljardit maailma täiskasvanud elanikkonnast ülekaalulised, neist omakorda üle 650 miljoni olid rasvunud [WHO, 2021]. Ülekaalulisus ja rasvumine on gastroösofagealse reflukshaiguse, ösofagiidi, söögitorulahisonga, söögitoru Barret metaplaasia, söögitoru adenokartsionoomi ja *Helicobacter pylori* riskifaktoriteks. Ülekaalulistel patsientidel esineb eelpool nimetatud patoloogiad 2–3 korda sagedamini kui normaalkaalulistel [Gerson LB., 2009].

Paralleelselt rasvunute arvu suurenemisega on pidevalt tõusnud bariaatriliste operatsioonide arv maailmas, kuna raskekujulise rasvumise korral on kirurgiline sekkumine ainus tõhus ja kestvat toimet pakkuv ravimeetod [Flum et al., 2007; Azagury et al., 2011].

Varasemad uuringud haavandtõve patsientidel vagotoomiate ja mao osaliste resektsioonide järgselt on näidanud, et operatsioonid maol tekitavad muutusi nii jääkmao limaskestal kui ka mao sekretsioonivõimes [Peetsalu et al., 1990]. Siiski on teadmised bariaatriliste operatsioonide järgselt tekkinud mao limaskesta muutuste osas piiratud.

Bariaatriliste patsientide pre- ja postoperatiivses käsitluses on mao seisundi hindamise ja patoloogiate tuvastamise kuldseks standardiks olnud seedetrakti ülaosa endoskoopiline uuring koos mao limaskesta biopsiaga. Samas puudub paljudel autoritel üksmeel asümptomaatilistele patsientidele teostatava rutiinse endoskoopilise uuringu vajalikkuse osas [Parikh et al., 2016; Csendes et al., 2007; D'Hondt et al., 2013; Almazeedi et al., 2013; Gerson LB., 2009].

Viimastel aastakümnetel on järjest enam populaarsust kogunud vereproovil baseeruv biomarkerite testpaneel (“seroloogiline biopsia”), mis aitab väheinvasiivset meetodit kasutades diagnoosida mao funktsionaalseid muutusi ja limaskesta patoloogiad, sh. *Helicobacter pylori* infektsiooni ja limaskesta atroofiat [Agreus et al., 2012]. Eelpool nimetatud testpaneel põhineb neljal maospetsiifilisel biomarkeril (Pepsinogeen I ja II (PgI, PgII), Gastriin-17 (G-17)), *Helicobacter pylori* antikeha (HPAK)), mille väärtuste põhjal saab eristada 8 erinevat markerite profiili [Agreus et al., 2012; Syrjänen et al. 2019].

Informatsioon bariaatrilise operatsiooni järgselt tekkinud muutuste kohta mao füsioloogias omab olulist kliinilist tähendust, kuna operatsioonil teosta-

tavate reseksioonide ja möödajuhtimistega kaasnevad muutused on püsivad ja kestavad aastaid. Arvestades bariaatriliste operatsioonide arvu kasvu, oleks “väheinvasiivse” meetodi kasutamine mao limaskesta seisundi hindamiseks pre- ja postoperatiivselt väga hea lahendus, eriti patsientide puhul, kellele on tehtud möödajuhtiv operatsioon.

Meie uurimistöö eesmärgiks oli välja selgitada, kas biomarkerite testi abil oleks võimalik vältida invasiivse endoskoopilise uuringu tegemist bariaatrilisele operatsioonile minevatele asümptomaatilistele patsientidele, ning kirjeldada mao biomarkerite profiili muutust mao vertikaalse reseksiooni (SG) ja maost möödajuhtiva operatsiooni (RYGB) järgselt. Muutused biomarkerite profiilis võiksid anda uut informatsiooni maovähendusoperatsiooni tõttu tekkinud võimalike komplikatsioonide ja kaebuste kohta ning aidata läbi viia patsiendipõhist ravi (sh. asendusravi) konkreetse operatsioonitüübi järgselt.

## 9.2. Uuringu eesmärgid

- 1) Selgitada välja, kas väheinvasiivse seroloogilise biomarkerite testi kasutamine aitaks preoperatiivselt vältida invasiivset endoskoopilist uuringut plaanilisele bariaatrilisele operatsioonile minevate patsientide hulgas.
- 2) Uurida mao limaskesta funktsiooni ja happe sekretsiooni bariaatrilise operatsiooni järgselt – millised muutused toimuvad mao biomarkerite profiilis kahe aasta jooksul peale operatsiooni.
- 3) Hinnata, kas postoperatiivselt tekkinud muutused mao biomarkerite profiilis on seotud operatsioonist tingitud anatoomiliste muutustega või on nende põhjuseks mao limaskesta patohistoloogilised muutused; samuti hinnata, kui kiiresti tekivad muutused biomarkerite tasemes postoperatiivselt.
- 4) Hinnata, kas mao biomarkerite pre- ja postoperatiivsed väärtused korreleeruvad ühe aasta liigse kehakaalu langusega.

## 9.3. Uuritavad ja meetodid

Antud uuring baseerub 105 patsiendi andmetel, kellele teostati ajaperioodil 2008–2016 Tartu Ülikooli Kliinikumi kirurgikliinikus maovähendusoperatsioon. Uuringusse kaasatud patsiendid olid vähemalt 18-aastased, vastasid bariaatrilise operatsiooni kriteeriumitele (kehamassiindeks (KMI)  $>40$  või  $>35$  koos rasvumisest põhjustatud kaasuva haigusega) ning kes andsid nõusoleku uuringus osalemiseks. Uuringusse ei kaasatud alla 18-aastaseid patsiente, kriteeriumitele mittevastavaid patsiente ning neid, kes ei soovinud uuringus osaleda.

Uuringus osalenud patsiente intervjueriti, neil võeti vereanalüüsid *Gastropanel* testi jaoks ning teostati endoskoopiline uuring koos mao limaskesta biopsiate võtmisega eri uuringugruppidel erinevatel ajaperioodidel. Kõikidele uuringu patsientidele teostati kas laparoskoopiline SG või laparoskoopiline RYGB.

Intervjuu läbiviimine ning füüsiliste parameetrite (pikkus, kaal, KMI, vööümbermõõt) fikseerimine toimus enne vereanalüüside võtmist ja endoskoopilist uuringut ning enne operatsiooni (Publikatsioon II), samuti postoperatiivsel jälgimisperioodil 1, 3, 12 kuu möödudes (Publikatsioon III) ja 2 aastat peale bariaatrilist operatsiooni (Publikatsioon I).

Vereanalüüsid *Gastropanel* testi jaoks võeti preoperatiivselt ning seejärel postoperatiivselt 1., 3. päeval, 1., 3., 12. kuul ning 2 aasta pärast (erinevates uuringu gruppides).

Endoskoopiline uuring koos mao limaskesta biopsiate võtmisega teostati preoperatiivselt (Publikatsioonid II ja III) ja 12 kuud peale bariaatrilist operatsiooni (ainult SG grupi patsientidel) (Publikatsioon III).

Uurimustöö esimeses etapis uuriti preoperatiivselt maospetsiifilisi biomarkereid ja mao limaskesta biopsiaid. *Gastropanel* testi tulemuste alusel jagati patsiendid kahte gruppi: terve magu (HS) ja haige magu (NHS). Mõlema grupi biomarkerite testi tulemusi võrreldi otseselt mao limaskesta biopsiate vastustega (Publikatsioon II).

Uurimustöö teises etapis võrreldi maospetsiifiliste biomarkerite dünaamikat kahe erineva operatsioonigrupi (SG ja RYGB) vahel kahe operatsioonijärgse aasta jooksul (Publikatsioonid III ja I).

## 9.4. Tulemused

*Gastropanel* testi tulemuste alusel kuulusid 22 patsienti HS gruppi ning 43 patsienti NHS gruppi. Biomarkerite (PgI, PgII, PgI/PgII ja IgG) väärtused erinesid kahe grupi vahel märkimisväärselt ( $p < 0,001$ ). *Gastropanel* testi tulemuste ja mao limaskesta biopsiate vastuste omavaheline kokkulangevus oli väga hea (Kappa=84,6%).

Mao biomarkerite tasemes ilmetusid postoperatiivselt kahe operatsioonigrupi (SG ja RYGB) vahel suured erinevused. SG grupis langes PgI tase kiiresti peale operatsiooni ning 1 aasta möödudes oli PgI tase märkimisväärselt madalam võrreldes operatsioonieelse tasemega ( $p < 0,0001$ ). Samasugune (madal) PgI tase esines ka kaks aastat peale operatsiooni. RYGB grupis langes PgI tase alates esimesest postoperatiivsest kuust ning esimeseks postoperatiivseks aastaks oli toimunud märkimisväärne langus võrreldes operatsioonieelse tasemega ( $p < 0,0001$ ), kusjuures madal tase püsis ka kahe aasta möödudes.

Võrreldes operatsioonieelsete väärtustega toimus ühe aasta möödudes PgII ja PgI/PgII tasemes märkimisväärne langus mõlemas operatsioonigrupis (SG grupp, PgII  $p < 0,0006$ ; PgI/PgII  $p < 0,0001$ ).

Basaal-G17 tase tõusis SG grupis esimese ja teise aasta lõpuks, samas kui RYGB grupis basaal-G17 tase langes 12 kuu möödudes 28% võrreldes algväärtusega.

SG grupis täheldati ühe ja kahe aasta möödudes operatsioonist stimuleeritud G17 (G17s) normaalset taset (vastus stimulatsioonile  $> 3$  pmol/l) vastavalt 40% ja 100% juhtudest. RYGB grupis oli G17 vastus stimulatsioonile adekvaatne vaid vastavalt 7,4% ja 25% juhtudest.

Postoperatiivsed gastrobiospiad (SG grupis) kirjeldasid kroonilise gastriidi esinemist 92% patsientidest, atroofiat ei esinenud.

Pre- ja postoperatiivsete (ühe aasta) ega preoperatiivsete/postoperatiivsete (ühe aasta) biomarkerite tase ei korreleerunud kummaski operatsioonigrupis 1 aasta liigse kehakaalu languse protsendiga.

## 9.5. Järeldused

- 1) Meie uuringu tulemused kinnitavad, et *Gastropanel* testi normaalne biomarkerite profiil peegeldab hästi terve mao seisundit, mistõttu võiks antud väheinvasiivne test asümptomaatilistel rasvunud patsientidel asendada preoperatiivset endoskoopilist uuringut. Käesoleva uuringu tulemuste alusel oleks preoperatiivne endoskoopiline uuring olnud välditav 31% uuringu patsientidest (20/65).
- 2) Muutused biomarkerite tasemes püsivad stabiilsena vähemalt kaks aastat peale operatsiooni. SG ja RYGB grupis toimunud muutused biomarkerite tasemes on sarnased 1. ja 2 aasta möödudes operatsioonist.
- 3) Meie uuringute tulemused näitavad, et bariaatrilise operatsiooni järgselt toimuvad lühikese aja jooksul (juba päevadega) märkimisväärsed muutused biomarkerite profiilis. Nende põhjuseks on operatsioon ise, mitte aga mao limaskesta patoloogia. Muutused on rohkem väljendunud SG patsientide grupis.
- 4) Preoperatiivne mao biomarkerite tase ei korreleeru esimese postoperatiivse aasta liigse kehakaalu languse protsendiga, mistõttu nende alusel ei saa ennustada operatsiooni edukust kaalulanguse aspektist.

## REFERENCES

- Agréus L, Kuipers EJ, Kupcinskis L, Malfertheiner P, Di Mario F, Leja M, Mahachai V, Yaron N, van Oijen M, Perez Perez G, Rugge M, Ronkainen J, Salaspuro M, Sipponen P, Sugano K, Sung J: Rationale in diagnosis and screening of atrophic gastritis with stomach-specific plasma biomarkers. *Scandinavian journal of gastroenterology* 2012;47:136–147.
- Agréus L, Storskrubb T, Aro P, Ronkainen J, Talley NJ, Sipponen P: Clinical use of proton-pump inhibitors but not H2-blockers or antacid/alginates raises the serum levels of amidated gastrin-17, pepsinogen I and pepsinogen II in a random adult population. *Scandinavian journal of gastroenterology* 2009;44:564–570.
- Alexandratos N BJ: World Agriculture Towards 2030/2050. <https://sefari.scot/sites/default/files/documents/FAO%20%282012%29%20World%20Agriculture%20Towards%202030%20and%202050.pdf>. Accessed April 12, 2023. 2012.
- Almazeedi S, Al-Sabah S, Al-Mulla A, Al-Murad A, Al-Mossawi A, Al-Enezi K, Jumaa T, Bastaki W: Gastric histopathologies in patients undergoing laparoscopic sleeve gastrectomies. *Obesity surgery* 2013;23:314–319.
- An S, Park HY, Oh SH, Heo Y, Park S, Jeon SM, Kwon JW: Cost-effectiveness of Bariatric Surgery for People with Morbid Obesity in South Korea. *Obesity surgery* 2020;30:256–266.
- Anderson MA, Gan SI, Fanelli RD, Baron TH, Banerjee S, Cash BD, Dominitz JA, Harrison ME, Ikenberry SO, Jagannath SB, Lichtenstein DR, Shen B, Lee KK, Van Guilder T, Stewart LE: Role of endoscopy in the bariatric surgery patient. *Gastrointestinal endoscopy* 2008;68:1–10.
- Angrisani L, Santonicola A, Iovino P, Ramos A, Shikora S, Kow L: Bariatric Surgery Survey 2018: Similarities and Disparities Among the 5 IFSO Chapters. *Obesity surgery* 2021;31:1937–1948.
- Angrisani L, Santonicola A, Iovino P, Vitiello A, Higa K, Himpens J, Buchwald H, Scopinaro N: IFSO Worldwide Survey 2016: Primary, Endoluminal, and Revisional Procedures. *Obesity surgery* 2018;28:3783–3794.
- Apovian CM, Aronne LJ, Bessesen DH, McDonnell ME, Murad MH, Pagotto U, Ryan DH, Still CD: Pharmacological management of obesity: an endocrine Society clinical practice guideline. *The Journal of clinical endocrinology and metabolism* 2015;100:342–362.
- Apovian CM, Istanbuli NW: Obesity: Guidelines, Best Practices, New Research. *Endocrinology and metabolism clinics of North America* 2016;45:xvii–xviii.
- Arapis K, Cavin JB, Gillard L, Cluzeaud F, Lettéron P, Ducroc R, Le Beyec J, Hourseau M, Couvelard A, Marmuse JP, Le Gall M, Bado A: Remodeling of the residual gastric mucosa after roux-en-y gastric bypass or vertical sleeve gastrectomy in diet-induced obese rats. *PloS one* 2015;10:e0121414.
- Armstrong D, Bennett JR, Blum AL, Dent J, De Dombal FT, Galmiche JP, Lundell L, Margulies M, Richter JE, Spechler SJ, Tytgat GN, Wallin L: The endoscopic assessment of esophagitis: a progress report on observer agreement. *Gastroenterology* 1996;111:85–92.
- Assef MS, Melo TT, Araki O, Marioni F: EVALUATION OF UPPER GASTRO-INTESTINAL ENDOSCOPY IN PATIENTS UNDERGOING BARIATRIC SURGERY. *Arquivos brasileiros de cirurgia digestiva: ABCD = Brazilian archives of digestive surgery* 2015;28 Suppl 1:39–42.

- Azagury DE, Abu Dayyeh BK, Greenwalt IT, Thompson CC: Marginal ulceration after Roux-en-Y gastric bypass surgery: characteristics, risk factors, treatment, and outcomes. *Endoscopy* 2011;43:950–954.
- Azagury DE, Lautz DB: Obesity overview: epidemiology, health and financial impact, and guidelines for qualification for surgical therapy. *Gastrointestinal endoscopy clinics of North America* 2011;21:189–201.
- Benotti PN, Forse RA: The role of gastric surgery in the multidisciplinary management of severe obesity. *American journal of surgery* 1995;169:361–367.
- Biohit: Gastropanel- interpretation guide snapshot. [www.gastropanel.com/healthcare-professionals-and-laboratories/interpreting-results](http://www.gastropanel.com/healthcare-professionals-and-laboratories/interpreting-results). Accessed April 17, 2023. 2023.
- Blüher M: Obesity: global epidemiology and pathogenesis. *Nature reviews Endocrinology* 2019;15:288–298.
- Borisenko O, Lukyanov V, Johnsen SP, Funch-Jensen P: Cost analysis of bariatric surgery in Denmark made with a decision-analytic model. *Danish medical journal* 2017a;64.
- Borisenko O, Mann O, Duprée A: Cost-utility analysis of bariatric surgery compared with conventional medical management in Germany: a decision analytic modeling. *BMC surgery* 2017b;17:87.
- Bray GA, Kim KK, Wilding JPH: Obesity: a chronic relapsing progressive disease process. A position statement of the World Obesity Federation. *Obesity reviews: an official journal of the International Association for the Study of Obesity* 2017;18:715–723.
- Buchwald H, Avidor Y, Braunwald E, Jensen MD, Pories W, Fahrenbach K, Schoelles K: Bariatric surgery: a systematic review and meta-analysis. *Jama* 2004;292:1724–1737.
- Buchwald H, Estok R, Fahrenbach K, Banel D, Sledge I: Trends in mortality in bariatric surgery: a systematic review and meta-analysis. *Surgery* 2007;142:621–632; discussion 632–625.
- Butland B JS, Kopelman P, McPherson K, Thomas S, Mardell J, Parry V: Tackling Obesities: Future Choices – Project report. [https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment\\_data/file/287937/07-1184x-tackling-obesities-future-choices-report.pdf](https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/287937/07-1184x-tackling-obesities-future-choices-report.pdf); in.
- Campbell JE, Drucker DJ: Pharmacology, physiology, and mechanisms of incretin hormone action. *Cell metabolism* 2013;17:819–837.
- Chooi YC, Ding C, Magkos F: The epidemiology of obesity. *Metabolism: clinical and experimental* 2019;92:6–10.
- Clément K, Vaisse C, Lahlou N, Cabrol S, Pelloux V, Cassuto D, Gormelen M, Dina C, Chambaz J, Lacorte JM, Basdevant A, Bougnères P, Lehoucq Y, Froguel P, Guy-Grand B: A mutation in the human leptin receptor gene causes obesity and pituitary dysfunction. *Nature* 1998;392:398–401.
- Coleman DL, Hummel KP: Effects of parabiosis of normal with genetically diabetic mice. *The American journal of physiology* 1969;217:1298–1304.
- Colquitt JL, Pickett K, Loveman E, Frampton GK: Surgery for weight loss in adults. *The Cochrane database of systematic reviews* 2014;2014:Cd003641.
- Colquitt JL, Picot J, Loveman E, Clegg AJ: Surgery for obesity. *The Cochrane database of systematic reviews* 2009;Cd003641.
- Csendes A, Burgos AM, Smok G, Beltran M: Endoscopic and histologic findings of the foregut in 426 patients with morbid obesity. *Obesity surgery* 2007;17:28–34.

- Csendes A, Orellana O, Martínez G, Burgos AM, Figueroa M, Lanzarini E: Clinical, Endoscopic, and Histologic Findings at the Distal Esophagus and Stomach Before and Late (10.5 Years) After Laparoscopic Sleeve Gastrectomy: Results of a Prospective Study with 93% Follow-Up. *Obesity surgery* 2019;29:3809–3817.
- Csendes A, Smok G, Burgos AM: Endoscopic and histologic findings in the gastric pouch and the Roux limb after gastric bypass. *Obesity surgery* 2006;16:279–283.
- Csendes A, Smok G, Burgos AM, Canobra M: Prospective sequential endoscopic and histologic studies of the gastric pouch in 130 morbidly obese patients submitted to Roux-en-Y gastric bypass. *Arquivos brasileiros de cirurgia digestiva: ABCD = Brazilian archives of digestive surgery* 2012;25:245–249.
- D'Hondt M, Steverlynck M, Pottel H, Elewaut A, George C, Vansteenkiste F, Van Rooy F, Devriendt D: Value of preoperative esophagogastroduodenoscopy in morbidly obese patients undergoing laparoscopic Roux-en-Y gastric bypass. *Acta chirurgica Belgica* 2013;113:249–253.
- Davies DJ, Baxter JM, Baxter JN: Nutritional deficiencies after bariatric surgery. *Obesity surgery* 2007;17:1150–1158.
- Di Lorenzo N, Antoniou SA, Batterham RL, Busetto L, Godoroja D, Iossa A, Carrano FM, Agresta F, Alarçon I, Azran C, Bouvy N, Balaguè Ponz C, Buza M, Copaescu C, De Luca M, Dicker D, Di Vincenzo A, Felsenreich DM, Francis NK, Fried M, Gonzalo Prats B, Goitein D, Halford JCG, Herlesova J, Kalogridaki M, Ket H, Morales-Conde S, Piatto G, Prager G, Pruijssers S, Pucci A, Rayman S, Romano E, Sanchez-Cordero S, Vilallonga R, Silecchia G: Clinical practice guidelines of the European Association for Endoscopic Surgery (EAES) on bariatric surgery: update 2020 endorsed by IFSO-EC, EASO and ESPCOP. *Surgical endoscopy* 2020;34:2332–2358.
- Dietz J, Ulbrich-Kulczynski JM, Souto KE, Meinhardt NG: Prevalence of upper digestive endoscopy and gastric histopathology findings in morbidly obese patients. *Arquivos de gastroenterologia* 2012;49:52–55.
- Dixon MF, Genta RM, Yardley JH, Correa P: Classification and grading of gastritis. The updated Sydney System. International Workshop on the Histopathology of Gastritis, Houston 1994. *The American journal of surgical pathology* 1996;20:1161–1181.
- Dumon KR, Murayama KM: Bariatric surgery outcomes. *The Surgical clinics of North America* 2011;91:1313–1338, x.
- EBMKS: Estonian Society for Metabolic and Bariatric Surgery. Number of bariatric operations. Operation types. Annual report. 2023.
- Eisenberg D, Shikora SA, Aarts E, Aminian A, Angrisani L, Cohen RV, De Luca M, Faria SL, Goodpaster KPS, Haddad A, Himpens JM, Kow L, Kurian M, Loi K, Mahawar K, Nimeri A, O'Kane M, Papasavas PK, Ponce J, Pratt JSA, Rogers AM, Steele KE, Suter M, Kothari SN: 2022 American Society for Metabolic and Bariatric Surgery (ASMBS) and International Federation for the Surgery of Obesity and Metabolic Disorders (IFSO): Indications for Metabolic and Bariatric Surgery. *Surgery for obesity and related diseases: official journal of the American Society for Bariatric Surgery* 2022;18:1345–1356.
- Engevik AC, Kaji I, Goldenring JR: The Physiology of the Gastric Parietal Cell. *Physiological reviews* 2020;100:573–602.
- Escalona A, Guzmán S, Ibáñez L, Meneses L, Huete A, Solar A: Gastric cancer after Roux-en-Y gastric bypass. *Obesity surgery* 2005;15:423–427.

- Farooqi IS: Defining the neural basis of appetite and obesity: from genes to behaviour. *Clinical medicine (London, England)* 2014;14:286–289.
- Farooqi IS, O'Rahilly S: 20 years of leptin: human disorders of leptin action. *The Journal of endocrinology* 2014;223:T63–70.
- Farrell TM, Haggerty SP, Overby DW, Kohn GP, Richardson WS, Fanelli RD: Clinical application of laparoscopic bariatric surgery: an evidence-based review. *Surgical endoscopy* 2009;23:930–949.
- Fernandes SR, Meireles LC, Carrilho-Ribeiro L, Velosa J: The Role of Routine Upper Gastrointestinal Endoscopy Before Bariatric Surgery. *Obesity surgery* 2016;26:2105–2110.
- Flum DR, Khan TV, Dellinger EP: Toward the Rational and Equitable Use of Bariatric Surgery. *Jama* 2007;298:1442–1444.
- Foster GD, Wyatt HR, Hill JO, Makris AP, Rosenbaum DL, Brill C, Stein RI, Mohammed BS, Miller B, Rader DJ, Zemel B, Wadden TA, Tenhave T, Newcomb CW, Klein S: Weight and metabolic outcomes after 2 years on a low-carbohydrate versus low-fat diet: a randomized trial. *Annals of internal medicine* 2010;153:147–157.
- Fried M, Yumuk V, Oppert JM, Scopinaro N, Torres A, Weiner R, Yashkov Y, Frühbeck G: Interdisciplinary European guidelines on metabolic and bariatric surgery. *Obesity surgery* 2014;24:42–55.
- Gehrer S, Kern B, Peters T, Christoffel-Courtin C, Peterli R: Fewer nutrient deficiencies after laparoscopic sleeve gastrectomy (LSG) than after laparoscopic Roux-Y-gastric bypass (LRYGB)-a prospective study. *Obesity surgery* 2010;20:447–453.
- Genco A, Soricelli E, Casella G, Maselli R, Castagneto-Gissey L, Di Lorenzo N, Basso N: Gastroesophageal reflux disease and Barrett's esophagus after laparoscopic sleeve gastrectomy: a possible, underestimated long-term complication. *Surgery for obesity and related diseases: official journal of the American Society for Bariatric Surgery* 2017;13:568–574.
- Gerbens-Leenes PW, Nonhebel S, Krol MS: Food consumption patterns and economic growth. Increasing affluence and the use of natural resources. *Appetite* 2010;55:597–608.
- Gerson LB: Impact of obesity on endoscopy. *Gastrointestinal endoscopy* 2009;70:758–762.
- Giordano S, Victorzon M: Bariatric surgery in elderly patients: a systematic review. *Clinical interventions in aging* 2015;10:1627–1635.
- Gómez V, Bhalla R, Heckman MG, Florit PT, Diehl NN, Rawal B, Lynch SA, Loeb DS: Routine Screening Endoscopy before Bariatric Surgery: Is It Necessary? *Bariatric surgical practice and patient care* 2014;9:143–149.
- Goni E, Riccò M, Franceschi M, Baldassarre G, Panozzo M, Antico A, Bastiani R, Scarpignato C, Mario F: Gastrin 17 As Non Invasive Marker of Reflux Disease. *Gastroenterology* 2015;146:616–617.
- Greenway FL, Whitehouse MJ, Guttadauria M, Anderson JW, Atkinson RL, Fujioka K, Gadde KM, Gupta AK, O'Neil P, Schumacher D, Smith D, Dunayevich E, Tollefson GD, Weber E, Cowley MA: Rational design of a combination medication for the treatment of obesity. *Obesity (Silver Spring, Md)* 2009;17:30–39.
- Gulliford MC, Charlton J, Prevost T, Booth H, Fildes A, Ashworth M, Littlejohns P, Reddy M, Khan O, Rudisill C: Costs and Outcomes of Increasing Access to Bariatric Surgery: Cohort Study and Cost-Effectiveness Analysis Using Electronic

- Health Records. Value in health: the journal of the International Society for Pharmacoeconomics and Outcomes Research 2017;20:85–92.
- Hawasli A, Reyes M, Hare B, Meguid A, Harriott A, Almahmeed T, Thatimatla N, Szpunar S: Can morbidly obese patients with reflux be offered laparoscopic sleeve gastrectomy? A case report of 40 patients. *American journal of surgery* 2016;211: 571–576.
- Kelly AS, Barlow SE, Rao G, Inge TH, Hayman LL, Steinberger J, Urbina EM, Ewing LJ, Daniels SR: Severe obesity in children and adolescents: identification, associated health risks, and treatment approaches: a scientific statement from the American Heart Association. *Circulation* 2013;128:1689–1712.
- Khitin L, Roses RE, Birkett DH: Cancer in the gastric remnant after gastric bypass: a case report. *Current surgery* 2003;60:521–523.
- Kikkas EM, Sillakivi T, Suumann J, Kirsimägi Ü, Tikk T, Värk PR: Five-Year Outcome of Laparoscopic Sleeve Gastrectomy, Resolution of Comorbidities, and Risk for Cumulative Nutritional Deficiencies. *Scandinavian journal of surgery: SJS: official organ for the Finnish Surgical Society and the Scandinavian Surgical Society* 2019;108:10–16.
- Kuga R, Safatle-Ribeiro AV, Faintuch J, Ishida RK, Furuya CK, Jr., Garrido AB, Jr., Ceconello I, Ishioka S, Sakai P: Endoscopic findings in the excluded stomach after Roux-en-Y gastric bypass surgery. *Archives of surgery (Chicago, Ill: 1960)* 2007; 142:942–946.
- Lee PC, Dixon J: Medical devices for the treatment of obesity. *Nature reviews Gastroenterology & hepatology* 2017;14:553–564.
- Lehmann FS, Renner EL, Meyer-Wyss B, Wilder-Smith CH, Mazzucchelli L, Ruchti C, Drewe J, Beglinger C, Merki HS: Helicobacter pylori and gastric erosions. Results of a prevalence study in asymptomatic volunteers. *Digestion* 2000;62:82–86.
- Marchesi F, Tartamella F, De Sario G, Forlini C, Caleffi A, Riccò M, Di Mario F: The Sleeping Remnant. Effect of Roux-En-Y Gastric Bypass on Plasma Levels of Gastric Biomarkers in Morbidly Obese Women: A Prospective Longitudinal Study. *Obesity surgery* 2017;27:1901–1905.
- Martin MJ, Beekley AC, Sebesta JA: Bowel obstruction in bariatric and nonbariatric patients: major differences in management strategies and outcome. *Surgery for obesity and related diseases: official journal of the American Society for Bariatric Surgery* 2011;7:263–269.
- McNickle AG, Bonomo SR: Predictability of first-year weight loss in laparoscopic sleeve gastrectomy. *Surgical endoscopy* 2017;31:4145–4149.
- Mechanick JI, Apovian C, Brethauer S, Timothy Garvey W, Joffe AM, Kim J, Kushner RF, Lindquist R, Pessah-Pollack R, Seger J, Urman RD, Adams S, Cleek JB, Correa R, Figaro MK, Flanders K, Grams J, Hurley DL, Kothari S, Seger MV, Still CD: Clinical Practice Guidelines for the Perioperative Nutrition, Metabolic, and Nonsurgical Support of Patients Undergoing Bariatric Procedures – 2019 Update: Cosponsored by American Association of Clinical Endocrinologists/American College of Endocrinology, The Obesity Society, American Society for Metabolic and Bariatric Surgery, Obesity Medicine Association, and American Society of Anesthesiologists. *Obesity (Silver Spring)* 2020;28:01–058.
- Melissas J, Braghetto I, Molina JC, Silecchia G, Iossa A, Iannelli A, Foletto M: Gastroesophageal Reflux Disease and Sleeve Gastrectomy. *Obesity surgery* 2015;25:2430–2435.

- Monkemuller K, Neumann H, Nocon M, Vieth M, Labenz J, Willich SN, Stolte M, Hocker M, Jaspersen D, Lind T, Malfertheiner P: Serum gastrin and pepsinogens do not correlate with the different grades of severity of gastro-oesophageal reflux disease: a matched case-control study. *Alimentary pharmacology & therapeutics* 2008;28:491–496.
- Montague CT, Farooqi IS, Whitehead JP, Soos MA, Rau H, Wareham NJ, Sewter CP, Digby JE, Mohammed SN, Hurst JA, Cheetham CH, Earley AR, Barnett AH, Prins JB, O'Rahilly S: Congenital leptin deficiency is associated with severe early-onset obesity in humans. *Nature* 1997;387:903–908.
- Mulla CM, Middelbeek RJW, Patti ME: Mechanisms of weight loss and improved metabolism following bariatric surgery. *Annals of the New York Academy of Sciences* 2018;1411:53–64.
- Needleman BJ, Happel LC: Bariatric surgery: choosing the optimal procedure. *The Surgical clinics of North America* 2008;88:991–1007, vi.
- Nomura A, Stemmermann GN, Chyou PH, Kato I, Perez-Perez GI, Blaser MJ: Helicobacter pylori infection and gastric carcinoma among Japanese Americans in Hawaii. *The New England journal of medicine* 1991;325:1132–1136.
- O'Brien PE, Hindle A, Brennan L, Skinner S, Burton P, Smith A, Crosthwaite G, Brown W: Long-Term Outcomes After Bariatric Surgery: a Systematic Review and Meta-analysis of Weight Loss at 10 or More Years for All Bariatric Procedures and a Single-Centre Review of 20-Year Outcomes After Adjustable Gastric Banding. *Obesity surgery* 2019;29:3–14.
- Ogden CL, Carroll MD, Curtin LR, McDowell MA, Tabak CJ, Flegal KM: Prevalence of overweight and obesity in the United States, 1999–2004. *Jama* 2006;295:1549–1555.
- Ogden CL, Carroll MD, Fryar CD, Flegal KM: Prevalence of Obesity Among Adults and Youth: United States, 2011–2014. *NCHS data brief* 2015:1–8.
- Onzi TR, d'Acampora AJ, de Araújo FM, Baratieri R, Kremer G, Lyra HF, Jr., Leitão JT: Gastric histopathology in laparoscopic sleeve gastrectomy: pre- and post-operative comparison. *Obesity surgery* 2014;24:371–376.
- Oona M, Utt M, Nilsson I, Uibo O, Vorobjova T, Maaros HI: Helicobacter pylori infection in children in Estonia: decreasing seroprevalence during the 11-year period of profound socioeconomic changes. *Helicobacter* 2004;9:233–241.
- Pallati PK, Shaligram A, Shostrom VK, Oleynikov D, McBride CL, Goede MR: Improvement in gastroesophageal reflux disease symptoms after various bariatric procedures: review of the Bariatric Outcomes Longitudinal Database. *Surgery for obesity and related diseases: official journal of the American Society for Bariatric Surgery* 2014;10:502–507.
- Papavramidis ST, Theocharidis AJ, Zaraboukas TG, Christoforidou BP, Kessissoglou, II, Aidonopoulos AP: Upper gastrointestinal endoscopic and histologic findings before and after vertical banded gastroplasty. *Surgical endoscopy* 1996;10:825–830.
- Parikh A, Alley JB, Peterson RM, Harnisch MC, Pfluke JM, Tapper DM, Fenton SJ: Management options for symptomatic stenosis after laparoscopic vertical sleeve gastrectomy in the morbidly obese. *Surgical endoscopy* 2012;26:738–746.
- Parikh M, Liu J, Vieira D, Tzimas D, Horwitz D, Antony A, Saunders JK, Ude-Welcome A, Goodman A: Preoperative Endoscopy Prior to Bariatric Surgery: a Systematic Review and Meta-Analysis of the Literature. *Obesity surgery* 2016;26:2961–2966.
- Parkin DM: International variation. *Oncogene* 2004;23:6329–6340.

- Parrott J, Frank L, Rabena R, Craggs-Dino L, Isom KA, Greiman L: American Society for Metabolic and Bariatric Surgery Integrated Health Nutritional Guidelines for the Surgical Weight Loss Patient 2016 Update: Micronutrients. *Surgery for obesity and related diseases: official journal of the American Society for Bariatric Surgery* 2017;13:727–741.
- Peetsalu A, Tamm A, Härkönen M, Varis K, Sipponen P, Karonen SL, Väli T, Villako K: The effect of vagotomy and antrectomy on serum pepsinogens I and II. *Scandinavian journal of gastroenterology* 1990;25:455–461.
- Peetsalu M, Valle J, Härkönen M, Maaros HI, Peetsalu A: Changes in the histology and function of gastric mucosa and in *Helicobacter pylori* colonization during a long-term follow-up period after vagotomy in duodenal ulcer patients. *Hepato-gastroenterology* 2005;52:785–791.
- Peitz U, Wex T, Vieth M, Stolte M, Willich S, Labenz J, Jaspersen D, Lind T, Malfertheiner P: Correlation of serum pepsinogens and gastrin-17 with atrophic gastritis in gastroesophageal reflux patients: a matched-pairs study. *Journal of gastroenterology and hepatology* 2011;26:82–89.
- Peleteiro B, Bastos A, Ferro A, Lunet N: Prevalence of *Helicobacter pylori* infection worldwide: a systematic review of studies with national coverage. *Digestive diseases and sciences* 2014;59:1698–1709.
- Peromaa-Haavisto P, Victorzon M: Is routine preoperative upper GI endoscopy needed prior to gastric bypass? *Obesity surgery* 2013;23:736–739.
- Perrone F, Bianciardi E, Ippoliti S, Nardella J, Fabi F, Gentileschi P: Long-term effects of laparoscopic sleeve gastrectomy versus Roux-en-Y gastric bypass for the treatment of morbid obesity: a monocentric prospective study with minimum follow-up of 5 years. *Updates in surgery* 2017;69:101–107.
- Peterli R, Wölnerhanssen BK, Peters T, Vetter D, Kröll D, Borbély Y, Schultes B, Beglinger C, Drewe J, Schiesser M, Nett P, Bueter M: Effect of Laparoscopic Sleeve Gastrectomy vs Laparoscopic Roux-en-Y Gastric Bypass on Weight Loss in Patients With Morbid Obesity: The SM-BOSS Randomized Clinical Trial. *Jama* 2018;319:255–265.
- Podnos YD, Jimenez JC, Wilson SE, Stevens CM, Nguyen NT: Complications after laparoscopic gastric bypass: a review of 3464 cases. *Archives of surgery (Chicago, Ill: 1960)* 2003;138:957–961.
- Ridaura VK, Faith JJ, Rey FE, Cheng J, Duncan AE, Kau AL, Griffin NW, Lombard V, Henrissat B, Bain JR, Muehlbauer MJ, Ilkayeva O, Semenkovich CF, Funai K, Hayashi DK, Lyle BJ, Martini MC, Ursell LK, Clemente JC, Van Treuren W, Walters WA, Knight R, Newgard CB, Heath AC, Gordon JI: Gut microbiota from twins discordant for obesity modulate metabolism in mice. *Science (New York, NY)* 2013;341:1241214.
- Rothman RB, Baumann MH, Dersch CM, Romero DV, Rice KC, Carroll FI, Partilla JS: Amphetamine-type central nervous system stimulants release norepinephrine more potently than they release dopamine and serotonin. *Synapse (New York, NY)* 2001;39:32–41.
- Sacks BC, Mattar SG, Qureshi FG, Eid GM, Collins JL, Barinas-Mitchell EJ, Schauer PR, Ramanathan RC: Incidence of marginal ulcers and the use of absorbable anastomotic sutures in laparoscopic Roux-en-Y gastric bypass. *Surgery for obesity and related diseases: official journal of the American Society for Bariatric Surgery* 2006;2:11–16.

- Safatle-Ribeiro AV, Kuga R, Iriya K, Ribeiro U, Jr., Faintuch J, Ishida RK, Corbett CE, Garrido AB, Jr., Ishioka S, Sakai P: What to expect in the excluded stomach mucosa after vertical banded Roux-en-Y gastric bypass for morbid obesity. *Journal of gastrointestinal surgery: official journal of the Society for Surgery of the Alimentary Tract* 2007;11:133–137.
- Safatle-Ribeiro AV, Petersen PA, Pereira Filho DS, Corbett CE, Faintuch J, Ishida R, Sakai P, Cecconello I, Ribeiro U, Jr.: Epithelial cell turnover is increased in the excluded stomach mucosa after Roux-en-Y gastric bypass for morbid obesity. *Obesity surgery* 2013;23:1616–1623.
- Salminen P, Grönroos S, Helmiö M, Hurme S, Juuti A, Juusela R, Peromaa-Haavisto P, Leivonen M, Nuutila P, Ovaska J: Effect of Laparoscopic Sleeve Gastrectomy vs Roux-en-Y Gastric Bypass on Weight Loss, Comorbidities, and Reflux at 10 Years in Adult Patients With Obesity: The SLEEVEPASS Randomized Clinical Trial. *JAMA surgery* 2022;157:656–666.
- Schubert ML: Gastric secretion. *Current opinion in gastroenterology* 2014;30:578–582.
- Schubert ML: Functional anatomy and physiology of gastric secretion. *Current opinion in gastroenterology* 2015;31:479–485.
- Sheppard CE, Sadowski DC, de Gara CJ, Karmali S, Birch DW: Rates of reflux before and after laparoscopic sleeve gastrectomy for severe obesity. *Obesity surgery* 2015; 25:763–768.
- Shrestha R, Koirala K, Raj KC, Batajoo KH: Helicobacter pylori infection among patients with upper gastrointestinal symptoms: prevalence and relation to endoscopy diagnosis and histopathology. *Journal of family medicine and primary care* 2014; 3:154–158.
- Sillakivi T, Suumann J, Kirsimägi U, Peetsalu A: Plasma levels of gastric biomarkers in patients after bariatric surgery: biomarkers after bariatric surgery. *Hepato-gastroenterology* 2013;60:2129–2132.
- Sipponen P, Graham DY: Importance of atrophic gastritis in diagnostics and prevention of gastric cancer: application of plasma biomarkers. *Scandinavian journal of gastroenterology* 2007;42:2–10.
- Sipponen P, Hyvärinen H: Role of Helicobacter pylori in the pathogenesis of gastritis, peptic ulcer and gastric cancer. *Scandinavian journal of gastroenterology Supplement* 1993;196:3–6.
- Sipponen P, Laxén F, Huotari K, Härkönen M: Prevalence of low vitamin B12 and high homocysteine in serum in an elderly male population: association with atrophic gastritis and Helicobacter pylori infection. *Scandinavian journal of gastroenterology* 2003;38:1209–1216.
- Sipponen P, Vauhkonen M, Helske T, Kaariainen I, Harkonen M: Low circulating levels of gastrin-17 in patients with Barrett's esophagus. *World journal of gastroenterology* 2005;11:5988–5992.
- Sjöström L: Review of the key results from the Swedish Obese Subjects (SOS) trial – a prospective controlled intervention study of bariatric surgery. *Journal of internal medicine* 2013;273:219–234.
- Sjöström L, Lindroos AK, Peltonen M, Torgerson J, Bouchard C, Carlsson B, Dahlgren S, Larsson B, Narbro K, Sjöström CD, Sullivan M, Wedel H: Lifestyle, diabetes, and cardiovascular risk factors 10 years after bariatric surgery. *The New England journal of medicine* 2004;351:2683–2693.
- Sjöström L, Narbro K, Sjöström CD, Karason K, Larsson B, Wedel H, Lystig T, Sullivan M, Bouchard C, Carlsson B, Bengtsson C, Dahlgren S, Gummesson A,

- Jacobson P, Karlsson J, Lindroos AK, Lönroth H, Näslund I, Olbers T, Stenlöf K, Torgerson J, Agren G, Carlsson LM: Effects of bariatric surgery on mortality in Swedish obese subjects. *The New England journal of medicine* 2007;357:741–752.
- Smith I, Hardy E, Mitchell S, Batson S: Semaglutide 2.4 Mg for the Management of Overweight and Obesity: Systematic Literature Review and Meta-Analysis. *Diabetes, metabolic syndrome and obesity: targets and therapy* 2022;15:3961–3987.
- Soybel DI: Anatomy and physiology of the stomach. *The Surgical clinics of North America* 2005;85:875–894, v.
- Speakman JR, Levitsky DA, Allison DB, Bray MS, de Castro JM, Clegg DJ, Clapham JC, Dulloo AG, Gruer L, Haw S, Hebebrand J, Hetherington MM, Higgs S, Jebb SA, Loos RJ, Luckman S, Luke A, Mohammed-Ali V, O'Rahilly S, Pereira M, Perusse L, Robinson TN, Rolls B, Symonds ME, Westerterp-Plantenga MS: Set points, settling points and some alternative models: theoretical options to understand how genes and environments combine to regulate body adiposity. *Disease models & mechanisms* 2011;4:733–745.
- Srivastava G, Apovian CM: Current pharmacotherapy for obesity. *Nature reviews Endocrinology* 2018;14:12–24.
- Stenard F, Iannelli A: Laparoscopic sleeve gastrectomy and gastroesophageal reflux. *World journal of gastroenterology* 2015;21:10348–10357.
- Stolte M, Eidt S, Ritter M, Bethke B: [Campylobacter pylori and gastritis. Association or induction?]. *Der Pathologe* 1989;10:21–26.
- Storskrubb T, Aro P, Ronkainen J, Sipponen P, Nyhlin H, Talley NJ, Engstrand L, Stolte M, Vieth M, Walker M, Agréus L: Serum biomarkers provide an accurate method for diagnosis of atrophic gastritis in a general population: The Kalixanda study. *Scandinavian journal of gastroenterology* 2008;43:1448–1455.
- Sucandy I, Chrestiana D, Bonanni F, Antanavicius G: Gastroesophageal Reflux Symptoms After Laparoscopic Sleeve Gastrectomy for Morbid Obesity. The Importance of Preoperative Evaluation and Selection. *North American journal of medical sciences* 2015;7:189–193.
- Sundbom M, Holdstock C, Engström BE, Karlsson FA: Early changes in ghrelin following Roux-en-Y gastric bypass: influence of vagal nerve functionality? *Obesity surgery* 2007;17:304–310.
- Sundbom M, Mårdh E, Mårdh S, Ohrvall M, Gustavsson S: Reduction in serum pepsinogen I after Roux-en-Y gastric bypass. *Journal of gastrointestinal surgery: official journal of the Society for Surgery of the Alimentary Tract* 2003;7:529–535.
- Syrjänen K: A Panel of Serum Biomarkers (GastroPanel®) in Non-invasive Diagnosis of Atrophic Gastritis. Systematic Review and Meta-analysis. *Anticancer research* 2016;36:5133–5144.
- Syrjänen K, Eskelinen M, Peetsalu A, Sillakivi T, Sipponen P, Härkönen M, Paloheimo L, Mäki M, Tiusanen T, Suovaniemi O, Di MF, Fan ZP: GastroPanel® Biomarker Assay: The Most Comprehensive Test for Helicobacter pylori Infection and Its Clinical Sequelae. A Critical Review. *Anticancer research* 2019;39:1091–1104.
- Zhang Y, Proenca R, Maffei M, Barone M, Leopold L, Friedman JM: Positional cloning of the mouse obese gene and its human homologue. *Nature* 1994;372:425–432.
- Zhou LY, Shen ZY, Lin SR, Jin Z, Ding SG, Huang XB, Xia ZW, Liu JJ, Guo HL, William C: [Changes of gastric mucosa histopathology after Helicobacter pylori eradication]. *Zhonghua nei ke za zhi* 2003;42:162–164.

- TAI: National institute of health development. Overweight and obese adolescents. [https://statistika.tai.ee/pxweb/et/Andmebaas/Andmebaas\\_\\_05Uuringud\\_\\_03HBSC\\_\\_03Kehaline\\_aktiivsus/KU32.px/table/tableViewLayout2/](https://statistika.tai.ee/pxweb/et/Andmebaas/Andmebaas__05Uuringud__03HBSC__03Kehaline_aktiivsus/KU32.px/table/tableViewLayout2/). 2019.
- TAI: National institute of health development. Overweight and obese adults. [https://statistika.tai.ee/pxweb/et/Andmebaas/Andmebaas\\_\\_05Uuringud\\_\\_02TKU\\_\\_04Liikumine/TKU40.px/table/tableViewLayout2/](https://statistika.tai.ee/pxweb/et/Andmebaas/Andmebaas__05Uuringud__02TKU__04Liikumine/TKU40.px/table/tableViewLayout2/) 2021.
- Tak YJ, Lee SY: Anti-Obesity Drugs: Long-Term Efficacy and Safety: An Updated Review. *The world journal of men's health* 2021;39:208–221.
- Telaranta-Keerie A, Kara R, Paloheimo L, Härkönen M, Sipponen P: Prevalence of undiagnosed advanced atrophic corpus gastritis in Finland: an observational study among 4,256 volunteers without specific complaints. *Scandinavian journal of gastroenterology* 2010;45:1036–1041.
- Turnbaugh PJ, Hamady M, Yatsunenko T, Cantarel BL, Duncan A, Ley RE, Sogin ML, Jones WJ, Roe BA, Affourtit JP, Egholm M, Henrissat B, Heath AC, Knight R, Gordon JI: A core gut microbiome in obese and lean twins. *Nature* 2009;457:480–484.
- Wadden TA, Sternberg JA, Letizia KA, Stunkard AJ, Foster GD: Treatment of obesity by very low calorie diet, behavior therapy, and their combination: a five-year perspective. *International journal of obesity* 1989;13 Suppl 2:39–46.
- van Can J, Sloth B, Jensen CB, Flint A, Blaak EE, Saris WH: Effects of the once-daily GLP-1 analog liraglutide on gastric emptying, glycemic parameters, appetite and energy metabolism in obese, non-diabetic adults. *International journal of obesity (2005)* 2014;38:784–793.
- Vandevijvere S, Chow CC, Hall KD, Umali E, Swinburn BA: Increased food energy supply as a major driver of the obesity epidemic: a global analysis. *Bulletin of the World Health Organization* 2015;93:446–456.
- Waters H GM: America`s Obesity Crisis. [https://milkeninstitute.org/sites/default/files/reports-pdf/Mi-Americas-Obesity-Crisis-WEB\\_2.pdf](https://milkeninstitute.org/sites/default/files/reports-pdf/Mi-Americas-Obesity-Crisis-WEB_2.pdf). Accessed 12 Apr. 2018.
- Weiner RA, Weiner S, Pomhoff I, Jacobi C, Makarewicz W, Weigand G: Laparoscopic sleeve gastrectomy--influence of sleeve size and resected gastric volume. *Obesity surgery* 2007;17:1297–1305.
- Velazquez A, Apovian CM: Updates on obesity pharmacotherapy. *Annals of the New York Academy of Sciences* 2018;1411:106–119.
- Williams EP, Mesidor M, Winters K, Dubbert PM, Wyatt SB: Overweight and Obesity: Prevalence, Consequences, and Causes of a Growing Public Health Problem. *Current obesity reports* 2015;4:363–370.
- World Health Organisation (WHO). (2021): Obesity and overweight. Available from <https://www.who.int/mediacentre/factsheets/fs311/en/>. Accessed 12 Apr 2023.
- Vorobjova T, Grünberg H, Oona M, Maaros HI, Nilsson I, Wadström T, Covacci A, Uibo R: Seropositivity to *Helicobacter pylori* and CagA protein in schoolchildren of different ages living in urban and rural areas in southern Estonia. *European journal of gastroenterology & hepatology* 2000;12:97–101.

## ACKNOWLEDGMENTS

I express my sincere gratitude to:

- My supervisor Ass. Prof. Toomas Sillakivi for his mentorship and also for showing me the beauty of laparoscopic surgery. I appreciate his support and patience during my PhD studies
- My supervisor Prof. Ants Peetsalu for offering me the opportunity to enter the world of science and for his support during my PhD studies
- My co-authors Dr. Zivile Riispere, Prof. Pentti Sipponen and Prof. Kari Syrjänen for performing histological evaluation of the gastric biopsies and for reviewing my publications
- Prof. Jaak Kals and Prof. Aare Märtson for critically reviewing and improving this thesis
- Prof. Osmo Suovoniemi for his help and support in this project
- Endoscopy unit, especially Ms. Evelin Arak for assisting with the endoscopies
- Dr. Ceith Nikkolo for comprehensive help whenever needed and for being the first reader to improve my thesis
- Mrs. Ülle Kirsimägi for enormous help with the statistics
- Mrs. Ester Jaigma for reading and improving the English text of my manuscripts
- Dr. Tatjana Veršinina for preparing the excellent illustrations for this thesis
- Staff members of the Department of General and Plastic Surgery and Department of Abdominal Surgery for being wonderful colleagues for me
- All patients who participated in this study and made this scientific project possible
- My family: my awesome wife Janelle who has supported me in everything and my children Tobias, Britta and Bastian – you are the best!



## **PUBLICATIONS**

## CURRICULUM VITAE

**Name:** Jaanus Suumann  
**Date of birth:** April 11, 1983  
**Citizenship:** Estonian  
**Phone:** +372 5331 9084; +372 731 8242  
**E-mail:** jaanus.suumann@kliinikum.ee

### Education:

2013–2023 University of Tartu, Faculty of Medicine, PhD studies  
2008–2013 University of Tartu, Faculty of Medicine,  
Residency in general surgery  
2002–2008 University of Tartu, Faculty of Medicine, degree of MD  
1990–2002 Miina Härma Gymnasium

### Professional employment:

2013– Tartu University Hospital, Surgery Clinic,  
Department of General Surgery and Plastic Surgery,  
General surgeon  
2008–2013 Tartu University Hospital, Surgery Clinic, Department of  
Abdominal Surgery; Department of General Surgery and Plastic  
Surgery, resident in general surgery  
2005–2007 Tartu University Hospital, Surgery Clinic, Department of Uro-  
logy and Kidney Transplantation, nurse

### Scientific work and professional organisations:

Research fields: bariatric surgery, endocrine surgery

Membership: Tartu Surgeons' Association  
Estonian Society for Bariatric and Metabolic Surgery

### Publications:

- 1) Sillakivi T, **Suumann J**, Kirsimägi U, Peetsalu A. Plasma levels of gastric biomarkers in patients after bariatric surgery: biomarker after bariatric surgery. *Hepatology*. 2013 Nov-Dec;60(128):2129–32.
- 2) Nikkolo C, Vaasna T, Murruste M, Seepter H, **Suumann J**, Tein A, Kirsimägi Ü, Lepner U. Single-center, single-blinded, randomized study of self-gripping versus sutured mesh in open inguinal hernia repair. *J Surg Res*. 2015 Mar;194(1):77–82.
- 3) Nikkolo C, Vaasna T, Murruste M, **Suumann J**, Kirsimägi Ü, Seepter H, Tein A, Lepner U. Three-year results of a randomized study comparing self-gripping mesh with sutured mesh in open inguinal hernia repair. *J Surg Res*. 2017 Mar;209:139–144.
- 4) Nikkolo C, Kirsimägi Ü, Vaasna T, Murruste M, **Suumann J**, Seepter H, Lepner U. Prospective study evaluating the impact of severity of chronic

- pain on quality of life after inguinal hernioplasty. *Hernia*. 2017 Apr;21(2): 199–205.
- 5) **Suumann J**, Sillakivi T, Riispere Ž, Syrjänen K, Sipponen P, Kirsimägi Ü, Peetsalu A. Serological biomarker testing helps avoiding unnecessary endoscopies in obese patients before bariatric surgery. *BMC Obes*. 2018 Feb 20;5:9.
  - 6) Kikkas EM, Sillakivi T, **Suumann J**, Kirsimägi Ü, Tikk T, Värk PR. Five-Year Outcome of Laparoscopic Sleeve Gastrectomy, Resolution of Comorbidities, and Risk for Cumulative Nutritional Deficiencies. *Scand J Surg*. 2018 Jul 1:1457496918783723.
  - 7) **Suumann J**, Sillakivi T, Riispere Ž, Syrjänen K, Sipponen P, Kirsimägi Ü, Peetsalu A. Serum gastric biomarker levels after sleeve gastrectomy and Roux-en-Y gastric bypass operations: a prospective study with 1-year follow-up. *Surgery Gastroenterology and Oncology*. 2019 Apr 24(2):86

## ELULOOKIRJELDUS

**Nimi:** Jaanus Suumann  
**Sünniaeg:** 11. aprill 1983  
**Kodakondsus:** Eesti  
**Telefon:** +372 5331 9084; +372 731 8242  
**E-mail:** jaanus.suumann@kliinikum.ee

### Hariduskäik:

2013–2023 Tartu Ülikool, Arstiteaduskond, arstiteaduse doktoriõpe  
2008–2013 Tartu Ülikool, Arstiteaduskond, arst-resident üldkirurgia erialal  
2002–2008 Tartu Ülikool, Arstiteaduskond, arstiteaduse põhiõpe  
1990–2002 Miina Härma Gümnaasium

### Teenistuskäik:

2013– SA Tartu Ülikooli Kliinikum, Kirurgiakliinik, üldkirurgia ja plastikakirurgia osakond, arst-õppejõud üldkirurgia erialal  
2008–2013 SA Tartu Ülikooli Kliinikum, Kirurgiakliinik, abdominaalkirurgia osakond; üldkirurgia ja plastikakirurgia osakond, arst-resident üldkirurgia erialal  
2005–2007 SA Tartu Ülikooli Kliinikum, Kirurgiakliinik, uroloogia ja neerusiirdamise osakond, õde

### Teadus- ja erialane tegevus:

Valdkonnad: bariatriline kirurgia, endokriinkirurgia  
Liikmelisus: Eesti Kirurgide Assotsiatsioon  
Tartu Kirurgide Selts

### Publikatsioonid:

- 1) Sillakivi T, **Suumann J**, Kirsimägi U, Peetsalu A. Plasma levels of gastric biomarkers in patients after bariatric surgery: biomarker after bariatric surgery. *Hepato-gastroenterology*. 2013 Nov-Dec;60(128):2129–32.
- 2) Nikkolo C, Vaasna T, Murruste M, Seepter H, **Suumann J**, Tein A, Kirsimägi Ü, Lepner U. Single-center, single-blinded, randomized study of self-gripping versus sutured mesh in open inguinal hernia repair. *J Surg Res*. 2015 Mar;194(1):77–82.
- 3) Nikkolo C, Vaasna T, Murruste M, **Suumann J**, Kirsimägi Ü, Seepter H, Tein A, Lepner U. Three-year results of a randomized study comparing self-gripping mesh with sutured mesh in open inguinal hernia repair. *J Surg Res*. 2017 Mar;209:139–144.
- 4) Nikkolo C, Kirsimägi Ü, Vaasna T, Murruste M, **Suumann J**, Seepter H, Lepner U. Prospective study evaluating the impact of severity of chronic

- pain on quality of life after inguinal hernioplasty. *Hernia*. 2017 Apr;21(2): 199–205.
- 5) **Suumann J**, Sillakivi T, Riispere Ž, Syrjänen K, Sipponen P, Kirsimägi Ü, Peetsalu A. Serological biomarker testing helps avoiding unnecessary endoscopies in obese patients before bariatric surgery. *BMC Obes*. 2018 Feb 20;5:9.
  - 6) Kikkas EM, Sillakivi T, **Suumann J**, Kirsimägi Ü, Tikk T, Värk PR. Five-Year Outcome of Laparoscopic Sleeve Gastrectomy, Resolution of Comorbidities, and Risk for Cumulative Nutritional Deficiencies. *Scand J Surg*. 2018 Jul 1:1457496918783723.
  - 7) **Suumann J**, Sillakivi T, Riispere Ž, Syrjänen K, Sipponen P, Kirsimägi Ü, Peetsalu A. Serum gastric biomarker levels after sleeve gastrectomy and Roux-en-Y gastric bypass operations: a prospective study with 1-year follow-up. *Surgery Gastroenterology and Oncology*. 2019 Apr 24(2):86

## DISSERTATIONES MEDICINAE UNIVERSITATIS TARTUENSIS

1. **Heidi-Ingrid Maaros.** The natural course of gastric ulcer in connection with chronic gastritis and *Helicobacter pylori*. Tartu, 1991.
2. **Mihkel Zilmer.** Na-pump in normal and tumorous brain tissues: Structural, functional and tumorigenesis aspects. Tartu, 1991.
3. **Eero Vasar.** Role of cholecystokinin receptors in the regulation of behaviour and in the action of haloperidol and diazepam. Tartu, 1992.
4. **Tiina Talvik.** Hypoxic-ischaemic brain damage in neonates (clinical, biochemical and brain computed tomographical investigation). Tartu, 1992.
5. **Ants Peetsalu.** Vagotomy in duodenal ulcer disease: A study of gastric acidity, serum pepsinogen I, gastric mucosal histology and *Helicobacter pylori*. Tartu, 1992.
6. **Marika Mikelsaar.** Evaluation of the gastrointestinal microbial ecosystem in health and disease. Tartu, 1992.
7. **Hele Everaus.** Immuno-hormonal interactions in chronic lymphocytic leukaemia and multiple myeloma. Tartu, 1993.
8. **Ruth Mikelsaar.** Etiological factors of diseases in genetically consulted children and newborn screening: dissertation for the commencement of the degree of doctor of medical sciences. Tartu, 1993.
9. **Agu Tamm.** On metabolic action of intestinal microflora: clinical aspects. Tartu, 1993.
10. **Katrin Gross.** Multiple sclerosis in South-Estonia (epidemiological and computed tomographical investigations). Tartu, 1993.
11. **Oivi Uiibo.** Childhood coeliac disease in Estonia: occurrence, screening, diagnosis and clinical characterization. Tartu, 1994.
12. **Viiu Tuulik.** The functional disorders of central nervous system of chemistry workers. Tartu, 1994.
13. **Margus Viigimaa.** Primary haemostasis, antiaggregative and anticoagulant treatment of acute myocardial infarction. Tartu, 1994.
14. **Rein Kolk.** Atrial versus ventricular pacing in patients with sick sinus syndrome. Tartu, 1994.
15. **Toomas Podar.** Incidence of childhood onset type 1 diabetes mellitus in Estonia. Tartu, 1994.
16. **Kiira Subi.** The laboratory surveillance of the acute respiratory viral infections in Estonia. Tartu, 1995.
17. **Irja Lutsar.** Infections of the central nervous system in children (epidemiologic, diagnostic and therapeutic aspects, long term outcome). Tartu, 1995.
18. **Aavo Lang.** The role of dopamine, 5-hydroxytryptamine, sigma and NMDA receptors in the action of antipsychotic drugs. Tartu, 1995.
19. **Andrus Arak.** Factors influencing the survival of patients after radical surgery for gastric cancer. Tartu, 1996.

20. **Tõnis Karki.** Quantitative composition of the human lactoflora and method for its examination. Tartu, 1996.
21. **Reet Mändar.** Vaginal microflora during pregnancy and its transmission to newborn. Tartu, 1996.
22. **Triin Remmel.** Primary biliary cirrhosis in Estonia: epidemiology, clinical characterization and prognostication of the course of the disease. Tartu, 1996.
23. **Toomas Kivastik.** Mechanisms of drug addiction: focus on positive reinforcing properties of morphine. Tartu, 1996.
24. **Paavo Pokk.** Stress due to sleep deprivation: focus on GABA<sub>A</sub> receptor-chloride ionophore complex. Tartu, 1996.
25. **Kristina Allikmets.** Renin system activity in essential hypertension. Associations with atherothrombogenic cardiovascular risk factors and with the efficacy of calcium antagonist treatment. Tartu, 1996.
26. **Triin Parik.** Oxidative stress in essential hypertension: Associations with metabolic disturbances and the effects of calcium antagonist treatment. Tartu, 1996.
27. **Svetlana Päi.** Factors promoting heterogeneity of the course of rheumatoid arthritis. Tartu, 1997.
28. **Maarika Sallo.** Studies on habitual physical activity and aerobic fitness in 4 to 10 years old children. Tartu, 1997.
29. **Paul Naaber.** *Clostridium difficile* infection and intestinal microbial ecology. Tartu, 1997.
30. **Rein Pähkla.** Studies in pinoline pharmacology. Tartu, 1997.
31. **Andrus Juhan Voitk.** Outpatient laparoscopic cholecystectomy. Tartu, 1997.
32. **Joel Starkopf.** Oxidative stress and ischaemia-reperfusion of the heart. Tartu, 1997.
33. **Janika Kõrv.** Incidence, case-fatality and outcome of stroke. Tartu, 1998.
34. **Ülla Linnamägi.** Changes in local cerebral blood flow and lipid peroxidation following lead exposure in experiment. Tartu, 1998.
35. **Ave Minajeva.** Sarcoplasmic reticulum function: comparison of atrial and ventricular myocardium. Tartu, 1998.
36. **Oleg Milenin.** Reconstruction of cervical part of esophagus by revascularised ileal autografts in dogs. A new complex multistage method. Tartu, 1998.
37. **Sergei Pakriev.** Prevalence of depression, harmful use of alcohol and alcohol dependence among rural population in Udmurtia. Tartu, 1998.
38. **Allen Kaasik.** Thyroid hormone control over  $\beta$ -adrenergic signalling system in rat atria. Tartu, 1998.
39. **Vallo Matto.** Pharmacological studies on anxiogenic and antiaggressive properties of antidepressants. Tartu, 1998.
40. **Maire Vasar.** Allergic diseases and bronchial hyperreactivity in Estonian children in relation to environmental influences. Tartu, 1998.
41. **Kaja Julge.** Humoral immune responses to allergens in early childhood. Tartu, 1998.

42. **Heli Grünberg.** The cardiovascular risk of Estonian schoolchildren. A cross-sectional study of 9-, 12- and 15-year-old children. Tartu, 1998.
43. **Epp Sepp.** Formation of intestinal microbial ecosystem in children. Tartu, 1998.
44. **Mai Ots.** Characteristics of the progression of human and experimental glomerulopathies. Tartu, 1998.
45. **Tiina Ristimäe.** Heart rate variability in patients with coronary artery disease. Tartu, 1998.
46. **Leho Kõiv.** Reaction of the sympatho-adrenal and hypothalamo-pituitary-adrenocortical system in the acute stage of head injury. Tartu, 1998.
47. **Bela Adojaan.** Immune and genetic factors of childhood onset IDDM in Estonia. An epidemiological study. Tartu, 1999.
48. **Jakov Shlik.** Psychophysiological effects of cholecystokinin in humans. Tartu, 1999.
49. **Kai Kisand.** Autoantibodies against dehydrogenases of  $\alpha$ -ketoacids. Tartu, 1999.
50. **Toomas Marandi.** Drug treatment of depression in Estonia. Tartu, 1999.
51. **Ants Kask.** Behavioural studies on neuropeptide Y. Tartu, 1999.
52. **Ello-Rahel Karelson.** Modulation of adenylate cyclase activity in the rat hippocampus by neuropeptide galanin and its chimeric analogs. Tartu, 1999.
53. **Tanel Laisaar.** Treatment of pleural empyema — special reference to intrapleural therapy with streptokinase and surgical treatment modalities. Tartu, 1999.
54. **Eve Pihl.** Cardiovascular risk factors in middle-aged former athletes. Tartu, 1999.
55. **Katrin Õunap.** Phenylketonuria in Estonia: incidence, newborn screening, diagnosis, clinical characterization and genotype/phenotype correlation. Tartu, 1999.
56. **Siiri Kõljalg.** *Acinetobacter* – an important nosocomial pathogen. Tartu, 1999.
57. **Helle Karro.** Reproductive health and pregnancy outcome in Estonia: association with different factors. Tartu, 1999.
58. **Heili Varendi.** Behavioral effects observed in human newborns during exposure to naturally occurring odors. Tartu, 1999.
59. **Anneli Beilmann.** Epidemiology of epilepsy in children and adolescents in Estonia. Prevalence, incidence, and clinical characteristics. Tartu, 1999.
60. **Vallo Volke.** Pharmacological and biochemical studies on nitric oxide in the regulation of behaviour. Tartu, 1999.
61. **Pilvi Ilves.** Hypoxic-ischaemic encephalopathy in asphyxiated term infants. A prospective clinical, biochemical, ultrasonographical study. Tartu, 1999.
62. **Anti Kalda.** Oxygen-glucose deprivation-induced neuronal death and its pharmacological prevention in cerebellar granule cells. Tartu, 1999.
63. **Eve-Irene Lepist.** Oral peptide prodrugs – studies on stability and absorption. Tartu, 2000.

64. **Jana Kivastik.** Lung function in Estonian schoolchildren: relationship with anthropometric indices and respiratory symptoms, reference values for dynamic spirometry. Tartu, 2000.
65. **Karin Kull.** Inflammatory bowel disease: an immunogenetic study. Tartu, 2000.
66. **Kaire Innos.** Epidemiological resources in Estonia: data sources, their quality and feasibility of cohort studies. Tartu, 2000.
67. **Tamara Vorobjova.** Immune response to *Helicobacter pylori* and its association with dynamics of chronic gastritis and epithelial cell turnover in antrum and corpus. Tartu, 2001.
68. **Ruth Kalda.** Structure and outcome of family practice quality in the changing health care system of Estonia. Tartu, 2001.
69. **Annika Krüüner.** *Mycobacterium tuberculosis* – spread and drug resistance in Estonia. Tartu, 2001.
70. **Marlit Veldi.** Obstructive Sleep Apnoea: Computerized Endopharyngeal Myotonometry of the Soft Palate and Lingual Musculature. Tartu, 2001.
71. **Anneli Uusküla.** Epidemiology of sexually transmitted diseases in Estonia in 1990–2000. Tartu, 2001.
72. **Ade Kallas.** Characterization of antibodies to coagulation factor VIII. Tartu, 2002.
73. **Heidi Annuk.** Selection of medicinal plants and intestinal lactobacilli as antimicrobial components for functional foods. Tartu, 2002.
74. **Aet Lukmann.** Early rehabilitation of patients with ischaemic heart disease after surgical revascularization of the myocardium: assessment of health-related quality of life, cardiopulmonary reserve and oxidative stress. A clinical study. Tartu, 2002.
75. **Maigi Eisen.** Pathogenesis of Contact Dermatitis: participation of Oxidative Stress. A clinical – biochemical study. Tartu, 2002.
76. **Piret Hussar.** Histology of the post-traumatic bone repair in rats. Elaboration and use of a new standardized experimental model – bicortical perforation of tibia compared to internal fracture and resection osteotomy. Tartu, 2002.
77. **Tõnu Rätsep.** Aneurysmal subarachnoid haemorrhage: Noninvasive monitoring of cerebral haemodynamics. Tartu, 2002.
78. **Marju Herodes.** Quality of life of people with epilepsy in Estonia. Tartu, 2003.
79. **Katre Maasalu.** Changes in bone quality due to age and genetic disorders and their clinical expressions in Estonia. Tartu, 2003.
80. **Toomas Sillakivi.** Perforated peptic ulcer in Estonia: epidemiology, risk factors and relations with *Helicobacter pylori*. Tartu, 2003.
81. **Leena Puksa.** Late responses in motor nerve conduction studies. F and A waves in normal subjects and patients with neuropathies. Tartu, 2003.
82. **Krista Lõivukene.** *Helicobacter pylori* in gastric microbial ecology and its antimicrobial susceptibility pattern. Tartu, 2003.

83. **Helgi Kolk.** Dyspepsia and *Helicobacter pylori* infection: the diagnostic value of symptoms, treatment and follow-up of patients referred for upper gastrointestinal endoscopy by family physicians. Tartu, 2003.
84. **Helena Soomer.** Validation of identification and age estimation methods in forensic odontology. Tartu, 2003.
85. **Kersti Oselin.** Studies on the human MDR1, MRP1, and MRP2 ABC transporters: functional relevance of the genetic polymorphisms in the *MDR1* and *MRP1* gene. Tartu, 2003.
86. **Jaan Soplepmann.** Peptic ulcer haemorrhage in Estonia: epidemiology, prognostic factors, treatment and outcome. Tartu, 2003.
87. **Margot Peetsalu.** Long-term follow-up after vagotomy in duodenal ulcer disease: recurrent ulcer, changes in the function, morphology and *Helicobacter pylori* colonisation of the gastric mucosa. Tartu, 2003.
88. **Kersti Klaamas.** Humoral immune response to *Helicobacter pylori* a study of host-dependent and microbial factors. Tartu, 2003.
89. **Pille Taba.** Epidemiology of Parkinson's disease in Tartu, Estonia. Prevalence, incidence, clinical characteristics, and pharmacoepidemiology. Tartu, 2003.
90. **Alar Veraksitš.** Characterization of behavioural and biochemical phenotype of cholecystikinin-2 receptor deficient mice: changes in the function of the dopamine and endopioidergic system. Tartu, 2003.
91. **Ingrid Kalev.** CC-chemokine receptor 5 (CCR5) gene polymorphism in Estonians and in patients with Type I and Type II diabetes mellitus. Tartu, 2003.
92. **Lumme Kadaja.** Molecular approach to the regulation of mitochondrial function in oxidative muscle cells. Tartu, 2003.
93. **Aive Liigant.** Epidemiology of primary central nervous system tumours in Estonia from 1986 to 1996. Clinical characteristics, incidence, survival and prognostic factors. Tartu, 2004.
94. **Andres, Kulla.** Molecular characteristics of mesenchymal stroma in human astrocytic gliomas. Tartu, 2004.
95. **Mari Järvelaid.** Health damaging risk behaviours in adolescence. Tartu, 2004.
96. **Ülle Pechter.** Progression prevention strategies in chronic renal failure and hypertension. An experimental and clinical study. Tartu, 2004.
97. **Gunnar Tasa.** Polymorphic glutathione S-transferases – biology and role in modifying genetic susceptibility to senile cataract and primary open angle glaucoma. Tartu, 2004.
98. **Tuuli Käämbre.** Intracellular energetic unit: structural and functional aspects. Tartu, 2004.
99. **Vitali Vassiljev.** Influence of nitric oxide syntase inhibitors on the effects of ethanol after acute and chronic ethanol administration and withdrawal. Tartu, 2004.

100. **Aune Rehema.** Assessment of nonhaem ferrous iron and glutathione redox ratio as markers of pathogeneticity of oxidative stress in different clinical groups. Tartu, 2004.
101. **Evelin Seppet.** Interaction of mitochondria and ATPases in oxidative muscle cells in normal and pathological conditions. Tartu, 2004.
102. **Eduard Maron.** Serotonin function in panic disorder: from clinical experiments to brain imaging and genetics. Tartu, 2004.
103. **Marje Oona.** *Helicobacter pylori* infection in children: epidemiological and therapeutic aspects. Tartu, 2004.
104. **Kersti Kokk.** Regulation of active and passive molecular transport in the testis. Tartu, 2005.
105. **Vladimir Järv.** Cross-sectional imaging for pretreatment evaluation and follow-up of pelvic malignant tumours. Tartu, 2005.
106. **Andre Õun.** Epidemiology of adult epilepsy in Tartu, Estonia. Incidence, prevalence and medical treatment. Tartu, 2005.
107. **Piibe Muda.** Homocysteine and hypertension: associations between homocysteine and essential hypertension in treated and untreated hypertensive patients with and without coronary artery disease. Tartu, 2005.
108. **Küllli Kingo.** The interleukin-10 family cytokines gene polymorphisms in plaque psoriasis. Tartu, 2005.
109. **Mati Merila.** Anatomy and clinical relevance of the glenohumeral joint capsule and ligaments. Tartu, 2005.
110. **Epp Songisepp.** Evaluation of technological and functional properties of the new probiotic *Lactobacillus fermentum* ME-3. Tartu, 2005.
111. **Tiia Ainla.** Acute myocardial infarction in Estonia: clinical characteristics, management and outcome. Tartu, 2005.
112. **Andres Sell.** Determining the minimum local anaesthetic requirements for hip replacement surgery under spinal anaesthesia – a study employing a spinal catheter. Tartu, 2005.
113. **Tiia Tamme.** Epidemiology of odontogenic tumours in Estonia. Pathogenesis and clinical behaviour of ameloblastoma. Tartu, 2005.
114. **Triine Annus.** Allergy in Estonian schoolchildren: time trends and characteristics. Tartu, 2005.
115. **Tiia Voor.** Microorganisms in infancy and development of allergy: comparison of Estonian and Swedish children. Tartu, 2005.
116. **Priit Kasenõmm.** Indicators for tonsillectomy in adults with recurrent tonsillitis – clinical, microbiological and pathomorphological investigations. Tartu, 2005.
117. **Eva Zusinaite.** Hepatitis C virus: genotype identification and interactions between viral proteases. Tartu, 2005.
118. **Piret Köll.** Oral lactoflora in chronic periodontitis and periodontal health. Tartu, 2006.
119. **Tiina Stelmach.** Epidemiology of cerebral palsy and unfavourable neurodevelopmental outcome in child population of Tartu city and county, Estonia Prevalence, clinical features and risk factors. Tartu, 2006.

120. **Katrin Pudersell.** Tropane alkaloid production and riboflavine excretion in the field and tissue cultures of henbane (*Hyoscyamus niger* L.). Tartu, 2006.
121. **Küllli Jaako.** Studies on the role of neurogenesis in brain plasticity. Tartu, 2006.
122. **Aare Märtsen.** Lower limb lengthening: experimental studies of bone regeneration and long-term clinical results. Tartu, 2006.
123. **Heli Tähepõld.** Patient consultation in family medicine. Tartu, 2006.
124. **Stanislav Liskmann.** Peri-implant disease: pathogenesis, diagnosis and treatment in view of both inflammation and oxidative stress profiling. Tartu, 2006.
125. **Ruth Rudissaar.** Neuropharmacology of atypical antipsychotics and an animal model of psychosis. Tartu, 2006.
126. **Helena Andreson.** Diversity of *Helicobacter pylori* genotypes in Estonian patients with chronic inflammatory gastric diseases. Tartu, 2006.
127. **Katrin Pruus.** Mechanism of action of antidepressants: aspects of serotonergic system and its interaction with glutamate. Tartu, 2006.
128. **Priit Põder.** Clinical and experimental investigation: relationship of ischaemia/reperfusion injury with oxidative stress in abdominal aortic aneurysm repair and in extracranial brain artery endarterectomy and possibilities of protection against ischaemia using a glutathione analogue in a rat model of global brain ischaemia. Tartu, 2006.
129. **Marika Tammaru.** Patient-reported outcome measurement in rheumatoid arthritis. Tartu, 2006.
130. **Tiia Reimand.** Down syndrome in Estonia. Tartu, 2006.
131. **Diva Eensoo.** Risk-taking in traffic and Markers of Risk-Taking Behaviour in Schoolchildren and Car Drivers. Tartu, 2007.
132. **Riina Vibo.** The third stroke registry in Tartu, Estonia from 2001 to 2003: incidence, case-fatality, risk factors and long-term outcome. Tartu, 2007.
133. **Chris Pruunsild.** Juvenile idiopathic arthritis in children in Estonia. Tartu, 2007.
134. **Eve Õiglane-Šlik.** Angelman and Prader-Willi syndromes in Estonia. Tartu, 2007.
135. **Kadri Haller.** Antibodies to follicle stimulating hormone. Significance in female infertility. Tartu, 2007.
136. **Pille Ööpik.** Management of depression in family medicine. Tartu, 2007.
137. **Jaak Kals.** Endothelial function and arterial stiffness in patients with atherosclerosis and in healthy subjects. Tartu, 2007.
138. **Priit Kampus.** Impact of inflammation, oxidative stress and age on arterial stiffness and carotid artery intima-media thickness. Tartu, 2007.
139. **Margus Punab.** Male fertility and its risk factors in Estonia. Tartu, 2007.
140. **Alar Toom.** Heterotopic ossification after total hip arthroplasty: clinical and pathogenetic investigation. Tartu, 2007.

141. **Lea Pehme.** Epidemiology of tuberculosis in Estonia 1991–2003 with special regard to extrapulmonary tuberculosis and delay in diagnosis of pulmonary tuberculosis. Tartu, 2007.
142. **Juri Karjagin.** The pharmacokinetics of metronidazole and meropenem in septic shock. Tartu, 2007.
143. **Inga Talvik.** Inflicted traumatic brain injury shaken baby syndrome in Estonia – epidemiology and outcome. Tartu, 2007.
144. **Tarvo Rajasalu.** Autoimmune diabetes: an immunological study of type 1 diabetes in humans and in a model of experimental diabetes (in RIP-B7.1 mice). Tartu, 2007.
145. **Inga Karu.** Ischaemia-reperfusion injury of the heart during coronary surgery: a clinical study investigating the effect of hyperoxia. Tartu, 2007.
146. **Peeter Padrik.** Renal cell carcinoma: Changes in natural history and treatment of metastatic disease. Tartu, 2007.
147. **Neve Vendt.** Iron deficiency and iron deficiency anaemia in infants aged 9 to 12 months in Estonia. Tartu, 2008.
148. **Lenne-Triin Heidmets.** The effects of neurotoxins on brain plasticity: focus on neural Cell Adhesion Molecule. Tartu, 2008.
149. **Paul Korrovits.** Asymptomatic inflammatory prostatitis: prevalence, etiological factors, diagnostic tools. Tartu, 2008.
150. **Annika Reintam.** Gastrointestinal failure in intensive care patients. Tartu, 2008.
151. **Kristiina Roots.** Cationic regulation of Na-pump in the normal, Alzheimer's and CCK<sub>2</sub> receptor-deficient brain. Tartu, 2008.
152. **Helen Puusepp.** The genetic causes of mental retardation in Estonia: fragile X syndrome and creatine transporter defect. Tartu, 2009.
153. **Kristiina Rull.** Human chorionic gonadotropin beta genes and recurrent miscarriage: expression and variation study. Tartu, 2009.
154. **Margus Eimre.** Organization of energy transfer and feedback regulation in oxidative muscle cells. Tartu, 2009.
155. **Maire Link.** Transcription factors FoxP3 and AIRE: autoantibody associations. Tartu, 2009.
156. **Kai Haldre.** Sexual health and behaviour of young women in Estonia. Tartu, 2009.
157. **Kaur Liivak.** Classical form of congenital adrenal hyperplasia due to 21-hydroxylase deficiency in Estonia: incidence, genotype and phenotype with special attention to short-term growth and 24-hour blood pressure. Tartu, 2009.
158. **Kersti Ehrlich.** Antioxidative glutathione analogues (UPF peptides) – molecular design, structure-activity relationships and testing the protective properties. Tartu, 2009.
159. **Anneli Rätsep.** Type 2 diabetes care in family medicine. Tartu, 2009.
160. **Silver Türk.** Etiopathogenetic aspects of chronic prostatitis: role of mycoplasmas, coryneform bacteria and oxidative stress. Tartu, 2009.

161. **Kaire Heilman.** Risk markers for cardiovascular disease and low bone mineral density in children with type 1 diabetes. Tartu, 2009.
162. **Kristi Rüütel.** HIV-epidemic in Estonia: injecting drug use and quality of life of people living with HIV. Tartu, 2009.
163. **Triin Eller.** Immune markers in major depression and in antidepressive treatment. Tartu, 2009.
164. **Siim Suutre.** The role of TGF- $\beta$  isoforms and osteoprogenitor cells in the pathogenesis of heterotopic ossification. An experimental and clinical study of hip arthroplasty. Tartu, 2010.
165. **Kai Kliiman.** Highly drug-resistant tuberculosis in Estonia: Risk factors and predictors of poor treatment outcome. Tartu, 2010.
166. **Inga Villa.** Cardiovascular health-related nutrition, physical activity and fitness in Estonia. Tartu, 2010.
167. **Tõnis Org.** Molecular function of the first PHD finger domain of Auto-immune Regulator protein. Tartu, 2010.
168. **Tuuli Metsvaht.** Optimal antibacterial therapy of neonates at risk of early onset sepsis. Tartu, 2010.
169. **Jaanus Kahu.** Kidney transplantation: Studies on donor risk factors and mycophenolate mofetil. Tartu, 2010.
170. **Koit Reimand.** Autoimmunity in reproductive failure: A study on associated autoantibodies and autoantigens. Tartu, 2010.
171. **Mart Kull.** Impact of vitamin D and hypolactasia on bone mineral density: a population based study in Estonia. Tartu, 2010.
172. **Rael Laugesaar.** Stroke in children – epidemiology and risk factors. Tartu, 2010.
173. **Mark Braschinsky.** Epidemiology and quality of life issues of hereditary spastic paraplegia in Estonia and implementation of genetic analysis in everyday neurologic practice. Tartu, 2010.
174. **Kadri Suija.** Major depression in family medicine: associated factors, recurrence and possible intervention. Tartu, 2010.
175. **Jarno Habicht.** Health care utilisation in Estonia: socioeconomic determinants and financial burden of out-of-pocket payments. Tartu, 2010.
176. **Kristi Abram.** The prevalence and risk factors of rosacea. Subjective disease perception of rosacea patients. Tartu, 2010.
177. **Malle Kuum.** Mitochondrial and endoplasmic reticulum cation fluxes: Novel roles in cellular physiology. Tartu, 2010.
178. **Rita Teek.** The genetic causes of early onset hearing loss in Estonian children. Tartu, 2010.
179. **Daisy Volmer.** The development of community pharmacy services in Estonia – public and professional perceptions 1993–2006. Tartu, 2010.
180. **Jelena Lissitsina.** Cytogenetic causes in male infertility. Tartu, 2011.
181. **Delia Lepik.** Comparison of gunshot injuries caused from Tokarev, Makarov and Glock 19 pistols at different firing distances. Tartu, 2011.
182. **Ene-Renate Pähkla.** Factors related to the efficiency of treatment of advanced periodontitis. Tartu, 2011.

183. **Maarja Krass.** L-Arginine pathways and antidepressant action. Tartu, 2011.
184. **Taavi Lai.** Population health measures to support evidence-based health policy in Estonia. Tartu, 2011.
185. **Tiit Salum.** Similarity and difference of temperature-dependence of the brain sodium pump in normal, different neuropathological, and aberrant conditions and its possible reasons. Tartu, 2011.
186. **Tõnu Vooder.** Molecular differences and similarities between histological subtypes of non-small cell lung cancer. Tartu, 2011.
187. **Jelena Štšepetova.** The characterisation of intestinal lactic acid bacteria using bacteriological, biochemical and molecular approaches. Tartu, 2011.
188. **Radko Avi.** Natural polymorphisms and transmitted drug resistance in Estonian HIV-1 CRF06\_cpx and its recombinant viruses. Tartu, 2011, 116 p.
189. **Edward Laane.** Multiparameter flow cytometry in haematological malignancies. Tartu, 2011, 152 p.
190. **Triin Jagomägi.** A study of the genetic etiology of nonsyndromic cleft lip and palate. Tartu, 2011, 158 p.
191. **Ivo Laidmäe.** Fibrin glue of fish (*Salmo salar*) origin: immunological study and development of new pharmaceutical preparation. Tartu, 2012, 150 p.
192. **Ülle Parm.** Early mucosal colonisation and its role in prediction of invasive infection in neonates at risk of early onset sepsis. Tartu, 2012, 168 p.
193. **Kaupo Teesalu.** Autoantibodies against desmin and transglutaminase 2 in celiac disease: diagnostic and functional significance. Tartu, 2012, 142 p.
194. **Maksim Zagura.** Biochemical, functional and structural profiling of arterial damage in atherosclerosis. Tartu, 2012, 162 p.
195. **Vivian Kont.** Autoimmune regulator: characterization of thymic gene regulation and promoter methylation. Tartu, 2012, 134 p.
196. **Pirje Hütt.** Functional properties, persistence, safety and efficacy of potential probiotic lactobacilli. Tartu, 2012, 246 p.
197. **Innar Tõru.** Serotonergic modulation of CCK-4- induced panic. Tartu, 2012, 132 p.
198. **Sigrid Vorobjov.** Drug use, related risk behaviour and harm reduction interventions utilization among injecting drug users in Estonia: implications for drug policy. Tartu, 2012, 120 p.
199. **Martin Serg.** Therapeutic aspects of central haemodynamics, arterial stiffness and oxidative stress in hypertension. Tartu, 2012, 156 p.
200. **Jaanika Kumm.** Molecular markers of articular tissues in early knee osteoarthritis: a population-based longitudinal study in middle-aged subjects. Tartu, 2012, 159 p.
201. **Kertu Rünkorg.** Functional changes of dopamine, endopioid and endocannabinoid systems in CCK2 receptor deficient mice. Tartu, 2012, 125 p.
202. **Mai Blöndal.** Changes in the baseline characteristics, management and outcomes of acute myocardial infarction in Estonia. Tartu, 2012, 127 p.

203. **Jana Lass.** Epidemiological and clinical aspects of medicines use in children in Estonia. Tartu, 2012, 170 p.
204. **Kai Truusalu.** Probiotic lactobacilli in experimental persistent *Salmonella* infection. Tartu, 2013, 139 p.
205. **Oksana Jagur.** Temporomandibular joint diagnostic imaging in relation to pain and bone characteristics. Long-term results of arthroscopic treatment. Tartu, 2013, 126 p.
206. **Katrin Sikk.** Manganese-ephedrone intoxication – pathogenesis of neurological damage and clinical symptomatology. Tartu, 2013, 125 p.
207. **Kai Blöndal.** Tuberculosis in Estonia with special emphasis on drug-resistant tuberculosis: Notification rate, disease recurrence and mortality. Tartu, 2013, 151 p.
208. **Marju Puurand.** Oxidative phosphorylation in different diseases of gastric mucosa. Tartu, 2013, 123 p.
209. **Aili Tagoma.** Immune activation in female infertility: Significance of autoantibodies and inflammatory mediators. Tartu, 2013, 135 p.
210. **Liis Sabre.** Epidemiology of traumatic spinal cord injury in Estonia. Brain activation in the acute phase of traumatic spinal cord injury. Tartu, 2013, 135 p.
211. **Merit Lamp.** Genetic susceptibility factors in endometriosis. Tartu, 2013, 125 p.
212. **Erik Salum.** Beneficial effects of vitamin D and angiotensin II receptor blocker on arterial damage. Tartu, 2013, 167 p.
213. **Maire Karelson.** Vitiligo: clinical aspects, quality of life and the role of melanocortin system in pathogenesis. Tartu, 2013, 153 p.
214. **Kuldar Kaljurand.** Prevalence of exfoliation syndrome in Estonia and its clinical significance. Tartu, 2013, 113 p.
215. **Raido Paasma.** Clinical study of methanol poisoning: handling large outbreaks, treatment with antidotes, and long-term outcomes. Tartu, 2013, 96 p.
216. **Anne Kleinberg.** Major depression in Estonia: prevalence, associated factors, and use of health services. Tartu, 2013, 129 p.
217. **Triin Eglit.** Obesity, impaired glucose regulation, metabolic syndrome and their associations with high-molecular-weight adiponectin levels. Tartu, 2014, 115 p.
218. **Kristo Ausmees.** Reproductive function in middle-aged males: Associations with prostate, lifestyle and couple infertility status. Tartu, 2014, 125 p.
219. **Kristi Huik.** The influence of host genetic factors on the susceptibility to HIV and HCV infections among intravenous drug users. Tartu, 2014, 144 p.
220. **Liina Tserel.** Epigenetic profiles of monocytes, monocyte-derived macrophages and dendritic cells. Tartu, 2014, 143 p.
221. **Irina Kerna.** The contribution of *ADAM12* and *CILP* genes to the development of knee osteoarthritis. Tartu, 2014, 152 p.

222. **Ingrid Liiv.** Autoimmune regulator protein interaction with DNA-dependent protein kinase and its role in apoptosis. Tartu, 2014, 143 p.
223. **Liivi Maddison.** Tissue perfusion and metabolism during intra-abdominal hypertension. Tartu, 2014, 103 p.
224. **Krista Ress.** Childhood coeliac disease in Estonia, prevalence in atopic dermatitis and immunological characterisation of coexistence. Tartu, 2014, 124 p.
225. **Kai Muru.** Prenatal screening strategies, long-term outcome of children with marked changes in maternal screening tests and the most common syndromic heart anomalies in Estonia. Tartu, 2014, 189 p.
226. **Kaja Rahu.** Morbidity and mortality among Baltic Chernobyl cleanup workers: a register-based cohort study. Tartu, 2014, 155 p.
227. **Klari Noormets.** The development of diabetes mellitus, fertility and energy metabolism disturbances in a Wfs1-deficient mouse model of Wolfram syndrome. Tartu, 2014, 132 p.
228. **Liis Toome.** Very low gestational age infants in Estonia. Tartu, 2014, 183 p.
229. **Ceith Nikkolo.** Impact of different mesh parameters on chronic pain and foreign body feeling after open inguinal hernia repair. Tartu, 2014, 132 p.
230. **Vadim Brjalin.** Chronic hepatitis C: predictors of treatment response in Estonian patients. Tartu, 2014, 122 p.
231. **Vahur Metsna.** Anterior knee pain in patients following total knee arthroplasty: the prevalence, correlation with patellar cartilage impairment and aspects of patellofemoral congruence. Tartu, 2014, 130 p.
232. **Marju Kase.** Glioblastoma multiforme: possibilities to improve treatment efficacy. Tartu, 2015, 137 p.
233. **Riina Runnel.** Oral health among elementary school children and the effects of polyol candies on the prevention of dental caries. Tartu, 2015, 112 p.
234. **Made Laanpere.** Factors influencing women's sexual health and reproductive choices in Estonia. Tartu, 2015, 176 p.
235. **Andres Lust.** Water mediated solid state transformations of a polymorphic drug – effect on pharmaceutical product performance. Tartu, 2015, 134 p.
236. **Anna Klugman.** Functionality related characterization of pretreated wood lignin, cellulose and polyvinylpyrrolidone for pharmaceutical applications. Tartu, 2015, 156 p.
237. **Triin Laisk-Podar.** Genetic variation as a modulator of susceptibility to female infertility and a source for potential biomarkers. Tartu, 2015, 155 p.
238. **Mailis Tõnisson.** Clinical picture and biochemical changes in blood in children with acute alcohol intoxication. Tartu, 2015, 100 p.
239. **Kadri Tamme.** High volume haemodiafiltration in treatment of severe sepsis – impact on pharmacokinetics of antibiotics and inflammatory response. Tartu, 2015, 133 p.

240. **Kai Part.** Sexual health of young people in Estonia in a social context: the role of school-based sexuality education and youth-friendly counseling services. Tartu, 2015, 203 p.
241. **Urve Paaver.** New perspectives for the amorphization and physical stabilization of poorly water-soluble drugs and understanding their dissolution behavior. Tartu, 2015, 139 p.
242. **Aleksandr Peet.** Intrauterine and postnatal growth in children with HLA-conferred susceptibility to type 1 diabetes. Tartu. 2015, 146 p.
243. **Piret Mitt.** Healthcare-associated infections in Estonia – epidemiology and surveillance of bloodstream and surgical site infections. Tartu, 2015, 145 p.
244. **Merli Saare.** Molecular Profiling of Endometriotic Lesions and Endometriosis of Endometriosis Patients. Tartu, 2016, 129 p.
245. **Kaja-Triin Laisaar.** People living with HIV in Estonia: Engagement in medical care and methods of increasing adherence to antiretroviral therapy and safe sexual behavior. Tartu, 2016, 132 p.
246. **Eero Merilind.** Primary health care performance: impact of payment and practice-based characteristics. Tartu, 2016, 120 p.
247. **Jaanika Kärner.** Cytokine-specific autoantibodies in AIRE deficiency. Tartu, 2016, 182 p.
248. **Kaido Paapstel.** Metabolomic profile of arterial stiffness and early biomarkers of renal damage in atherosclerosis. Tartu, 2016, 173 p.
249. **Liidia Kiisk.** Long-term nutritional study: anthropometrical and clinico-laboratory assessments in renal replacement therapy patients after intensive nutritional counselling. Tartu, 2016, 207 p.
250. **Georgi Nellis.** The use of excipients in medicines administered to neonates in Europe. Tartu, 2017, 159 p.
251. **Aleksei Rakitin.** Metabolic effects of acute and chronic treatment with valproic acid in people with epilepsy. Tartu, 2017, 125 p.
252. **Eveli Kallas.** The influence of immunological markers to susceptibility to HIV, HBV, and HCV infections among persons who inject drugs. Tartu, 2017, 138 p.
253. **Tiina Freimann.** Musculoskeletal pain among nurses: prevalence, risk factors, and intervention. Tartu, 2017, 125 p.
254. **Evelyn Aaviksoo.** Sickness absence in Estonia: determinants and influence of the sick-pay cut reform. Tartu, 2017, 121 p.
255. **Kalev Nõupuu.** Autosomal-recessive Stargardt disease: phenotypic heterogeneity and genotype-phenotype associations. Tartu, 2017, 131 p.
256. **Ho Duy Binh.** Osteogenesis imperfecta in Vietnam. Tartu, 2017, 125 p.
257. **Uku Haljasorg.** Transcriptional mechanisms in thymic central tolerance. Tartu, 2017, 147 p.
258. **Živile Riispere.** IgA Nephropathy study according to the Oxford Classification: IgA Nephropathy clinical-morphological correlations, disease progression and the effect of renoprotective therapy. Tartu, 2017, 129 p.

259. **Hiie Soeorg**. Coagulase-negative staphylococci in gut of preterm neonates and in breast milk of their mothers. Tartu, 2017, 216 p.
260. **Anne-Mari Anton Willmore**. Silver nanoparticles for cancer research. Tartu, 2017, 132 p.
261. **Ott Laius**. Utilization of osteoporosis medicines, medication adherence and the trend in osteoporosis related hip fractures in Estonia. Tartu, 2017, 134 p.
262. **Alar Aab**. Insights into molecular mechanisms of asthma and atopic dermatitis. Tartu, 2017, 164 p.
263. **Sander Pajusalu**. Genome-wide diagnostics of Mendelian disorders: from chromosomal microarrays to next-generation sequencing. Tartu, 2017, 146 p.
264. **Mikk Jürisson**. Health and economic impact of hip fracture in Estonia. Tartu, 2017, 164 p.
265. **Kaspar Tootsi**. Cardiovascular and metabolomic profiling of osteoarthritis. Tartu, 2017, 150 p.
266. **Mario Saare**. The influence of AIRE on gene expression – studies of transcriptional regulatory mechanisms in cell culture systems. Tartu, 2017, 172 p.
267. **Piia Jõgi**. Epidemiological and clinical characteristics of pertussis in Estonia. Tartu, 2018, 168 p.
268. **Elle Põldoja**. Structure and blood supply of the superior part of the shoulder joint capsule. Tartu, 2018, 116 p.
269. **Minh Son Nguyen**. Oral health status and prevalence of temporomandibular disorders in 65–74-year-olds in Vietnam. Tartu, 2018, 182 p.
270. **Kristian Semjonov**. Development of pharmaceutical quench-cooled molten and melt-electrospun solid dispersions for poorly water-soluble indomethacin. Tartu, 2018, 125 p.
271. **Janne Tiigimäe-Saar**. Botulinum neurotoxin type A treatment for sialorrhea in central nervous system diseases. Tartu, 2018, 109 p.
272. **Veiko Vengerfeldt**. Apical periodontitis: prevalence and etiopathogenetic aspects. Tartu, 2018, 150 p.
273. **Rudolf Bichele**. TNF superfamily and AIRE at the crossroads of thymic differentiation and host protection against *Candida albicans* infection. Tartu, 2018, 153 p.
274. **Olga Tšuiiko**. Unravelling Chromosomal Instability in Mammalian Pre-implantation Embryos Using Single-Cell Genomics. Tartu, 2018, 169 p.
275. **Kärt Kriisa**. Profile of acylcarnitines, inflammation and oxidative stress in first-episode psychosis before and after antipsychotic treatment. Tartu, 2018, 145 p.
276. **Xuan Dung Ho**. Characterization of the genomic profile of osteosarcoma. Tartu, 2018, 144 p.
277. **Karit Reinson**. New Diagnostic Methods for Early Detection of Inborn Errors of Metabolism in Estonia. Tartu, 2018, 201 p.

278. **Mari-Anne Vals.** Congenital N-glycosylation Disorders in Estonia. Tartu, 2019, 148 p.
279. **Liis Kadastik-Eerme.** Parkinson's disease in Estonia: epidemiology, quality of life, clinical characteristics and pharmacotherapy. Tartu, 2019, 202 p.
280. **Hedi Hunt.** Precision targeting of intraperitoneal tumors with peptide-guided nanocarriers. Tartu, 2019, 179 p.
281. **Rando Porosk.** The role of oxidative stress in Wolfram syndrome 1 and hypothermia. Tartu, 2019, 123 p.
282. **Ene-Ly Jõgeda.** The influence of coinfections and host genetic factor on the susceptibility to HIV infection among people who inject drugs. Tartu, 2019, 126 p.
283. **Kristel Ehala-Aleksejev.** The associations between body composition, obesity and obesity-related health and lifestyle conditions with male reproductive function. Tartu, 2019, 138 p.
284. **Aigar Ottas.** The metabolomic profiling of psoriasis, atopic dermatitis and atherosclerosis. Tartu, 2019, 136 p.
285. **Elmira Gurbanova.** Specific characteristics of tuberculosis in low default, but high multidrug-resistance prison setting. Tartu, 2019, 129 p.
286. **Van Thai Nguyeni.** The first study of the treatment outcomes of patients with cleft lip and palate in Central Vietnam. Tartu, 2019, 144 p.
287. **Maria Yakoreva.** Imprinting Disorders in Estonia. Tartu, 2019, 187 p.
288. **Kadri Rekker.** The putative role of microRNAs in endometriosis pathogenesis and potential in diagnostics. Tartu, 2019, 140 p.
289. **Ülle Võhma.** Association between personality traits, clinical characteristics and pharmacological treatment response in panic disorder. Tartu, 2019, 121 p.
290. **Aet Saar.** Acute myocardial infarction in Estonia 2001–2014: towards risk-based prevention and management. Tartu, 2019, 124 p.
291. **Toomas Toomsoo.** Transcranial brain sonography in the Estonian cohort of Parkinson's disease. Tartu, 2019, 114 p.
292. **Lidiia Zhytnik.** Inter- and intrafamilial diversity based on genotype and phenotype correlations of Osteogenesis Imperfecta. Tartu, 2019, 224 p.
293. **Pilleriin Soodla.** Newly HIV-infected people in Estonia: estimation of incidence and transmitted drug resistance. Tartu, 2019, 194 p.
294. **Kristiina Ojamaa.** Epidemiology of gynecological cancer in Estonia. Tartu, 2020, 133 p.
295. **Marianne Saard.** Modern Cognitive and Social Intervention Techniques in Paediatric Neurorehabilitation for Children with Acquired Brain Injury. Tartu, 2020, 168 p.
296. **Julia Maslovskaja.** The importance of DNA binding and DNA breaks for AIRE-mediated transcriptional activation. Tartu, 2020, 162 p.
297. **Natalia Lobanovskaya.** The role of PSA-NCAM in the survival of retinal ganglion cells. Tartu, 2020, 105 p.

298. **Madis Rahu.** Structure and blood supply of the postero-superior part of the shoulder joint capsule with implementation of surgical treatment after anterior traumatic dislocation. Tartu, 2020, 104 p.
299. **Helen Zirnask.** Luteinizing hormone (LH) receptor expression in the penis and its possible role in pathogenesis of erectile disturbances. Tartu, 2020, 87 p.
300. **Kadri Toome.** Homing peptides for targeting of brain diseases. Tartu, 2020, 152 p.
301. **Maarja Hallik.** Pharmacokinetics and pharmacodynamics of inotropic drugs in neonates. Tartu, 2020, 172 p.
302. **Raili Müller.** Cardiometabolic risk profile and body composition in early rheumatoid arthritis. Tartu, 2020, 133 p.
303. **Sergo Kasvandik.** The role of proteomic changes in endometrial cells – from the perspective of fertility and endometriosis. Tartu, 2020, 191 p.
304. **Epp Kaleviste.** Genetic variants revealing the role of STAT1/STAT3 signaling cytokines in immune protection and pathology. Tartu, 2020, 189 p.
305. **Sten Saar.** Epidemiology of severe injuries in Estonia. Tartu, 2020, 104 p.
306. **Kati Braschinsky.** Epidemiology of primary headaches in Estonia and applicability of web-based solutions in headache epidemiology research. Tartu, 2020, 129 p.
307. **Helen Vaher.** MicroRNAs in the regulation of keratinocyte responses in *psoriasis vulgaris* and atopic dermatitis. Tartu, 2020, 242 p.
308. **Liisi Raam.** Molecular Alterations in the Pathogenesis of Two Chronic Dermatoses – Vitiligo and Psoriasis. Tartu, 2020, 164 p.
309. **Artur Vetkas.** Long-term quality of life, emotional health, and associated factors in patients after aneurysmal subarachnoid haemorrhage. Tartu, 2020, 127 p.
310. **Teele Kasepalu.** Effects of remote ischaemic preconditioning on organ damage and acylcarnitines' metabolism in vascular surgery. Tartu, 2020, 130 p.
311. **Prakash Lingasamy.** Development of multitargeted tumor penetrating peptides. Tartu, 2020, 246 p.
312. **Lille Kurvits.** Parkinson's disease as a multisystem disorder: whole transcriptome study in Parkinson's disease patients' skin and blood. Tartu, 2021, 142 p.
313. **Mariliis Pöld.** Smoking, attitudes towards smoking behaviour, and nicotine dependence among physicians in Estonia: cross-sectional surveys 1982–2014. Tartu, 2021, 172 p.
314. **Triin Kikas.** Single nucleotide variants affecting placental gene expression and pregnancy outcome. Tartu, 2021, 160 p.
315. **Hedda Lippus-Metsaots.** Interpersonal violence in Estonia: prevalence, impact on health and health behaviour. Tartu, 2021, 172 p.

316. **Georgi Dzaparidze.** Quantification and evaluation of the diagnostic significance of adenocarcinoma-associated microenvironmental changes in the prostate using modern digital pathology solutions. Tartu, 2021, 132 p.
317. **Tuuli Sedman.** New avenues for GLP1 receptor agonists in the treatment of diabetes. Tartu, 2021, 118 p.
318. **Martin Padar.** Enteral nutrition, gastrointestinal dysfunction and intestinal biomarkers in critically ill patients. Tartu, 2021, 189 p.
319. **Siim Schneider.** Risk factors, etiology and long-term outcome in young ischemic stroke patients in Estonia. Tartu, 2021, 131 p.
320. **Konstantin Ridnõi.** Implementation and effectiveness of new prenatal diagnostic strategies in Estonia. Tartu, 2021, 191 p.
321. **Risto Vaikjärv.** Etiopathogenetic and clinical aspects of peritonsillar abscess. Tartu, 2021, 115 p.
322. **Liis Preem.** Design and characterization of antibacterial electrospun drug delivery systems for wound infections. Tartu, 2022, 220 p.
323. **Keerthie Dissanayake.** Preimplantation embryo-derived extracellular vesicles: potential as an embryo quality marker and their role during the embryo-maternal communication. Tartu, 2022, 203 p.
324. **Laura Viidik.** 3D printing in pharmaceuticals: a new avenue for fabricating therapeutic drug delivery systems. Tartu, 2022, 139 p.
325. **Kasun Godakumara.** Extracellular vesicle mediated embryo-maternal communication – A tool for evaluating functional competency of pre-implantation embryos. Tartu, 2022, 176 p.
326. **Hindrekk Teder.** Developing computational methods and workflows for targeted and whole-genome sequencing based non-invasive prenatal testing. Tartu, 2022, 138 p.
327. **Jana Tuusov.** Deaths caused by alcohol, psychotropic and other substances in Estonia: evidence based on forensic autopsies. Tartu, 2022, 157 p.
328. **Heigo Reima.** Colorectal cancer care and outcomes – evaluation and possibilities for improvement in Estonia. Tartu, 2022, 146 p.
329. **Liisa Kuhi.** A contribution of biomarker collagen type II neopeptide C2C in urine to the diagnosis and prognosis of knee osteoarthritis. Tartu, 2022, 157 p.
330. **Reeli Tamme.** Associations between pubertal hormones and physical activity levels, and subsequent bone mineral characteristics: a longitudinal study of boys aged 12–18. Tartu, 2022, 118 p.
331. **Deniss Sõritsa.** The impact of endometriosis and physical activity on female reproduction. Tartu, 2022, 152 p.
332. **Mohammad Mehedi Hasan.** Characterization of follicular fluid-derived extracellular vesicles and their contribution to periconception environment. Tartu, 2022, 194 p.
333. **Priya Kulkarni.** Osteoarthritis pathogenesis: an immunological passage through synovium-synovial fluid axis. Tartu, 2022, 268 p.

334. **Nigul Ilves.** Brain plasticity and network reorganization in children with perinatal stroke: a functional magnetic resonance imaging study. Tartu, 2022, 169 p.
335. **Marko Murruste.** Short- and long-term outcomes of surgical management of chronic pancreatitis. Tartu, 2022, 180 p.
336. **Marilin Ivask.** Transcriptomic and metabolic changes in the WFS1-deficient mouse model. Tartu, 2022, 158 p.
337. **Jüri Lieberg.** Results of surgical treatment and role of biomarkers in pathogenesis and risk prediction in patients with abdominal aortic aneurysm and peripheral artery disease. Tartu, 2022, 160 p.
338. **Sanna Puusepp.** Comparison of molecular genetics and morphological findings of childhood-onset neuromuscular disorders. Tartu, 2022, 216 p.
339. **Khan Nguyen Viet.** Chemical composition and bioactivity of extracts and constituents isolated from the medicinal plants in Vietnam and their nanotechnology-based delivery systems. Tartu, 2023, 172 p.
340. **Getnet Balcha Midekessa.** Towards understanding the colloidal stability and detection of Extracellular Vesicles. Tartu, 2023, 172 p.
341. **Kristiina Sepp.** Competency-based and person-centred community pharmacy practice – development and implementation in Estonia. Tartu, 2023, 242 p.
342. **Linda Sõber.** Impact of thyroid disease and surgery on patient's quality of voice and swallowing. Tartu, 2023, 114 p.
343. **Anni Lepland.** Precision targeting of tumour-associated macrophages in triple negative breast cancer. Tartu, 2023, 160 p.
344. **Sirje Sammul.** Prevalence and risk factors of arterial hypertension and cardiovascular mortality: 13-year longitudinal study among 35- and 55-year-old adults in Estonia and Sweden. Tartu, 2023, 158 p.
345. **Maarjaliis Paavo.** Short-Wavelength and Near-Infrared Autofluorescence Imaging in Recessive Stargardt Disease, Choroideremia, *PROM1*-Macular Dystrophy and Ocular Albinism. Tartu, 2023, 202 p.
346. **Kaspar Ratnik.** development of predictive multimarker test for pre-eclampsia in early and late pregnancy. Tartu, 2023, 134 p.
347. **Kärt Simre.** Development of coeliac disease in two populations with different environmental backgrounds. Tartu, 2023, 161 p.
348. **Qurat Ul Ain Reshi.** Characterization of the maternal reproductive tract and spermatozoa communication during periconception period via extracellular vesicles. Tartu, 2023, 182 p.
349. **Stanislav Tjagur.** *Mycoplasma genitalium* and other sexually transmitted infections causing urethritis – their prevalence, impact on male fertility parameters and prostate health. Tartu, 2023, 225 p.
350. **Lagle Lehes.** The first study of voice and resonance related treatment outcomes of Estonian cleft palate children. Tartu, 2023, 126 p.

351. **Liis Ilves.** Metabolomic profiling of chronic inflammatory skin diseases. Tartu, 2023, 146 p.
352. **Marina Šunina.** Flow cytometric analysis of T and B cell properties in healthy donors and subjects with vitiligo. Tartu, 2023, 164 p.